

NF- κ B pathway as a molecular target for curcumin in diabetes mellitus treatment: Focusing on oxidative stress and inflammation

Mohammad Yasin Zamanian¹, Hashem O. Alsaab², Maryam Golmohammadi³, Alexey Yumashev⁴, Abeer mhussan jabbar⁵, Mohammed Kadhem Abid⁶, Abhishek Joshi⁷, Ahmed Alawadi⁸, Noor S. Jafer⁹, Farzaneh Kianifar¹⁰, and Samuel Baker Obakiro¹¹

¹Hamadan University of Medical Sciences Medical School

²Taif University

³Shahid Beheshti University of Medical Sciences School of Medicine

⁴Pervyj Moskovskij gosudarstvennyj medicinskij universitet imeni I M Secenova

⁵National University of Science and Technology

⁶Al-Ayen University

⁷King Mongkut's University of Technology Thonburi School of Liberal Arts

⁸The Islamic University Technical Engineering College

⁹Al-Rafidain University College

¹⁰Iran University of Medical Sciences School of Behavioral Sciences and Mental Health

¹¹Busitema University

April 05, 2024

Abstract

Diabetes mellitus (DM), a chronic metabolic disorder associated with hyperglycemia and other complications, is one of the five priority non communicable diseases of global interest with unprecedented rise in developing countries. Whereas, the current treatment with insulin and oral hypoglycemic agents is aimed at managing the hyperglycemia and associated complications, there is need to explore other critical pathways in the pathogenesis of DM that can act as potential drug targets with better treatment outcomes. This study comprehensively explains the role of cellular and molecular elements, like hyperglycemia-induced oxidative stress, endothelial dysfunction, and Nuclear Factor Kappa B (NF- κ B)'s involvement in inflammation and immune regulation, in the onset of DM. With bioactive compounds from natural products gaining popularity as novel drug molecules due to their diverse pharmacological actions, the study also extensively explores the prospective therapeutic benefits of curcumin (CUR), a bioactive compound known for its antioxidant, anti-inflammatory, and hypoglycemic properties, in addressing diabetic complications, predominantly via the modulation of the NF- κ B pathway. The findings reveal that CUR administration effectively lowered blood glucose elevation, reinstated diminished serum insulin levels, and enhanced body weight in Streptozotocin -induced diabetic rats. CUR exerts its beneficial effects in management of diabetic complications through regulation of signaling pathways, such as CaMKII, PPAR- γ , NF- κ B, and TGF- β 1. Moreover, CUR reversed the heightened expression of inflammatory cytokines (TNF- α , IL-1 β , IL-6) and chemokines like MCP-1 in diabetic specimens, vindicating its anti-inflammatory potency in counteracting hyperglycemia-induced alterations. CUR diminishes oxidative stress, avert structural kidney damage linked to diabetic nephropathy, and suppress NF- κ B activity. Furthermore, CUR exhibited a protective effect against diabetic cardiomyopathy, lung injury, and diabetic gastroparesis. Conclusively, the study posits that CUR could potentially offer therapeutic benefits in relieving diabetic complications through its influence on the NF- κ B pathway.

ΝΦ-κΒ παθηωψ ας α μολεςυλαρ ταρχετ φορ ςυρςυμιν ιν διαβετες μελλιτυς τρεατμεντ: Φοςυσινγ ον οξειδατιε στρεσς ανδ ινφλαμματιον

Mohammad Yasin Zamanian ^{1,2*}, Hashem O. Alsaab³, Maryam Golmohammadi ⁴, Alexey Yumashev ⁵, Abeer mhussan jabbar ⁶, Mohammed Kadhem Abid ⁷, Abhishek Joshi⁸, Ahmed Hussien Alawadi^{9,10,11}, Noor S. Jafer ¹², Farzaneh Kianifar ¹³, Samuel Baker Obakiro^{14*}

¹Department of Physiology, Hamadan University of Medical Sciences, Hamadan 6718773654, Iran.

²Department of Pharmacology and Toxicology, School of Pharmacy, Hamadan University of Medical Sciences, Hamadan 6718773654, Iran.

³ Department of Pharmaceutics and Pharmaceutical Technology, Taif University, Taif 21944, Saudi Arabia

⁴ School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran 198887355, Iran.

⁵ Department of Prosthetic Dentistry, Sechenov First Moscow State Medical University, Trubetskaya st. 8-2, Moscow 119991, Russian Federation

⁶ colleges of pharmacy, National University of Science and Technology, Dhi Qar, Iraq

⁷ Department of Anesthesia, College of Health & medical Technology, Al-Ayen University, Thi-Qar, Iraq

⁸Department of Liberal Arts School of Liberal Arts, Uttaranchal University, Dehradun-248007, India

⁹College of technical engineering, the Islamic University, Najaf, Iraq

¹⁰ College of technical engineering, the Islamic University of Al Diwaniyah, Iraq

¹¹ College of technical engineering, the Islamic University of Babylon, Iraq

¹² Department of Medical Laboratory Technologies, Al Rafidain University College, Bagdad, Iraq

¹³ School of Medicine, Iran University of Medical Sciences, Tehran, Iran

¹⁴ Department of Pharmacology and Therapeutics, Faculty of Health Sciences, Busitema University, P.O. Box 1460, Mbale, Uganda

***Corresponding Authors:**

Mohammad Yasin Zamanian (mzamanian52@yahoo.com, Tel: +989187018850)

,Dr. Samuel Baker Obakiro (sobakiro@gmail.com)

