Development of a deep-water carbonate ion concentration proxy based on preservation of planktonic foraminifera shells quantified by X-ray CT scanning

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Abstract

The quantitative and objective characterization of dissolution intensity in fossil planktonic foraminiferal shells could be used to reconstruct past changes in bottom water carbonate ion concentration. Among proxies measuring the degree of dissolution of planktonic foraminiferal shells, X-ray micro-Computed Tomography (CT) based characterization of apparent shell density appears to have good potential to facilitate quantitative reconstruction of carbonate chemistry. However, unlike the wellestablished benthic foraminiferal B/Ca ratio-based proxy, only a regional calibration of the CT-based proxy exists based on a limited number of data points covering mainly low-saturation state waters. Here we determined by CT-based proxy the shell dissolution intensity of planktonic foraminifera Globigerina bulloides, Globorotalia inflata, Globigerinoides ruber, and Trilobatus sacculifer from a collection of core top samples in the Southern Atlantic covering higher saturation states, and assessed the characteristics and reliability of CT-based proxy. We observed that the CT-based proxy is generally controlled by deep-water Δ [°O32–] λ uxe the B/ α προξψ, β ut itc εφφεςtic ρανγε οφ Δ [°O32–] ic β ετωεεν –20 to 10 µmolkg–1. In this range, the CT-based proxy appears directly and strongly related to deep-water Δ [°O32–], ωρερεας the B/ α σφ β εντηις φοραμινιφερα αππεαφε to β ε αφφεετεδ β ψ πορεωατερ σατυρατιον is capβονατε-ριξη συβστρατες. Ον τηε στηε pl α σ β ξυτηρε ϕ σεξψ ρεχυιρες σπεειεσ-σπεειφις caλιβρατιον, β ut τηε εφφεετ οφ σπεειεσ-σπεειφις σηελλ διφφερενεε is συσεεπτιβιλιτψ το δισσολυτιον ov τηε προξψ ic σμαλλ.

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1 Development of a deep-water carbonate ion concentration proxy based on 2 preservation of planktonic foraminifera shells quantified by X-ray CT 3 scanning 4 5 S. Iwasaki^{1*}, K. Kimoto², M. Kucera¹ 6 7 ¹MARUM - Center for Marine Environmental Sciences, University of Bremen, Bremen, 8 Germany 9 ² Japan Agency for Marine-Earth Science and Technology, Research Institute for Global Change, 10 Yokosuka, Japan 11 12 13 14 Corresponding author: Shinya Iwasaki (siwasaki@marum.de) 15 16 **Key Points:** 17 Planktonic foraminiferal shell dissolution intensity was determined by CT-based proxy • 18 using core top samples in the Southern Atlantic Ocean. 19 • The characteristics and reliability of CT-based proxy were assessed by comparing with 20 21 conventional proxy. • The CT-based proxy is available for reconstructing deep seawater carbonate ion 22 concentration under the appropriate condition. 23 24 25

26 Abstract

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- reliability of CT-based proxy. We observed that the CT-based proxy is generally controlled by
- 39 deep-water $\Delta[CO_3^{2^-}]$ like the B/Ca proxy, but its effective range of $\Delta[CO_3^{2^-}]$ is between -20 to
- 40 $10 \,\mu\text{molkg}^{-1}$. In this range, the CT-based proxy appears directly and strongly related to deep-
- 41 water $\Delta[CO_3^{2-}]$, whereas the B/Ca of benthic foraminifera appears to be affected by porewater
- 42 saturation in carbonate-rich substrates. On the other hand, the CT-based proxy is affected by
- 43 supralysoclinal dissolution in areas with high productivity. Like the B/Ca proxy, the CT-based
- 44 proxy requires species-specific calibration, but the effect of species-specific shell difference in
- 45 susceptibility to dissolution on the proxy is small.
- 46

47 **1 Introduction**

48 The atmospheric CO_2 concentration has fluctuated by ~80 ppm between glacial and interglacial periods, implying a large and rapid exchange of carbon between the atmosphere and 49 the ocean on those time scales [Barnola et al., 1987; Petit et al., 1999]. This is because the 50 oceanic carbon pool is about 50 times larger than that of the atmosphere [Sigman and Boyle, 51 2000], and carbon storage in the deep ocean and changes in deep-water circulation can 52 substantially alter atmospheric pCO_2 . However, data concerning the amount of carbon storage in 53 the deep sea and their temporal and spatial variation, which are essential to understand the 54 glacial-interglacial pCO_2 exchange between ocean and atmosphere, is insufficient. Because the 55 deep seawater carbonate ion concentration ($[CO_3^{2-}]$) is governed primarily by the concentration 56 of dissolved inorganic carbon and alkalinity, its reconstruction can provide valuable insights into 57 the changes in the global carbon cycle. 58

The B/Ca ratio of epifaunal benthic foraminifera has been proposed as a quantitative 59 deep seawater [CO₃²⁻] proxy [e.g., Yu and Elderfield, 2007]. This proxy relies on the fact that 60 the ratio of the two major boron species in the deep seawater, $B(OH)_3$ and $B(OH)_4^-$, varies with 61 pH [Hemming et al., 1992] and these variations are reflected in the B/Ca ratio in shell calcite. 62 Indeed, a significant correlation between B/Ca ratio in the shells of Cibicidoides wuellerstorfi 63 and deep seawater $\Delta[CO_3^{2-}]$ was shown by Yu and Elderfield [2007]. The sensitivity of C. 64 wuellerstorfi B/Ca ratio to deep seawater $\Delta [CO_3^{2-}]$ was evaluated based on core-top calibration 65 at more than 200 sites, indicating an uncertainty of $\pm 5 \ \mu mol \ kg^{-1}$ in $[CO_3^{2-}]$ (Brown and 66 Elderfield, 1996, Rae et al., 2011, Raitzsch et al., 2011, Yu and Elderfield., 2007, Yu et al., 2013, 67 2014, Brown et al., 2011). Based on this calibration, subsequent studies reconstructed temporal 68 variations in deep seawater $[CO_3^{2-}]$ at several sites, where the sediment samples contain 69

70 sufficient amount of benthic foraminifera shells of the target species [e.g., Yu et al., 2016, 2020, Allen et al., 2015, 2019]. Next to the most frequently used calibration based on C. wuellerstorfi, 71 there also exist data for another epifaunal species, C. mundulus, confirming a relationship 72 between shell B/Ca and deep seawater $\Delta[CO_3^{2-}]$, but the calibration shows a different slope, 73 indicating that the incorporation of B into the shell calcite is affected by species-specific 74 processes. Therefore, the B/Ca method can only be applied where the same species occurs 75 throughout the studied interval and also where a sufficient number of shells can be recovered to 76 facilitate the chemical analysis. This prevents applications in settings where the regional 77 environmental change caused large shifts in food availability and bottom water oxygen content, 78 79 resulting in shifts in benthic communities, as well as in settings with high sedimentation rate, where the concentration of foraminifera shells is low (e.g. Kitazato et al., 2000, Gooday, 2003, 80 81 Geslin et al., 2004).