Abstract

Diabetes mellitus (DM), a chronic metabolic disorder associated with hyperglycemia and other complications, is one of the five priority non communicable diseases of global interest with unprecedented rise in developing countries. Whereas, the current treatment with insulin and oral hypoglycemic agents is aimed at managing the hyperglycemia and associated complications, there is need to explore other critical pathways in the pathogenesis of DM that can act as potential drug targets with better treatment outcomes. This study comprehensively explains the role of cellular and molecular elements, like hyperglycemia-induced oxidative stress, endothelial dysfunction, and Nuclear Factor Kappa B (NF- κ B)'s involvement in inflammation and immune regulation, in the onset of DM. With bioactive compounds from natural products gaining popularity as novel drug molecules due to their diverse pharmacological actions, the study also extensively explores the prospective therapeutic benefits of curcumin (CUR), a bioactive compound known for its antioxidant, anti-inflammatory, and hypoglycemic properties, in addressing diabetic complications, predominantly via the modulation of the NF- κ B pathway.

The findings reveal that CUR administration effectively lowered blood glucose elevation, reinstated diminished serum insulin levels, and enhanced body weight in Streptozotocin -induced diabetic rats. CUR exerts its beneficial effects in management of diabetic complications through regulation of signaling pathways, such

as CaMKII, PPAR- γ , NF- κ B, and TGF- β 1. Moreover, CUR reversed the heightened expression of inflammatory cytokines (TNF- α , IL-1 β , IL-6) and chemokines like MCP-1 in diabetic specimens, vindicating its anti-inflammatory potency in counteracting hyperglycemia-induced alterations. CUR diminishes oxidative stress, avert structural kidney damage linked to diabetic nephropathy, and suppress NF- κ B activity. Furthermore, CUR exhibited a protective effect against diabetic cardiomyopathy, lung injury, and diabetic gastroparesis. Conclusively, the study posits that CUR could potentially offer therapeutic benefits in relieving diabetic complications through its influence on the NF- κ B pathway.

Keywords: Curcumin; Diabetes mellitus; NF- κ B pathway; Antioxidant, Anti-inflammatory

Significance statement:

This study explores the role of cellular and molecular elements, such as hyperglycemia-induced oxidative stress, endothelial dysfunction, and the involvement of the NF- κ B pathway in inflammation and immune regulation in the onset of diabetes mellitus (DM).

It extensively investigates the potential therapeutic benefits of curcumin (CUR), a bioactive compound known for its antioxidant, anti-inflammatory, and hypoglycemic properties, in addressing diabetic complications, predominantly via the modulation of the NF- κ B pathway.

The study highlights the importance of the NF- κ B pathway as a molecular target for CUR in the treatment of diabetes mellitus, focusing on oxidative stress and inflammation.

Abbreviations:

COX-2: Cyclooxygenase-2

TNF- α : Tumor necrosis factor α

IRS1: Insulin receptor substrate 1

GLUT2: Glucose transporter 2

GLUT4: Glucose transporter 4

STAT: Signal transducer and activator of transcription

MMP-3: Matrix metalloproteinase-3

MMP-9: matrix metalloproteinase-9

IL-1: Interleukin-1

IL-6: Interleukin-6

IL-8: Interleukin-8

IL-12: Interleukin-12

LPS: Lipopolysaccharides

I κ B: Inhibitor of nuclear factor kappa B

CYLD: Cylindromatosis

CD40: Cluster of differentiation 40

RANK: Receptor activator of nuclear factor kappa B

BAFF: B-cell activating factor

CaMKII: Calcium-calmodulin (CaM)-dependent protein kinase II

PPAR- γ : Peroxisome proliferator-activated receptor gamma

TGF- β 1: Transforming growth factor beta 1

TXA2: Thromboxane A2

PGI2: Prostacyclin

NO: Nitric oxide

PGE2: Prostaglandin E2

iNOS: Inducible nitric oxide synthase

MDA: Malondialdehyde

MPO: Myeloperoxidase

SOD: Superoxide dismutase

GSH: Glutathione

JNK: c-Jun N-terminal kinase

ERK1: Extracellular signal-regulated kinase 1

p38MAPK: p38 mitogen-activated protein kinases are a class of mitogen-activated protein kinase

AMPK: AMP-activated protein kinase

MCP-1: Monocyte chemoattractant protein-1

ICAM-1: Intercellular adhesion molecule 1

NADPH: Nicotinamide adenine dinucleotide phosphate

eNOS: Endothelial nitric oxide synthase

PKC β : Protein kinase C β

MPTP: Mitochondrial permeability transition pore

FOXO-3a: Forkhead box O3

DOP mouse: Diabetic osteoporosis mouse

SCF: Stem cell factor

ICCs: Interstitial cells of Cajal

OS: Oxidative stress

ROS: Reactive oxygen species

1. Introduction

Diabetes mellitus (DM), a widespread chronic metabolic anomaly, manifests through heightened blood sugar levels (hyperglycemia) accompanied by symptoms including, excessive urination (polyuria), intensified thirst (polydipsia) and pronounced hunger (polyphagia) [1]. The worldwide incidence of DM is surging swiftly, positioning it as a leading metabolic disturbance globally [2]. The World Health Organization predicts that diabetes will ascend to the seventh predominant cause of mortality by 2030 [3]. DM presents in four principal forms. Type 1 diabetes mellitus (T1DM) originates as an autoimmune condition, characterized by the destruction of insulin-producing β -cells in the pancreas [4]. The autoimmune origin of T1DM gains validation from the detection of autoantibodies aimed at pancreatic islet cells, their invasion by T-cells, B-cells, and macrophages, and the presence of abnormalities in cellular immunity [5]. Commonly identified during childhood, this diabetes type constitutes 5–10% of all diabetes instances [2].