Planktonic foraminifera shells are a primary component of carbonate in the deep-sea 82 sediment (Schiebel, 2002), and the preservation of these shells is intimately associated with the 83 deep seawater Δ [CO₃^{2–}] (e.g., Berger et al., 1982). Therefore, another way to reconstruct deep 84 seawater $[CO_3^{2-}]$ is by the quantification of the degree of dissolution of planktonic foraminifera 85 shells (e.g. Berger et al., 1982). Previous studies made use of this relationship, but applied 86 different ways to quantify dissolution intensity, such as by shell fragmentation, proportion of 87 dissolution resistant species or the ratio between benthic and planktonic foraminifera (Berger et 88 al., 1982, Peterson and Prell, 1985, Kucera, 2007). These proxies all show the expected direction 89 of the relationship with deep seawater $\Delta[CO_3^{2-}]$, but are either hard to objectively quantify or are 90 affected by initial conditions (assemblage composition) (Kucera, 2007). The size-normalized 91 92 weight of sedimentary shell was proposed to quantify carbonate dissolution intensity more objectively (Lohmann, 1995, Broecker and Clark, 2001). However, the initial weight of the 93 planktonic foraminifera shell is also controlled by surface water properties, such as surface water 94 carbonate chemistry (e.g., Barker and Elderfield, 2002; Marshall et al., 2013) making 95 quantitative reconstructions of deep-water $[CO_3^{2-}]$ impossible in settings where surface 96 properties have changed as well. 97

More recent studies suggested that the problem of quantifying preservation state of 98 planktonic foraminifera shells can be circumvented by measurements of shell architecture using 99 X-ray micro-CT scanning (Johnstone et al., 2010, Iwasaki et al., 2015, Iwasaki et al., 2019). In 100 particular, the quantification of shell preservation by measuring the proportion of more strongly 101 dissolved calcite, identified by their lower CT number, provides a means to objectively 102 determine the preservation state in a way that is not dependent on the initial size or weight of the 103 shell. As long as the ratio of the different types of calcites, making up the lamellar structure of 104 the shells remains the same, the proportion of the more dissolved calcite should only be related 105 to deep-water [CO₃²⁻] (Iwasaki et al., 2015, Iwasaki et al., 2019). However, so far only a regional 106 South pacific calibration of the CT-based proxy exists (Iwasaki et al., 2022). This calibration is 107 108 based on a limited number of data points, covering mainly low saturation state waters, and it has not been directly compared with the B/Ca proxy. 109

In addition, there are factors other than deep seawater $[CO_3^{2-}]$ variation that control carbonate dissolution on the deep seafloor and which may affect the CT-based proxy differently than the B/Ca proxy. The first factor is the variation in sedimentation rate, which alters the exposure time to deep seawater and results in differences in dissolution intensity of planktonic foraminifera shells (Berger et al., 1982). The second factor is the decomposition of organic material in the sediment, which decreases ambient seawater (or porewater) $[CO_3^{2-}]$ and may cause carbonate dissolution in seafloor sediments deposited above the regional lysocline (Berger

- et al., 1970, Milliman, 1993, Hales and Emerson, 1996, Hales, 2003). Finally, it is known from earlier attempts to use the preservation of planktonic foraminifera shells as a proxy for deep-
- earlier attempts to use the preservation of planktonic foraminifera shells as a proxy for deepwater properties that significant changes in preservation are only encountered below a certain
- critical threshold value of deep seawater $[CO_3^{2-}]$, and shells deposited above this foraminiferal
- 121 lysocline all appear well preserved (Berger et al., 1982). Therefore, to develop the method using
- 122 CT-scanning of planktonic foraminiferal shells as paleo-deep seawater $[CO_3^{2-}]$ proxy, assessing
- the effect of these secondary factors on the preservation state of shells deposited in various
- sedimentary settings is necessary.
- To assess the accuracy and reliability of the CT-based proxy for deep seawater $[CO_3^{2^-}]$
- reconstruction, we have quantified planktonic foraminifera shell preservation in a collection of
- samples from the South Atlantic, covering a gradient towards higher ambient $\Delta[CO_3^{2-}]$ values,
- and representing different sedimentation rates and productivity regimes. Importantly, in all the
- analyzed samples, B/Ca of C. wuellerstorfi have been determined by (Rae et al., 2011, Raitzsch
- et al., 2013), allowing direct proxy intercomparison. We quantified the degree of shell
- dissolution in four planktonic foraminifera species (*G. bulloides*, *G. inflata*, *G. ruber*, and *T*.
- *sacculifer*) and propose a new way to quantify the proportion of more dissolved calcite, which
- 133 does not require instrument-specific calibration of the strength of the X-ray beam. We combine
- the new measurements with previously obtained CT images (Iwasaki et al., 2019, 2022) to
- determine the regional and species-specific effects and establish a new calibration formula,
- allowing reconstructions of deep seawater $\Delta[\text{CO}_3^{2^-}]$.
- 137

138 2 Materials and Methods

139 2.1. Multiple core samples and physicochemical properties

In this study, we used planktonic foraminifera shells extracted from 13 core top samples 140 collected by multiple corer between 940 and 4173 m water depth in the South Atlantic (Figure 141 1). The samples represent a subset of core tops where benthic foraminiferal (Cibicidoides 142 wuellerstorfi) B/Ca ratio, a conventional proxy of deep seawater [CO₃²⁻], has been determined 143 by previous studies (Rae et al., 2011, Raitzsch et al., 2013) (Figure 2). The subset contains all 144 samples used in the previous studies for which the material is stored in the GeoB repository in 145 Bremen and where information is available on sedimentation rate and accumulation rate of 146 organic carbon. From each core top sample, eight shells of each of four planktic foraminiferal 147 species (G. bulloides, G. inflata, G. ruber, and T. sacculifer) were collected for analysis by X-ray 148 micro-CT scanning. Among the 13 sites, shells of G. bulloides could be collected from all sites, 149 shells of G. inflata and T. sacculifer were collected from 12 sites, and shells of G. ruber were 150 151 collected from 11 sites. The all shells were collected from the 300-355 µm size fraction, and are not fragmented or containing fractures and holes, or peeling of the surface. Estimates of 152 sedimentation rate (cm kyr⁻¹) at the studied sites were taken from age models of sediment cores 153 sampled at the same site (Wefer et al., 1990, Schulz et al., 1992, 1996; Bleil et al. 2001), and 154 used to calculate accumulation rate of organic carbon ($g \text{ cm}^{-2} \text{ kyr}^{-1}$) making use of total organic 155 carbon content (%TOC) and dry bulk density (g cm-3) data for each site (Mollenhauer et al., 156 157 2004).