Type 2 DM (T2DM) stands as the predominant form of diabetes, resulting from inadequate insulin production and / or insensitivity of insulin receptors, leading to the inability of glucose to enter the cells [6]. T2DM accounts for 90–95% of all the DM cases and mainly affects the adults and elderly [7]. T2DM is renowned for its diverse complications, ranging from additional cardiovascular issues like obesity, hypertension, and an atherogenic dyslipidemia profile typified by high levels of triglycerides and low levels of high-density lipoprotein cholesterol among others to neuropathy [7]. Gestational diabetes mellitus (GDM) manifests exclusively during gestation, affecting roughly 5–15% of gravid females, with incidence rates demonstrating variability across ethnic groups and geographical territories [8]. Monogenic diabetes, commonly misclassified as T1DM or T2DM, originates from a mutation in a singular gene or a gene conglomerate [9]. This type of DM is transmitted through an autosomal dominant fashion, affected individuals present with heterogeneous clinical manifestations, symptomatic profiles, and disease trajectories [10].

The etiology of DM on the cellular and molecular scales entails a multifaceted interaction of elements. Hyperglycemia, a hallmark of diabetes, induces increased reactive oxygen species (ROS) levels, precipitating enduring modifications in the structure and functionality of macromolecules like proteins, lipids, and nucleic acids [11]. This oxidative stress (OS) is implicated in the malfunction of endothelial cells, a prevalent condition in diabetic individuals [12]. The compromise of endothelial cells correlates with disturbances in nitric oxide (NO) synthesis, pivotal for maintaining vascular functional equilibrium [13]. The diacylglycerol (DAG)–protein kinase C (PKC) axis is a pivotal pathway implicated in the augmented generation of ROS within endothelial cells [14]. Furthermore, elevated glucose concentrations stimulate PKC, recognized for its substantial involvement in precipitating diabetic endothelial dysfunction [15]. Such activation of PKC results in the enhanced expression of nuclear factor- κ B (NF- κ B) and COX-2, fostering OS and modulating the synthesis of vasoconstrictive prostanoids in diabetes [16]. These Cellular and molecular pathways instigate the emergence of vascular abnormalities correlated with diabetes.

NF- κ B, a primordial protein transcription factor, is instrumental in orchestrating innate immunity, inflammation, oncogenesis, and neural system operations [17]. NF- κ B plays various roles in diabetes, particularly in inflammation and immune regulation [18]. This transcription factor regulates gene expression pertinent to immune and inflammatory reactions, cellular viability, and cellular adhesion [19, 20]. Heightened NF- κ B activity is identified as a pathological element in sustained inflammation seen in autoimmune conditions, including T1DM [21]. Suppression of the NF- κ B signaling cascade has demonstrated efficacy in mitigating cardiomyocyte hypertrophy and fibrosis associated with T1DM [22]. Additionally, NF- κ B significantly influences the development of T2DM [23]. Research indicates that activating NF- κ B can replicate the insulin insensitivity observed in rodents subjected to high-fat diets or obesity [24, 25]. Furthermore, pro-inflammatory cytokines such as TNF- α are linked to enhancing insulin insensitivity by the serine phosphorylation of IRS1 through NF- κ B activation [26]. Moreover, NF- κ B is pivotal in gene expression, including GLUT2, which is pivotal for insulin release from β cells [27]. NF- κ B is crucial in diabetic nephropathy (DN) development. ALTamimia et al. found that NF- κ B contributes to DN progression by triggering intrinsic cell apoptosis, marked by Bax enhancement, Bcl-2 diminution, and cytochrome-c discharge inducement. Excessive NF- κ B activity also incites inflammation, fibrosis, and apoptosis in the renal tissues of diabetic patients [28].

Mitigating DM is essential for lessening its global burden. Growing evidence endorses the use of medicinal plant supplements for DM prevention and management, with curcumin (CUR) receiving notable attention as a promising option [29, 30]. CUR is a natural active substance present in the rhizome of the *Curcuma longa* plant, which is more commonly recognized as turmeric [31]. It exhibits a spectrum of therapeutic properties including antioxidative, cardiac protective, anti-inflammatory, glucose-lowering, and antiarthritic capacities, corroborated through comprehensive *in vitro* and *in vivo* experimental studies [32–34]. In animal studies, CUR extract has demonstrated the ability to postpone diabetes onset, bolster β -cell function, shield β -cells from demise, and lessen insulin resistance [35, 36].

Jiménez-Flores et al. identified curcumin’s capacity to suppress NF- κ B expression in diabetic mice’s livers, highlighting its potential as an anti-inflammatory mediator in the context of diabetes contexts. This points to the therapeutic potential of NF- κ B targeting for managing T2DM’s inflammatory aspects and related

complications [30]. Furthermore, ALTamimi et al. observed that curcumin’s protective impact in reversing diabetic nephropathy (DN) in rats is linked to NF- κ B inhibition, underlining NF- κ B’s role in DN pathophysiology and its viability as a therapeutic target [28]. This study aims to scrutinize the available scientific data and synthesize compelling evidence for curcumin’s potential therapeutic roles in treating diabetic complications, particularly through modulating the NF- κ B pathway. The curcumin’s effects on different types of DM, liver complications tied to diabetes, and its capability to mitigate inflammation, OS and apoptotic cellular demise in tissues affected by diabetes are fully discussed.

2. Chemistry and pharmacokinetics of curcumin

CUR (Figure 1), a molecule with the formula $C_{21}H_{20}O_6$ and the structure ((1E,6E)-1,7-bis(4-hydroxy-3-methoxyphenyl) hepta-1,6-dien-3,5-dione) was initially extracted from turmeric in the year 1815 [37].

Figure 1. Curcumin chemical structure [38].

As a polyphenolic compound from the diarylheptanoid class, CUR is characterized by its symmetrically substituted aromatic rings bearing methoxy groups and phenolic OH groups in the ortho position, linked via a conjugated heptadiene chain featuring an enone segment and a 1,3-diketone structure. The molecule’s bioactivity stems from its functional groups: two ortho-methoxy and two hydroxyl phenolic groups, a duo of double bonds in the aliphatic chain, and the 1,3-keto-enol structure [39].

CUR maintains stability under heat ($<120\text{ }^\circ\text{C}$) and within a pH range of 2.5–6.5, transitioning from yellow in acidic conditions to reddish-brown in alkaline settings. It dissolves well in ethanol, acetone, methanol, and oils, yet exhibits minimal water solubility [40]. Its low bioavailability at 12 g/day in humans is owing to limited intestinal uptake, accelerated hepatic metabolism, and rapid systemic elimination [41]. Predominantly excreted unmetabolized in feces, a minor absorbed fraction undergoes metabolic transformations[42].

Metabolism starts with reductase-driven reduction within enterocytes and hepatocytes, resulting in the synthesis of dihydrocurcumin, tetrahydrocurcumin, hexahydrocurcumin, and octahydrocurcumin [42]. This process is catalyzed by enzymes such as the NADPH-dependent reductase and alcohol dehydrogenase, inclusive of a certain unidentified microsomal enzyme [43]. Hassaninasab’s team identified the curcumin-reducing enzyme in *Escherichia coli*, noting a two-stage reduction to tetrahydrocurcumin via an intermediate, dihydrocurcumin, facilitated by NADPH [44]. *In vivo* and *in vitro*, CUR, along with its metabolites, easily form conjugates with glucuronic acid and sulfate [45, 46]. Glucuronidation and sulfation, primarily in the hepatic and intestinal regions of both rodents and humans, involve glucuronyl transferase and sulfotransferase, respectively, producing glucuronide and sulfate conjugates detectable in human plasma post-oral administration[42]. Human phenol sulfotransferase (SULT1A1, SULT1A3) and uridine diphosphate-glucuronosyltransferase (UGT) facilitate these processes [47].

However, curcumin’s reduction or conjugation generates derivatives with less COX-2 suppression than CUR itself [48]. Tetrahydrocurcumin, hexahydrocurcumin, and CUR sulfate exhibit decreased prostaglandin E2 inhibition, with hexahydrocurcuminol showing no inhibition [49]. Except for tetrahydrocurcumin, these metabolites’ biological activities are significantly reduced compared to CUR [50, 51].

Curcumin’s limited solubility and absorption, coupled with its rapid conversion to inactive metabolites in the gastrointestinal tract, hinder its effectiveness as a health aid and dietary supplement. To counteract this, recent studies have adopted various nanoformulation strategies to boost curcumin’s efficacy. These include employing adjuvants, stabilizers, conjugates/polymer conjugates, lipids/liposomes, and nano-sized hydrogels, microgels, and nanoparticles [52, 53].

3. Pharmacological actions of curcumin

CUR has shown potency against numerous chronic conditions like Alzheimer’s disease, T2DM, rheumatoid arthritis, and metabolic syndrome [54-57]. It possesses a broad spectrum of therapeutic properties, including antineoplastic, antimicrobial, anti-carcinogenic, anti-mutagenic, anti-aging, anti-inflammatory, anti-proliferative, anti-amyloid, and anti-hypercholesterolemic effects [58, 59]. Mechanistically, CUR inhibits the

initiation of the radical-sensitive transcription factor NF- κ B, reduces cytokine production, and impedes vital cellular survival processes [60]. It also inhibits STAT proteins and NF- κ B-DNA binding, diminishing the expression of pro-inflammatory molecules MMP-9, MMP-3, and cytokines like TNF- α , IL-1, and IL-8. Furthermore, CUR attaches to the COX-2 protein, curtailing COX-2 expression and the synthesis of prostaglandins and thromboxanes [61].

Clinical trials have consistently affirmed curcumin's assurance, acceptability, and effectiveness in addressing diverse chronic disorders in humans. These trials reported no toxicity when CUR was given by mouth at a daily dose of 6 g for a duration of 4 to 7 weeks [62-64]. Specific studies, like that of Greil et al., determined the maximum safe dosage of liposomal CUR (Lipocure) for cancer treatment to be 300 mg/m² [65]. Saghatelian et al.'s research on the combined intravenous infusion of CUR and paclitaxel in individuals with progressive breast cancer demonstrated no significant adverse effects or deterioration in life quality after a 12-week regimen [66].

4. Overview of NF- κ B

NF- κ B, identified in 1986 by David Baltimore's team, is a nuclear factor in B cells binding the kappa light chain immunoglobulin enhancer [67, 68]. In its dormant state, NF- κ B is located in the cytoplasm, it becomes active and moves to the nucleus to initiate the transcription of more than 400 genes vital for immunity, growth, apoptosis, and inflammation [68-70]. It modulates a spectrum of crucial genes, encompassing chemokines and pro-inflammatory cytokines, both positively and negatively. Potent activators like IL-1 β and TNF- α induce NF- κ B, which also tempers inflammation, thereby regulating its own activity [68, 71-73].