In addition to the above sample set, we used a subset of published 16 core top samples from the Pacific Ocean to extend the dissolution intensity calibration over a larger range of deep

seawater $\Delta[CO_3^{2-}]$ values (Iwasaki et al., 2019, 2022). The core top sediment samples were 160 obtained by multiple core sampling from the water depth between 1500 and 4000 m, with most 161 of them located below the dissolution transition depth level ($\Delta [CO_3^{2-}] < 10 \ \mu mol \ kg^{-1}$). From 162 each core top sample, more than eight shells of each of three planktic foraminiferal species (G. 163 bulloides, G. ruber, and T. sacculifer) without external damage were collected from 200-355 µm 164 size fraction for analysis by X-ray micro-CT scanning in previous studies, and the existing scans 165 were here used for the calculation of the newly defined CT based dissolution index (Iwasaki et 166 al., 2019 and 2022). To remain consistent with the existing data and due to the difficulty to 167 unambiguously and consistently separating both species (Morard et al., 2019), specimens 168 designated as G. ruber may contain both G. ruber albus and G. elongatus. 169

Bottom water values of $\Delta[CO_3^{2^-}]$ for the all of studied sites were calculated from physicochemical parameters (temperature, salinity, total alkalinity, total inorganic carbon, and concentrations of phosphate and silicate) measured at nearby Global Ocean Data Analysis Project (GLODAP) profiles (Key et al., 2004). The site information and the estimated values of bottom water physicochemical properties for all Atlantic and Pacific sites are summarized in Table 1.





Figure 1. The locations of multiple corer samples in the South Atlantic used in this study. The section along the yellow line is showing the spatial distribution of the degree of carbonate saturation ($\Delta[CO_3^{2-}]$) in the study area.



Figure 2. B/Ca ratio in shells of the benthic foraminifera Cibicidoides wuellerstorfi in core top samples plotted against deep-water $\Delta[CO_3^{2-}]$. The data of B/Ca ratio of benthic foraminifera in the core-top sample used in this study are shown by blue squares. Data are derived from Rae et al. (2011), Raitzch et al. (2011), and Yu and Elderfield (2007).

Cruise	Site	Latitude (S)	longitude (W)	Depth (m)	Deep water Δ[CO ₃ ²] (µmol kg ⁻¹)	Linear Sedimentation Rate (cm kyr ⁻¹)	TOC accumulation rate (g m ⁻² y ⁻¹)	Caronate content (%)	C. w_B/Ca (µmol mol ⁻¹)	error
M 12/1	1206-1	24.67	-6.48	940	25.1	No data	No data	No data	193	4
M 12/1	1207-2	24.59	-6.85	2593	29.9	1.5	0.17	95.5	217	7
M 12/1	1209-1	24.50	-7.28	3303	14.6	3.3	0.41	95.9	191	9
M 12/1	1210-3	24.48	-7.43	3750	6.5	1.4	0.16	93.7	192	5
M 20/2	1715-1	26.47	-11.63	4097	-5.9	10.0	1.73	83.0	191	8
M 20/2	1720-3	29.00	-13.82	2004	33.2	6.0	3.30	78.8	208	9
M 20/2	1721-5	29.17	-13.08	3045	18.9	6.0	1.41	84.8	186	9
M 20/2	1728-3	29.83	-2.40	2887	23.8	2.0	0.30	94.9	229	7
M 20/2	1729-1	28.88	-1.00	4401	-0.1	0.6	0.08	93.6	211	6
M 34/3	3803-1	30.34	8.57	4173	3.1	0.8	0.16	90.0	199	10
M 34/3	3804-2	30.74	8.77	3882	7.8	0.7	0.12	92.2	184	10
M 34/3	3827-1	25.02	38.54	3842	-4.3	0.9	0.18	50.5	176	9
M 46/3	6330-1	46.15	57.56	3874	-13.1	No data	No data	No data	162	7

Table 1: Multiple core sample locations, depth (m), Deep water degree of Δ [CO₃^{2–}] (µmol kg⁻¹), Linear Sedimentation Rate (cm kyr⁻¹), TOC (Total Organic Caron) accumulation rate (g m⁻² y⁻¹)

and *Cibicidoides wuellerstorfi* B/Ca ratio (μ mol mol⁻¹) at each sampling site.

2.2. X-ray Micro-CT Scanning 195

196 The XMCT system (ScanXmate-D160TSS105/11000, WhiteRabbit Corp., Tokyo, Japan) at the Japan Agency for Marine-Earth Science and Technology, Yokosuka, Japan, was used to 197 obtain three-dimensional X-ray images of foraminiferal shells. The imaging was carried out with 198 a high-resolution setting (X-ray focus spot diameter, 0.8 µm; X-ray tube voltage, 80 kV; detector 199 array size, 2000 × 1336; 1500 projections/360°; 0.5 s/projection). After XMCT scanning, 200 ConeCTexpress software (WhiteRabbit Corp., Tokyo, Japan) was used to convert the raw 201 tomography data into segmented images of foraminifera shells. Image cross-sections were 202 reconstructed from filtered back projections following the general principle of Feldkamp cone-203 beam reconstruction. The scanning and data processing methods followed a previous study 204 [Iwasaki et al., 2015]. The CT number, indicating calcite density and visualizable porosity, was 205 calculated based on the X-ray attenuation coefficient of each sample. We used Molcer Plus 3-D 206 imaging software (WhiteRabbit Corp., Tokyo, Japan) to obtain isosurface images of the shells. 207 Then, we evaluated the CT number histograms of each shell based on the 3-D tomography data. 208

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2.3. Protocol of CT data processing for calculating CTDX 210