NF- κ B activation occurs via canonical or alternative pathways [74]. The principal route, crucial for inherent immunity, inflammation, and programmed cell death prevention, is stimulated by factors like TNF- α , IL-1, and LPS [75]. Within this pathway, NF- κ B, primarily a p50 and RelA heterodimer, is held inactive within the cytoplasm by ankyrin repeat-containing inhibitors interacting with NF- κ B's Rel homology domains, with I κ B α being the most common [68, 69, 76]. Activation involves specific I κ B kinase (IKK) triggering, resulting in the phosphorylation, polyubiquitination, and ensuing proteasomal breakdown of I κ B proteins. This reveals a nuclear positioning indicator, permitting the NF- κ B heterodimer to access the nucleus and initiate transcription of target genes [68, 69, 77]. Conversely, the alternative pathway, vital for B-cell maturation, secondary lymphoid organ formation, and high-affinity antibody production, is initiated by NF- κ B-inducing kinase (NIK) [75, 78]. NIK phosphorylates IKK α , which subsequently phosphorylates pre-existing p100/NF- κ B2:RelB heterodimers. This action converts inhibitory p100/NF- κ B2 into p52, allowing the active p52:RelB complexes to enter the nucleus and stimulate target genes [68, 79].

In normal conditions, mechanisms exist to regulate NF- κ B overactivation, ensuring controlled NF- κ B nuclear presence. I κ B α , possessing a nuclear export sequence, can bind NF- κ B and escort it out of the nucleus. Additionally, proinflammatory signals trigger deubiquitinating enzymes like CYLD, A20, and Cezanne, which inhibit IKK activation by removing polyubiquitin chains from I κ B α . This stabilizes newly synthesized I κ B α , thus curtailing further NF- κ B activation [68, 80].

Figure 2. Illustrates the NF- κ B signaling cascade, delineating the canonical pathway on the left and the alternative pathway on the right, with respective agonists listed.

Figure 2. In the canonical pathway, the NF- κ B heterodimer, typically consisting of p50 and RelA subunits, plays a pivotal role in innate immunity, inflammation, and apoptosis prevention. Residing in the cytoplasm, this heterodimer is inactivated by an inhibitory molecule, commonly I κ B α . Activation occurs when a specific I κ B kinase (IKK) phosphorylates two conserved serine residues on the N-terminal domains of I κ B proteins, leading to their polyubiquitination and proteasomal degradation. This reveals a nuclear localization signal, permitting NF- κ B's migration into the nucleus to trigger target gene transcription. In the alternative pathway, NF- κ B-inducing kinase phosphorylates inhibitory IKK α , which in turn phosphorylates preexisting p100/NF- κ B2:RelB heterodimers. This process triggers the processing of inhibitory p100/NF- κ B2 to p52. The active p52:RelB complexes can then translocate and activate downstream target genes. This pathway is crucial for appropriate B-cell maturation, formation of lymphoid organs, and the production of high-affinity

antibodies. The response element (RE) serves as a binding site for active NF- κ B.

5. Εμφραγμα οφ ὑπερλιπιδαιμιας ον Τ1ΔΜ ανδ Τ2ΔΜ: Φορσος ον ΝΦ- κ B πατηωαψ

T1DM, primarily identified in children and young adults, is characterized by the body's inability to synthesize and secrete insulin, necessitating lifelong insulin treatment [81, 82]. Streptozotocin (STZ) is a preferred agent to induce experimental T1DM in animals [83], as it selectively destroys insulin-generating beta cells in the pancreas, mimicking T1DM characteristics [4, 84]. Curcumin's beneficial impact on hyperglycemia in STZ-induced diabetic rats is widely reported, yet its underlying mechanisms require further exploration [85]. It's hypothesized that curcumin's cholesterol-lowering, antioxidant properties, and its ability to elevate plasma insulin levels contribute to its potential in managing metabolic syndrome, obesity, and diabetes-related issues [86-88].

Diabetic cardiomyopathy (DCM), a grave complication of diabetes, notably in T1DM, [89] involves heart structure and function alterations, leading to compromised cardiac performance [90]. Despite being a major mortality cause in diabetic patients, DCM lacks specific treatments due to its complex pathogenesis [91]. It's occurrence in T1DM patients is characterized by disrupted lipid metabolism, increased cardiac injury markers, and escalated OS [89]. JM-2, a CUR derivative, exhibited therapeutic benefit in combating DCM in mouse models of STZ-induced T1DM and diet-induced T2DM [92]. Wang et al. reported that JM-2 combats cardiac functional and structural deficits in both T1DM and T2DM models. JM-2 administration curtailed cardiac dysfunction, inflammation, fibrosis, and inhibited the NF- κ B pathway in diabetic mouse hearts. Particularly, JM-2 eradicated STZ-induced cardiac issues in T1DM mice, forestalled heart fibrosis in T2DM mice, and mitigated hypertrophy and fibrosis in high glucose-exposed cardiac cells, owing to its NF- κ B-mediated anti-inflammatory action [92]. These insights bolster the therapeutic potential of JM-2 for DCM and accentuate the necessity of novel drug development for this condition, positioning JM-2 as a promising candidate for DCM treatment.

CaMKII, a serine/threonine kinase, is pivotal in cardiac pathologies like DCM, regulating cardiac hypertrophy genes, amplifying pro-inflammatory signaling, elevating inflammatory cytokine expression, and modulating cardiac fibroblast growth and collagen production [93, 94]. Its heightened activity in diabetic hearts contributes to pathological cardiac remodeling [95]. PPAR- γ , a nuclear hormone receptor, is instrumental in controlling lipid and glucose metabolism genes [96]. Clinically utilized for T2DM treatment, its activation can ameliorate cardiovascular disorders including DCM, by optimizing myocardial lipid profiles, mitigating endoplasmic reticulum stress, curbing inflammation, and diminishing ROS [97]. PPAR- γ also suppresses myocardial pro-inflammatory markers like NF- κ B and TGF- β 1 [97, 98]. TGF- β 1, a multifunctional cytokine, plays a crucial role in cellular proliferation, differentiation, and immune regulation [99]. It significantly influences myocardial fibrosis in DCM by regulating fibroblast function and extracellular matrix accumulation [100]. Additionally, its link to OS and inflammation highlights its involvement in the onset of cardiac fibrosis [98, 101].