211 The CT scanning-based dissolution index (CTDX) of the planktic foraminiferal test employed in this study follows the same concept as the proxy of %Low-CT-number calcite 212 volume previously suggested (Iwasaki et al., 2019, 2022). However, in this study, we modify the 213 %Low-CT-number calcite volume proxy in a way that the resulting value is not dependent on the 214 exact settings of the scanner, enabling applications using other X-ray micro-CT scanners, which 215 had been modified to remove beam-hardening effects for assessing quantitative bulk density (i.e. 216 submicron scale porosity) of organic carbonate. The protocol to calculate the value of CTDX 217 from Serial Cross-Sectional Images of each specimen is shown in Figure 3. First, from the initial 218 raw data (sequential sectional images) (Figure 3a), contaminants (e.g., detritus, fragments and 219 other small foraminifera shells) stuck inside the test were removed by manual digital image 220 processing (Figure 3b). After that, to segment shell material from other environments, we 221 determined the lower threshold CT value by visual inspection of multiple specimens, ensuring 222 only shell material is included. Under the process of segmentation, to facilitate a replicable 223 smoothing of the shell surface and to make sure all voxels attached to the shell surface are 224 included, we introduce a step where the six-neighboring voxels of a voxel located on the air-225 calcite boundary are retained. This is important to maintain voxels that partially contain shell 226 material for the analysis. CT values in voxels retained after cleaning, segmentation, and 227 thresholding were used to generate a CT histogram of the foraminiferal shell that could be used 228 to calculate the dissolution index (Figure 3c). Previous studies [Iwasaki et al. 2019, 2022] 229 suggested that the CT number histogram changes from monomodal to bimodal distribution with 230 increasing dissolution. Based on this characteristic of change in the histogram shape with test 231 dissolution, they proposed the relative volume of low-CT-number calcite to the volume of calcite 232 in the whole shell (%Low-CT-number calcite volume) as a quantitative carbonate dissolution 233 proxy for dissolution. Here we build on this approach but develop a new Dissolution Index 234 (CTDX), which defines the threshold CT value independently of the scanner setting, using the 235 shape of the CT value histogram. To this end, we first scaled the original CT number histogram 236 by assigning 0 and 100 to the voxels with the lowest and highest CT values (Figure 3d). After 237 that, we calculated the CTDX as the area ratio of the lower part of the histogram (X-axis: 0-50) 238 to the whole histogram (X-axis: 0-100) (Figure 3e). Finally, we assigned a CTDX value to a 239 sample as the average CTDX value of at least eight individual specimens. The minimum number 240

- of shells needed to provide a representative estimate of the average preservation state of a given
- foraminifera species was determined by Iwasaki et al. (2022).
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- 244





Figure 3. The protocol to calculate the foraminiferal shell dissolution index (CTDX) starts from raw sequential X-ray image data (a). (b) Contaminants in the shell sample are removed by manual deleting. (c) Apply threshold to distinguish between calcite material and surrounding environment. Extract CT number histogram from cleaned and threshold 3D data. (d) Scaling X
and Y-axes of CT number histogram. (e) Calculation of CTDX (%).

251

252 **3 Results**

253 **3.1.Evaluating test dissolution by X-ray micro-CT scanning**

The X-ray micro-CT scanning enables us to observe the internal structure of the 254 foraminiferal shell and calcite density distribution in a single specimen's shell. The CT number 255 histogram provides an objective and quantitative means to evaluate microscale calcite density 256 distribution in each specimen. The Figure 4 shows the changes in the internal shell structure and 257 CT number histogram of four species of planktic foraminifera in three seafloor sediment samples 258 representing different locations and bottom water $\Delta[CO_3^{2-}]$ conditions. As expected, and in 259 agreement with previous studies (Johnson et al., 2010, Iwasaki et al., 2019), we find a 260 progression of shell dissolution with decreasing $\Delta[CO_3^{2-}]$. The dissolution first affects the septa 261 among the juvenile chambers and then spreads to the inner layer of the chambers in the final 262 whorl. With progressive dissolution, the CT number histograms show a shift from a unimodal 263 distribution with a single, high CT number peak to a bimodal distribution, followed by a trend 264 toward a unimodal density distribution with a peak at a low CT number. These are consistent 265 with the results of previous studies (Iwasaki et al., 2015, 2019), suggesting that the shape of the 266 CT number histogram, irrespective of the CT scanning settings, can be used to describe the 267 preservation state of the foraminiferal shell. 268

Assuming that the shape of the CT number histogram, converted to the CTDX value as 269 described above, quantify the degree of shell alteration due to selective dissolution (removal) of 270 the most susceptible parts of the shell, the CTDX-based average state of shells of a given species 271 of planktonic foraminifera should show a systematic relationship with deep-water chemistry. 272 This is important, because the CTDX value is independent of the size and shape of the analyzed 273 individual shells. It describes the portion of the shell that is affected by dissolution, irrespective 274 of the analyzed calcite volume. In theory, the CTDX could even be applied to multiple species of 275 foraminifera, but because the shell architectures among species vary significantly, we began by 276 277 comparing the G. bulloides CTDX with that of three other species (G. ruber, T. sacculifer, and G. inflata) in the same sediment samples to also evaluate the inter-species variation in 278 279 CTDX (Figure 5). The results shows that CTDX of G. bulloides significantly correlates with the other three species (G. ruber: $R^2 = 0.7$, T. sacculifer: $R^2 = 0.72$, G. inflata: $R^2 = 0.81$), suggesting 280 that the CTDX of each of the species should be applicable as a dissolution proxy. However, the 281 CTDX values among the species are offset, in particular for the relationship between CTDX of 282 283 G. bulloides, T. sacculifer and G. inflata, the slopes of regression lines are relatively high (T. sacculifer: 1.69, G. inflata: 1.39). This implies that T. sacculifer and G. inflata are more sensitive 284 to dissolution expressed by the CTDX index than G. bulloides. 285



Figure 4. Changes in cross-sectional isosurface images and CT number histograms of four major species of planktonic foraminiferal shells (*G. bulloides, G. inflata, G. ruber* and *T. sacculifer*) with the progression of dissolution. Shells were obtained from three core top samples (GeoB-

292 1728, 1729, and 3827). The condition of selected tests, which have CTDX similar to average of

293 each sample set, are shown. $\Delta [CO_3^{2^-}]$ at the nearest GLODAP stations at the core-top sample 294 depths ranged from -4.3 to 23.8 µmol kg⁻¹. The dashed line in the CT number histograms shows

- the threshold (50) between low and high values.
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- 297



303 **3.2.** Comparison between planktonic foraminiferal CTDX, bottom water $\Delta[CO_3^{2-}]$, MAR, 304 and organic carbon flux