A research conducted by Gbr et al. investigated the cardioprotective attributes of pioglitazone (a PPAR- γ agonist) and CUR in DCM within a T1DM rat model. The findings indicated notable reductions in heart weight, blood glucose levels, lipid and cardiac injury markers, OS, and fibrosis following pioglitazone and CUR treatment. The duo therapy notably outperformed individual treatments, with observed cardioprotective effects tied to modulations in CaMKII, PPAR- γ , NF- κ B, and TGF- β 1 pathways. Histological analyses corroborated the cardiac structural improvements in the treated groups, suggesting pioglitazone and curcumin's potential in mitigating DCM in T1DM via these signaling pathways [98].

COX-2, an integral enzyme in prostaglandin synthesis, plays a key role in inflammation, pain, and fever. Often upregulated by inflammatory triggers, COX-2 is linked to prostanoid production, like TXA2 and PGI2 which are crucial in vascular tone and inflammation regulation [102]. NF- κ B p65, a component of the NF- κ B transcription factor, is central in regulating genes pertinent to inflammation, immunity, cell proliferation, and survival. Typically bound to inhibitory proteins in the cytoplasm, it relocates to the nucleus upon activation by stress or inflammation, initiating the expression of related genes [68, 103]. Protein kinase C

(PKC) represents a family of enzymes critical in cellular functions including proliferation, differentiation, and programmed cell death [104].

Rungseesantivanon and colleagues showed that daily supplementation of CUR at 30 and 300 mg/kg notably reduced blood glucose in T1DM rats by 18.73% and 30.26%, respectively. The research also highlighted curcumin's capacity to balance the prostanoid ratio in diabetic mesenteric arteries and decrease average arterial blood pressure. Importantly, CUR therapy significantly diminished COX-2 and NF- κ B p65 expression in the small mesenteric arterial walls of the rats. Emphasizing PKC's contribution to diabetic vascular complications, the study indicated that elevated glucose levels trigger PKC, resulting in increased ROS production and raised NF- κ B p65 and COX-2 expressions, elements linked with OS and modified vasoconstrictor prostanoid production in diabetes. These observations endorse curcumin's potential in alleviating diabetes-induced endothelial dysfunction through its antioxidant and anti-inflammatory characteristics [16].

In another study, Zhang et al. explored impacts of CUR on diabetes-induced lung injury in rats. Their findings indicated that CUR not only diminished blood glucose, triglycerides, and cholesterol levels in diabetic rats but also diminished lung tissue inflammation. CUR significantly decreased Pro-inflammatory mediators like TNF- α , IL-1 β , and IL-6 in pulmonary structures. Furthermore, CUR lessened NO and PGE2 levels, crucial inflammatory response effectors during tissue injury, and curtailed inflammatory mediators and enzymes like iNOS and COX-2 in the pulmonary structures. The research also highlighted curcumin's role in reducing OS, evidenced by decreased MDA and MPO levels, and enhancing the antioxidant system, indicated by increased SOD and reduced GSH levels. These results underscore curcumin's potential in ameliorating diabetic lung injury through its antioxidative and counter-inflammatory responses, particularly by inhibiting the NF- κ B pathway, thus mitigating pulmonary inflammation and OS in the STZ-diabetic rats [105].

C66, a novel derivative of curcumin, has garnered attention for its potential therapeutic properties [106]. Exhibiting anti-inflammatory effects, it curtails high glucose-provoked inflammatory reactions within both laboratory conditions and living organisms, effectively mitigating renal damage in diabetic rat models through its Counter-inflammatory effects [107]. TNF- α , a pivotal pro-inflammatory cytokine, is integral to immune regulation, inflammation, and apoptosis, primarily produced by cells like macrophages [108, 109]. Pan et al. revealed that C66 diminishes NO and TNF- α production in macrophages under high glucose conditions. Moreover, C66 suppresses mRNA transcription of pro-inflammatory cytokines (IL-1 β , TNF- α , IL-6, IL-12) and inducible enzymes (COX-2, iNOS), offering renal protection. This is evident from the amelioration of histopathological anomalies and fibrosis in diabetic kidneys, marked by decreased glycogen deposition, type IV collagen levels, serum creatinine, and kidney/body weight ratio. Additionally, C66 treatment has been found to result in the dephosphorylation of JNK and inhibition of the activation of NF- κ B. This inhibition is associated with the suppression of I κ Ba phosphorylation and degradation, as well as the nuclear translocation of the p65 subunit of NF- κ B. Notably, these therapeutic benefits of C66 manifest without altering blood glucose or body weight, underscoring its promise as a diabetic nephropathy (DN) treatment by targeting inflammation and renal damage [29].

Inflammation is pivotal in both the onset and advancement of T1DM [110]. The progression of T1DM results in the migration of inflammatory cells into pancreatic islets, which triggers the release of proinflammatory cytokines, notably TNF- α , IFN- γ , and IL-1 β [111, 112]. These cytokines initiate the degradation of pancreatic β cells by activating intracellular signaling that favors apoptotic pathways [113, 114]. Excessive cytokine release activates NF- κ B in β cells, further inducing the transcription of proinflammatory cytokines [115]. The existence of T cells that produce IFN- γ in pancreatic islets marks an early sign of T1DM [116]. Elevated levels of cytokines like IL-1 β and IL-6 correlate with hampered insulin secretion, diminished β cell multiplication, and heightened cell death, underscoring the vital role of inflammation in β cell destruction and T1DM pathogenesis [117].