To assess the main controlling factors on foraminifera shell dissolution on the deep seafloor, we compared the planktonic foraminiferal shell CTDX values as a carbonate dissolution index, with three parameters: Bottom water $\Delta[CO_3^{2^-}]$ representing ambient seawater corrosiveness, Linear Sedimentation Rate describing the exposure time of the shells on the seafloor, and Organic carbonate accumulation rate approximating supralysoclinal dissolution due to elevated porewater CO_2 from organic matter remineralization, for the 13 Atlantic samples where all parameters were available (Figure 6). First, we found a significant correlation between the

planktonic foraminiferal CTDX and the bottom water $\Delta[CO_3^{2^-}]$ under the low $\Delta[CO_3^{2^-}]$ condition (< 10 µmol kg⁻¹): *G. bulloides*; R² = 0.87, *G. inflata*; R² = 0.85, *G. ruber*; R² = 0.77, *T.* 312 313 sacculifer; $R^2 = 0.80$ (Figure 6a). In contrast, the comparison between CTDX and sedimentation 314 rate site did not show any support for the hypothesis that higher sedimentation rate facilitates 315 better preservation of carbonate by faster burial (Figure 6b). These results suggest that the 316 carbonate dissolution intensity in the South Atlantic is generally governed by ottom water 317 Δ [CO₃^{2–}] under low Δ [CO₃^{2–}] condition (< 10 µmol kg⁻¹), not by sedimentation rate. Under high 318 Δ [CO₃²⁻] conditions (> 10 µmol kg⁻¹), high CTDX values are observed at two sites (GeoB 1720 319 and 1721), suggesting significant dissolution of carbonate, despite the supersaturated conditions 320 for calcite in the ambient seawater. Here, a comparison with organic carbon accumulation rate (g 321 $cm^{-2} kyr^{-1}$) reveals that the organic carbon inputs at both sites is more than three times higher 322 than at the other sites (Figure 6c). These results indicate that organic carbon decomposition in the 323 seafloor sediment may affect preservation of planktonic foraminifera shells even under 324 supralysoclinal conditions (> 10 μ mol kg⁻¹). 325

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Bottom water $\Delta[CO_3^{2-1}]$ (µmol kg⁻¹) Linear Sedimentation Rate (cm kyr⁻¹) Organic carbon accumulation rate (g cm⁻² kyr⁻¹) Bottom water $\Delta[CO_3^{2-1}]$ (µmol kg⁻¹), (b) Liner Sedimentation Rate (cm kyr⁻¹), and (c) Organic carbon accumulation rate (g cm⁻² kyr⁻¹) versus Planktonic foraminifera CTDX (%) at each sampling site. The data points at the sites locate in the Benguela upwelling system (GeoB-1715, 1720 and 1721) with high organic carbon accumulation rate are surrounded by dotted line.

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335 **3.3.**Comparison with the proxy of benthic foraminiferal B/Ca ratio

To investigate the applicability of the CTDX as a deep seawater Δ [CO₃²⁻] proxy, we next 336 compared the CTDX values in the 13 South Atlantic samples with the B/Ca ratio of benthic 337 foraminifera (C. wuellerstorfi) (Figure 7). The B/Ca ratio of benthic foraminifera (C. 338 *wuellerstorfi*), a conventional quantitative proxy of deep seawater $\Delta[CO_3^{2-}]$, derived from the 339 same sediment samples showed a weak correlation with the CTDX of each species: G. bulloides: 340 $R^2 = 0.42, p = 0.0169^*, G. inflata; R^2 = 0.24, p = 0.1034, G. ruber; R^2 = 0.27, p = 0.1034, T.$ 341 sacculifer; $R^2 = 0.28$, p = 0.0960 (Figure 7). The correlation has in all cases the expected sign 342 (both proxies indicating more dissolution in the same samples), but the observation that the 343 CTDX and B/Ca ratio correlation is not as pronounced as the correlation between CTDX and the 344 bottom water $\Delta[CO_3^{2^-}]$ is puzzling. Next, we focused on the G. bulloides CTDX, because this 345 species was found at all sites, and showed the most significant correlation with the benthic 346 347 foraminiferal B/Ca ratio among four species. The Figure 8 shows the relationship between two

proxies (G. bulloides CTDX and benthic foraminifera B/Ca ratio) and the bottom water Δ [CO₃²⁻] 348 under the lower deep seawater $\Delta[CO_3^{2-}]$ condition (< 10 µmol kg⁻¹). Based on this comparison, 349 we identified the sites where either proxy is discrepant, and investigated the factors that may be 350 responsible for the discrepancy in these two proxies. In this plot, the regression lines between G. 351 *bulloides* CTDX and bottom water $\Delta[CO_3^{2^-}]$ obtained from the results of this study and that 352 between the benthic foraminiferal B/Ca ratio and bottom water $\Delta[CO_3^{2-}]$ obtained from the 353 results of calibration using more than 200 core-top samples (Yu et al., 2013) were presented. The 354 result shows that the benthic foraminiferal B/Ca ratio is at least 20 µmol mol⁻¹ higher than the 355 regression line at two sites: Site 1715 (Depth: 4097 m, bottom water $\Delta[CO_3^{2-}]$: -5.87 µmol kg⁻¹); Site 1729 (Depth: 4401 m, bottom water $\Delta[CO_3^{2-}]$: -0.10 µmol kg⁻¹). At those sites, the 356 357 estimated value of bottom water $\Delta[CO_3^{2-}]$ derived from the benthic foraminiferal B/Ca ratio 358 imply higher than observed $\Delta[CO_3^{2-}]$. 359







Figure 7. Relationship between planktonic foraminiferal CTDX of (a) *G. bulloides*, (b) *G. inflata*, (c) *G. ruber*, and (d) *T. sacculifer* and B/Ca ratio of *C. wuellerstorfi* (μ mol mol⁻¹) in each core-top sample. The sampling sites located in the Benguela upwelling system with high organic carbon flux are shown by gray circle. The regression line and the square of the correlation coefficient (\mathbb{R}^2) are also shown.

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Figure 8. Comparison of the plots of *G. bulloides* CTDX and B/Ca ratio of *C. wuellerstorfi* against bottom seawater Δ [CO₃²⁻] under the condition of low bottom seawater Δ [CO₃²⁻] (< 10 µmol kg⁻¹). The B/Ca ratio of *C. wuellerstorfi* at the site GeoB-1715, 1729 and 3803 are relatively higher than *G. bulloides* CTDX.

378 **3.4.Extended calibration between CTDX and deep water** Δ [CO₃²⁻]

In addition to the 13 core top samples collected in the South Atlantic, we added the 16 core 379 top samples collected in the Pacific Ocean to assess the behavior of the CTDX proxy under the 380 condition of low deep seawater $\Delta[CO_3^{2-}]$ (< 10 µmol kg⁻¹) and to obtain an extended 381 calibration equation between CTDX and deep seawater $\Delta[CO_3^{2-}]$ in the wider range of global 382 ocean. The plot shows the relationship between the CTDX of three species of major planktic 383 for a miniferal shell (G. bulloides, G. ruber and T. sacculifer) and deep water Δ [CO₃²⁻] obtained 384 from nearby bottle water sampling at each site (Figure 9). The sites at GeoB 1720 and 1721, 385 with high organic material deposition, are excluded from the plot. The plot provides the 386 relationship between CTDX and a wide range of deep seawater $\Delta [CO_3^{2-}]$ (-27 to 30 μ mol kg⁻¹), 387 and suggests that CTDX of each species significantly correlate with deep seawater $\Delta [CO_3^{2^-}]$ between the range of -20 to 10 µmol kg⁻¹. The results of calibrations in each species are as 388 389 390 follows:

392	G. bulloides: Deep seawater $\Delta [CO_3^{2^-}] = -0.65 * CTDX + 24.8 (R^2 = 0.57, N = 15)$
393	<i>G. ruber</i> : Deep seawater Δ [CO ₃ ²⁻] = -0.56 * CTDX + 20.5 (R ² = 0.88, N = 12)
394	<i>T. sacculifer:</i> Deep seawater $\Delta [CO_3^{2-}] = -0.43 * CTDX + 15.3 (R^2 = 0.88, N = 13)$

These calibrations revealed that the CTDX of planktonic foraminifera are able to works as deep-396 water $[CO_3^{2^-}]$ proxy in the specific range of the deep seawater $\Delta[CO_3^{2^-}]$ (-20 to 10 μ mol kg⁻¹). 397 However, the variation in the slope of regression line for each species indicated that the 398 sensitivity of CTDX to dissolution varied among species. Based on the regression analysis, the 399 uncertainties of the above regression equations are $\pm 5.7 \ \mu mol \ kg^{-1}$ (G. bulloides, N = 15), ± 2.8 400 μ mol kg⁻¹ (G. ruber, N = 12), and $\pm 2.6 \mu$ mol kg⁻¹ (T. sacculifer, N = 13). On the other hand, the 401 CTDX shows stable values of around 25 and 60 under the high (> 10 μ mol kg⁻¹) and low (< -20 402 μ mol kg⁻¹) carbonate saturation state condition, respectively. 403





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Figure 9. The CTDX of planktonic foraminifera (*G. bulloides, G. ruber* and *T. sacculifer*) in core top samples plotted against deep-water Δ [CO₃^{2–}]. Core-top samples are obtained from this study and previous studies (Iwasaki et al., 2019 and 2022).

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411 4 Discussion

412 **4.1.Controlling factors of the planktonic foraminiferal CTDX**

Dissolution of the planktonic foraminiferal shell is considered to occur mainly on deep 413 seafloor under the control of ambient seawater Δ [CO₃^{2–}]. At the same time, it can be affected by 414 the variance in sedimentation rate and organic carbon respiration at the sediment surface [e.g., 415 Berger et al., 1982, Milliman, 1993]. The direct comparisons between three numerical indexes 416 derived from deep seafloor in this study (i.e., planktic foraminiferal CTDX, bottom water 417 Δ [CO₃^{2–}], the sedimentation rate, and the organic carbon accumulation rate at each site) provided 418 the critical information to identify the controlling factor of carbonate dissolution at the deep 419 seafloor. First, the significant correlation between the proxy of CTDX and bottom water $\Delta [CO_3^2]$ 420

] proves that dissolution of the planktic foraminiferal shell is undoubtedly controlled by bottom 421 water $\Delta[CO_3^{2-}]$ in the most sites under the lower condition of bottom water $\Delta[CO_3^{2-}]$ (< 10 µmol 422 kg⁻¹). Second, the sedimentation rate is not correlated with the planktic foraminiferal CTDX. 423 This result suggests that the variance in sedimentation rate, which corresponds to approx. 5-to-424 10-fold difference in sedimentation rate does not significantly impact the carbonate dissolution 425 variance at the deep seafloor. Third, the CTDX showed significantly higher values at two data 426 sites (GeoB-1720 and 1721) than at the other data points under the condition of higher deep 427 seawater $\Delta [CO_3^{2-}]$ (> 10 µmol kg⁻¹), which implies the exceeded carbonate dissolution under the 428 supersaturated condition. These two sites are located at the Benguela Upwelling System in the 429 eastern South Atlantic, one of the significant continental margins upwelling systems and belongs 430 to the high productivity area of the ocean [Berger, 1989]. Previous research in the Benguela 431 Upwelling System suggested that carbonate dissolution occurs at 400-1600 m above the 432 lysocline by the above process (Vobers and Hinrich, 2002), induced by the release of CO₂ due to 433 the oxidation of organic material within the sediment (Emerson and Bender, 1981; Archer, 1991; 434 Jahnke et al., 1994). In addition, the other potential factor is that the porous shell, possibly 435 response to rapid chamber formation, resulting in the formation of shells that are prone to 436 437 dissolution. Culture studies have shown that the chamber formation rate increases under the food abundant condition as like as upwelling region (Bé et al., 2009), and such high chamber 438 formation rate could provide the porous shell formation (Berger, 1970). In either case, we 439 440 consider that the observed dissolution of planktonic foraminifera shells in the sediments derived from the Benguela Upwelling System in this study is not controlled by deep seawater $\Delta[CO_3^{2-}]$, 441 but was probably brought yb other factors such as decomposition of organic material occurring 442 above the regional lysocline and ontogenetic growth condition at sea surface. 443

On the other hand, the relationship between benthic foraminiferal B/Ca ratio and deep-444 water $\Delta[CO_3^{2-}]$ did not show any unusual values at the sites from the Benguela Upwelling 445 System, suggesting that the organic material decomposition does not have notable impact on 446 benthic foraminiferal B/Ca ratio. We suppose that the reduction in seawater $\Delta[CO_3^{2-}]$, caused by 447 organic material decomposition, has a more substantial influence on the dissolution of the 448 planktonic foraminiferal test than the reduction in the B/Ca ratio of the benthic foraminiferal test. 449 The mechanism to explain these results is to hypothesize that the decrease in seawater Δ [CO₃²⁻] 450 due to organic material decomposition occurs mainly inside the sediment or aggregates of 451 organic material like marine snow or fecal pellets. Such organic material oxidization affects 452 planktonic foraminifera shells co-deposited with such aggregates, but, it does not significantly 453 affect the seafloor surface or outside of the aggregates (i.e., the habitat area of benthic 454 foraminifera). These findings suggests that the planktonic foraminiferal CTDX is not a suitable 455 proxy at sites with high organic material deposition like upwelling system. The benthic 456 foraminiferal B/Ca ratio is rather appropriate for use in such sample sites. 457

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460 **4.2. Deep seawater** $[CO_3^{2-}]$ proxies: The characteristics of CTDX and B/Ca proxy

In this study, the characteristics of CTDX proxy were determined by measuring the dissolution intensity of four species of planktic foraminifera and comparing the several parameters derived from seafloor sediment samples in the South Atlantic. First, the results of inter-species comparisons (Figure 5) showed that the CTDX varies among the species, suggesting that species-specific application is required for accurate reconstruction. Nevertheless, our results suggested that the CTDX of all species are applicable as shell dissolution proxy. This indicates that multiple species of planktonic foraminifera are applicable as deep-water $[CO_3^{2^-}]$ proxy by establishing proxy calibration for each species. This is one of the essential advantages of using CTDX over conventional proxies based on the B/Ca ratio of benthic foraminifera. Therefore, it enables us to apply this new proxy to the foraminifera poor sediment cores, where it is challenging to collect single species continuously. It also allows us to compare the reconstructed data between sites from different locations with different planktic locations foraminiferal species compositions.