Rashid et al. explored the impact of CUR on diabetes-induced OS and related spleen complications. Key findings include: 1) CUR successfully reduced blood glucose, normalized serum insulin levels, and enhanced body weight in STZ-diabetic rats, 2) it countered the heightened expression of inflammatory cytokines (TNF- α ,

IL-1 β , IL-6) and chemokines (MCP-1), showcasing its anti-inflammatory role in ameliorating hyperglycemia-driven splenic damage, 3) CUR modulated the detrimental impacts of the NF- κ B-mediated inflammatory cascade in diabetic spleen tissue, 4) it altered ER-dependent apoptotic proteins and decreased intracellular Ca²⁺ in the spleen, and 5) CUR significantly raised the GSH/GSSG ratio, indicating its efficacy in attenuating OS in the spleen. Overall, the study posits CUR as a promising agent against diabetes-induced OS, inflammation, and apoptosis in the spleen [118].

In a study by Badr et al., the influence of CUR on STZ-diabetic mice was assessed. Findings reported significant blood glucose reduction. The study underscored the role of NF- κ B in T1DM pathogenesis, particularly in β cell dysfunction and apoptosis, driven by cytokines like IL-1 β and IFN- γ . They identified phosphorylated p65 as a marker of NF- κ B activation. CUR administration, with its anti-inflammatory properties, inhibited NF- κ B activation and curtailed the expression of proinflammatory cytokines (TNF- α , IL-1, IL-6). These outcomes highlight NF- κ B's involvement in diabetic inflammation and suggest curcumin's therapeutic potential in modulating these responses [119]. In another study, Mojtabavi et al. highlighted that CUR not only reduced blood glucose levels but also demonstrated significant analgesic properties, likely due to its influence on diminishing the expression of inflammatory genes like NF-B, IL6, and TNF- α . Notably, in neuropathy assessments, both CUR and metformin treatment groups exhibited significant improvements compared to the diabetic control group [120].

Inflammation plays a crucial role in insulin resistance, especially in obesity and T2DM contexts [121]. Elevated levels of pro-inflammatory cytokines like TNF- α and IL-6 in these states disrupt insulin signaling, particularly in liver, muscle, and adipose tissues [122]. These cytokines play a critical role in the development of insulin resistance and can impair insulin signaling pathways [122]. These cytokines impede insulin signaling molecules like IRS-1 and GLUT4, leading to reduced insulin receptor signaling and increased insulin resistance [123, 124]. Addressing this inflammatory cascade is thus crucial in managing insulin resistance-related conditions.

3T3-L1 adipocytes, derived from mouse embryonic fibroblasts, are key in adipose and obesity research due to their role in adipocyte differentiation, lipid metabolism, and inflammation studies [125, 126]. Wang et al. reported curcumin's efficacy in countering palmitate-induced insulin resistance in these cells by enhancing insulin-stimulated glucose uptake, blocking NF-B p65 nuclear translocation, and reducing MAPKs activity. The spice also reversed the pro-inflammatory state induced by palmitate, downregulating TNF- α and IL-6 expression and decreasing JNK, ERK1/2, and p38MAPK activities dose-dependently, suggesting its potential in managing obesity-related insulin resistance and inflammation [127].

AMPK, a vital enzyme in cellular energy regulation, is activated during low energy states to stimulate energy production and inhibit energy consumption, thereby managing glucose and fatty acid metabolism [128-131]. Jiménez-Flores et al. explored curcumin's impact on diabetic db/db mice livers, noting increased AMPK and PPAR γ expression and reduced NF- κ B protein levels post-treatment. These results point to curcumin's therapeutic promise in T2DM by modulating inflammatory pathways and metabolic dysfunctions, as evidenced by its ability to significantly lower NF- κ B expression, indicating its anti-inflammatory potential in diabetic contexts. Therefore, the findings of this study support the potential of CUR as a modulator of NF- κ B expression, indicating its possible role in managing the inflammatory response associated with T2DM [30].

6. Εμφραξτε οφ ὕρςυμιν ον διαβετις νεπεροπατηψ ιν ΣΤΖ-ινδυσεδ διαβετις ανιμαλ μονελ: φοσυς ον ΝΦ-κΒ πατηωαψ

Diabetic nephropathy (DN), also termed diabetic kidney disease, is a grave complication of both type 1 and type 2 diabetes, impairing the kidneys' capacity to eliminate waste and excess fluids from the body [132]. It stems from elevated blood sugar levels and hypertension, which may harm the renal blood vessels responsible for filtering waste [133]. Prolonged DN can result into chronic kidney disease culminating in kidney failure.

Inflammation is pivotal in the onset and advancement of DN. Inflammation contributes to DN's pathology at multiple junctures, including the escalation of chemokine production, influx of inflammatory cells into

the kidney, synthesis of pro-inflammatory cytokines, and resultant tissue damage. In diabetic individuals, inflammatory markers such as chemokines (MCP-1), pro-inflammatory cytokines, and cell adhesion molecules intensify in the renal tissues. Moreover, the concentration of these cytokines and cell adhesion molecules in the serum and urine correlates with albuminuria [134]. The modulation of inflammatory signaling pathways has gained attention as a prospective therapeutic avenue for DN. Numerous investigations indicate that anti-inflammatory compounds, like CUR, may mitigate diabetic renal injury and avert kidney deterioration by curbing inflammation [134-136].