Second, the relationship between the CTDX and the deep seawater Δ [CO₃²⁻] suggested 474 that the CTDX significantly correlated with deep seawater $\Delta [CO_3^{2-}]$ only under the low deep 475 seawater $\Delta[CO_3^{2^-}]$ condition (< 10 µmol kg⁻¹) (Figure 6, and Figure 9). However, under the 476 high deep seawater $\Delta[CO_3^{2-}]$ condition (> 10 µmol kg⁻¹), the values of planktic 477 foraminiferal CTDX were almost constant. Similar results were also represented by the 478 comparison between shell dissolution proxy and deep seawater Δ [CO₃²⁻], suggesting the 479 saturation of shell dissolution proxy at the sites of high deep seawater $\Delta [CO_3^{2-}]$ (> 10 480 µmol/kg) [Johnstone et al., 2010]. Furthermore, under the extremely low deep seawater 481 Δ [CO₃²⁻] condition (< -20 µmol kg⁻¹), the CTDX seems to saturate around the values of 60 482 (Figure 9). This implies that planktonic foraminiferal shells which appear intact (i.e., which 483 are not broken or fragmented and retain the outermost chamber) show maximum CTDX 484 values of around 60. It is likely that with further progressing dissolution the shells lose 485 structural integrity, break into fragments and would fall outside of the sample selection for 486 CT scanning. From the above, we conclude that the CTDX proxy works effectively under 487 deep seawater $\Delta[CO_3^{2-}]$ lower than 10 µmol kg⁻¹ and higher than -20 µmol kg⁻¹. 488 Alternatively, we suppose that the analysis of fragmented individual is useful for applying 489 CTDX under the significantly low deep seawater Δ [CO₃²⁻] condition (< -20 µmol kg⁻¹). 490

We also evaluated the accuracy of the deep seawater $[CO_3^{2-}]$ reconstruction by each 491 proxy. Under the low deep seawater $\Delta [CO_3^{2-}]$ (< 10 µmol/kg) condition, the proxy of CTDX 492 showed a correlation with the deep seawater $\Delta [CO_3^{2^-}]$ and the conventional proxy of benthic 493 foraminiferal B/Ca ratio (Figure 7). These results support our interpretation that the proxy of 494 CTDX can serve as an indicator of deep seawater $\Delta [CO_3^{2-}]$. Nevertheless, comparing these two 495 proxies, we found two peculiar sites (Sites GeoB-1715 and 1729) where the CTDX and B/Ca 496 ratio values deviate (Figure 8). The sediment samples at these two sites are rich in carbonate (> 497 80%), and the proxy of CTDX shows the occurrence of carbonate dissolution and relatively high 498 values of benthic foraminiferal B/Ca ratio than general estimation. We speculate that the process 499 of carbonate compensation in surface sediment may have raised $[CO_3^{2-}]$ in the porewater and 500 contributed to the higher values of the benthic foraminiferal B/Ca ratio in a way similar to the 501 speculated protective effect of aragonite dissolution on carbonate preservation described by 502 Sulpice et al. (2022). To evaluate this speculation, we investigated the difference in total 503 alkalinity of porewater in the sediment surface (upper 5 cm) and the overlying bottom water 504 (Hensen et al., 2003a-d). The total alkalinity is a valuable indicator that relates to the seawater 505 $[CO_3^{2-}]$ and is controlled by the carbonate dissolution in the sediment, which enables us to 506 presume the geochemical process in the sediment surface. The profiles of chemical parameters at 507 508 Site GeoB-1715 and 1729 showed that the total alkalinity in the pore water (average of the upper 5 cm) is more than 200 μ mol kg⁻¹ higher than in overlying bottom water as follows: GeoB-1715 (bottom water: 2354 μ mol kg⁻¹, porewater average: 2583 μ mol kg⁻¹) and GeoB-1729 (bottom 509 510 water: 2359 µmol kg⁻¹, porewater average: 2653 µmol kg⁻¹). In contrast, at Site GeoB-3804 and 511 3827 with appropriate benthic foraminiferal B/Ca ratio for the general estimation, the difference 512

in the total alkalinity between pore water and overlying bottom water is relatively small as follows: GeoB-3804 (bottom water: 2348 µmol kg⁻¹, porewater average: 2458 µmol kg⁻¹) and GeoB-3827 (bottom water: 2342 µmol kg⁻¹, porewater average: 2275 µmol kg⁻¹). These results imply that carbonate dissolution at the sediment surface discharges carbonate ions and raises the porewater's total alkalinity, altering the calcification condition for benthic foraminifera. From the above, we consider that the strong carbonate dissolution in the carbonate-rich sediment is one of the principles controlling factors of the benthic foraminiferal B/Ca ratio.

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521 5 Conclusions

The assessments in planktic foraminiferal shell dissolution intensity, represented by 522 CTDX, were directly compared with the data of B/Ca in benthic foraminifera in the same sample 523 and the other proxies of conditions at each site (deep-water Δ [CO₃²⁻], sedimentation rate, and 524 Organic carbon accumulation rate). In addition, inter-species variation in CTDX of four species 525 of planktic foraminifera were assessed. Our results proved that CTDX of each planktic 526 for a for a second deep-water $\Delta [CO_3^{2-}]$ variation like a conventional proxy 527 of benthic foraminiferal B/Ca ratio. We suggested that four species (G. bulloides, G. inflata, G. 528 *ruber*, and *T. sacculifer*) of planktonic foraminiferal CTDX are applicable as a quantitative proxy 529 for carbonate dissolution intensity, while we recommend species-specific use of CTDX for 530 accurate reconstruction because there is slight difference in sensitivity to dissolution. On the 531 other hand, the effect of variation in sedimentation rate is not a significant factor in carbonate 532 533 dissolution in the sediment surface if the difference in sedimentation rate is 5-to-10-fold difference or less. Based on the above results, we concluded that the proxy of planktic 534 for a for a miniferal CTDX is applicable as a quantitative deep-water $[CO_3^{2-}]$ proxy as well as a 535 conventional proxy of benthic foraminiferal B/Ca ratio. Nevertheless, there are several caveats in 536 the application of each proxy. First, the proxy of CTDX is useful under the specific condition of 537 deep-water Δ [CO₃^{2–}] between -20 to 10 µmol kg⁻¹, suggesting that the CTDX has a detection 538 limit range of deep-water $[CO_3^{2-}]$. Therefore, we should exclude the seafloor sediment sample 539 obtained from high deep-water $\Delta[CO_3^{2-}]$ condition (> 10 µmol kg⁻¹) in the process of 540 establishing the calibration equation, and we have to note that this proxy is not reliable for 541 reconstructing the higher deep-water $\Delta [CO_3^{2-}]$ in the application of CTDX into sediment core 542 samples. Second, we also observed the excessive dissolution of planktic foraminifera in the 543 supersaturated condition for calcite, which seems to be caused by the intensive input of organic 544 material into seafloor sediment. Therefore, we consider that we should refrain from applying the 545 CTDX proxy to the sediment sample located in the upwelling system where organic material 546 547 input from the sea surface is exceptionally high. Third, we revealed that carbonate compensation, which occurs in carbonate-rich and carbonate dissolution affected sediment samples, may alter 548 the benthic foraminiferal B/Ca ratio higher during their calcification, which may contribute to 549 uncertainty of deep-water $[CO_3^{2-}]$ reconstruction by benthic foraminiferal B/Ca ratio. 550

Based on the suggested characteristics of CTDX, we provided the calibration formula between three species planktic foraminiferal (*G. bulloides, G. ruber* and *T. sacculifer*) CTDX and deep seawater Δ [CO₃^{2–}] using a number (N = 22) of seafloor sediment samples. Although the number of sediment samples and the distribution range of samples are inferior to the conventional proxy of benthic foraminiferal B/Ca ratio, our calibration showed that the proxy of CTDX works as well as the B/Ca proxy for deep seawater [CO₃^{2–}] reconstruction under the suitable conditions. This proxy may help to fill the blank of paleo-deep-water [CO₃^{2–}] data, in particular, in the mid-high latitude of the North Pacific, where benthic foraminiferal B/Ca ratio

- 559 data are insufficient.
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568 569

570 Open Research

- 571 The data used in this study will be available in PANGAEA (https://www.pangaea.de/).
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761 **Figure chaptions**

Figure 1. The locations of multiple corer samples in the South Atlantic used in this study. The section along the yellow line is showing the spatial distribution of the degree of carbonate saturation ($\Delta[CO_3^{2^-}]$) in the study area.

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Figure 2. B/Ca ratio in shells of the benthic foraminifera *Cibicidoides wuellerstorfi* in core top samples plotted against deep-water $\Delta[CO_3^{2^-}]$. The data of B/Ca ratio of benthic foraminifera in the core-top sample used in this study are shown by blue squares. Data are derived from Rae et al. (2011), Raitzch et al. (2011), and Yu and Elderfield (2007).

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Figure 3. Protocol to calculate the foraminiferal shell dissolution index (CTDX) start from raw sequential X-ray image data (a). (b) Contaminants in the shell sample are removed by manual deleting. (c) Apply threshold to distinguish between calcite material and surrounding environment. Extract CT number histogram from cleaned and threshold 3D data. (d) Scaling X and Y-axes of CT number histogram. (e) Calculation of CTDX (%).

- 777 Figure 4. Changes in cross-sectional isosurface images and CT number histograms of four major species of planktonic foraminiferal shells (G. bulloides, G. inflata, G. ruber and T. sacculifer) 778 779 with the progression of dissolution. Shells were obtained from three core top samples (GeoB-1728, 1729, and 3827). The condition of selected tests, which have CTDX similar to average of 780 each sample set, are shown. $\Delta[CO_3^{2-}]$ at the nearest GLODAP stations at the core-top sample 781 depths ranged from -4.3 to 23.8 µmol kg⁻¹. The dashed line in the CT number histograms shows 782 the threshold (50) between low and high values.
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Figure 5. Inter species comparison of G. bulloides CTDX with other three species obtained from 785 same core-top samples. The square of the correlation coefficient (\mathbb{R}^2) and the type I error rate (p-786 value) are also shown. 787

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Figure 6. Investigation in controlling factor of foraminiferal shell dissolution intensity. (a) 789 Bottom water $\Delta[CO_3^{2^-}]$ (µmol kg⁻¹), (b) Liner Sedimentation Rate (cm kyr⁻¹), and (c) Organic carbon accumulation rate (g cm⁻² kyr⁻¹) versus Planktonic foraminifera CTDX (%) at each 790 791 sampling site. The data points at the sites locate in the Benguela upwelling system (GeoB-1715, 792 1720 and 1721) with high organic carbon accumulation rate are surrounded by dotted line. 793

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Figure 7. Relationship between planktonic foraminiferal CTDX of (a) G. bulloides, (b) G. 795 *inflata*, (c) G. *ruber*, and (d) T. *sacculifer* and B/Ca ratio of C. *wuellerstorfi* (μ mol mol⁻¹) in each 796 core-top sample. The sampling sites located in the Benguela upwelling system with high organic 797 carbon flux are shown by gray circle. The regression line and the square of the correlation 798 coefficient (R^2) are also shown. 799

800 Figure 8. Comparison of the plots of G. bulloides CTDX and B/Ca ratio of C. wuellerstorfi 801 against bottom seawater $\Delta[CO_3^{2-}]$ under the condition of low bottom seawater $\Delta[CO_3^{2-}]$ (< 10 802 μ mol kg⁻¹). The B/Ca ratio of C. wuellerstorfi at the site GeoB-1715, 1729 and 3803 are 803 relatively higher than G. bulloides CTDX. 804

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Figure 9. The CTDX of planktonic foraminifera (G. bulloides, G. ruber and T. sacculifer) in 806 core top samples plotted against deep-water Δ [CO₃^{2–}]. Core-top samples are obtained from this 807 study and previous studies (Iwasaki et al., 2019 and 2022). 808

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Table 1. Multiple core sample locations, depth (m), Deep water degree of $\Delta[CO_3^{2-}]$ (µmol kg⁻¹), 810 Linear Sedimentation Rate (cm kyr⁻¹), TOC (Total Organic Caron) accumulation rate (g $m^{-2} y^{-1}$) 811

- and *Cibicidoides wuellerstorfi* B/Ca ratio (μ mol mol⁻¹) at each sampling site. 812
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