NF- κ B orchestrates the expression of genes pivotal in renal disease progression, including chemokines like MCP-1. In DN, MCP-1 is instrumental in drawing monocytes/macrophages, and its elevated presence in human DN tubulointerstitial lesions implies its role in advanced DN pathogenesis. In DN, blocking NF- κ B activation has emerged as a viable therapeutic approach, curtailing gene transcription and impeding the inflammatory cascade. Numerous investigations reveal that persistent suppression of NF- κ B mitigates kidney damage in DN models [137-140]. MCP-1, a pivotal chemokine, orchestrates the recruitment and congregation of monocytes and macrophages in various inflammatory scenarios, DN included [141]. ICAM-1, a 90-kD membrane glycoprotein, crucially moderates interactions with immune cells and is notably elevated at inflammation sites [142].

Soetikno and colleagues demonstrated that CUR therapy mitigated DN in a T1DM rat model, achieved by curtailing macrophage penetration in the glomerulus via its anti-inflammatory properties. Furthermore, CUR hindered NF- κ B activity, diminishing the release of proinflammatory and profibrotic cytokines. The study also highlighted curcumin's ability to significantly reduce blood glucose and 24-hour urinary protein, alongside mitigating weight loss and enhancing DN-related biochemical indicators like plasma creatinine, blood urea nitrogen, and creatinine clearance. Histological analysis showed CUR reduced glomerular sclerosis and macrophage presence in diabetic rat kidneys. Moreover, CUR suppressed proinflammatory proteins such as ICAM-1, TGF- β 1, and MCP-1, underscoring its potential as an adjunct therapy in DN prevention [143].

Oxidative stress (OS) is central in DN pathogenesis, predominantly fueled by excessive ROS generation due to hyperglycemia, resulting in oxidative damage, inflammation, and fibrosis in the renal system [144]. Mitochondria-derived ROS are pinpointed as key catalysts in DN onset and progression [145]. Diabetic individuals face additional ROS sources, including NADPH oxidase activation and eNOS uncoupling [146]. Notably, hindering mitochondrial ROS production via antioxidant agents or transgenic antioxidant expression has shown efficacy in curbing DN and other microvascular complications [147].

The PKC β /p66Shc pathway involves PKC β II and p66Shc [28]. Stress conditions like hyperglycemia, hydrogen peroxide (H₂O₂), and UV exposure lead to PKC β II and JNK-mediated Ser36 phosphorylation of p66Shc in the cytoplasm [148]. This phosphorylation is essential for p66Shc's mitochondrial migration, disrupting the electron transport chain, cytochrome-c release, and MPTP opening [149]. These actions trigger extensive mitochondrial ROS production and activate intrinsic cell death mechanisms [28]. The PKC β /p66Shc axis is implicated in DN's evolution and is a prospective therapeutic target for the disorder [28]. FOXO-3a, a key transcription factor, regulates a spectrum of cellular functions, including OS resistance, apoptosis, cell cycle regulation, and metabolism [150, 151]. It belongs to the FOXO transcription factor family, recognized for regulating genes tied to cellular longevity and survival [152]. Predominantly situated in the nucleus, FOXO-3a modulates the expression of genes linked to antioxidant defense, DNA repair, and cell cycle arrest [153].

ALTamimi and colleagues underscored curcumin's renal protective role in STZ-induced diabetic rats, primarily through thwarting the PKC β /p66Shc pathway and invigorating FOXO-3a. They observed curcumin's reversal of DN symptoms, encompassing proteinuria, glomerular and tubular deterioration, and interstitial fibrosis. CUR showcased its therapeutic prowess by mitigating OS, inflammation, and fibrosis, thus exhibiting antioxidant, anti-inflammatory, and anti-fibrotic properties. It also safeguarded mitochondrial integrity, obstructed the mitochondrial permeability transition pore, and enhanced mitochondrial function indicators. Notably, CUR impeded NF- κ B in the renal tissues of both normal and T1DM-induced rats, markedly diminishing TNF- α and IL-6 levels, as well as NF- κ B P65 nuclear presence and activity in T1DM

+ CUR -treated rats compared to their untreated T1DM counterparts. Further, CUR was found to inhibit cytochrome-c release and downregulate mRNA and protein expressions of collagen I/III in the renal tubules and mitochondria. These insights vindicate curcumin's potential as an efficacious therapeutic agent in DN management [28].

ECM proteins are a set of structural proteins secreted by cells, forming an intricate matrix in the extracellular space [154]. They bolster structural integrity for adjacent cells and partake in various cellular processes including adhesion, migration, differentiation, and proliferation [155, 156].

Chiu et al. observed a halt in the diabetes-prompted surge of vasoactive factors (eNOS and endothelin-1), transforming growth factor- β -1, and ECM proteins (fibronectin and extradomain-B-containing fibronectin) within the kidneys of diabetic rats treated with curcumin. These alterations correlated with escalated OS, mesangial proliferation, and heightened activity of p300 and NF- κ B, all of which were subdued by CUR intervention. Furthermore, CUR was shown to diminish OS and avert structural kidney damage linked to DN. The curative impacts of CUR were primarily through the suppression of p300 and NF- κ B pathways. Conclusively, the research posited CUR as a promising agent for patients grappling with chronic diabetic complications, especially for the prevention of diabetes-induced renal anomalies [157].

7. Εμφραγςτς οφ ύρςυμιν ον οττερ ςομπλινςατιονς οφ διαβητες: Φοςυς ον ΝΦ-κΒ παττωαψ

Diabetic osteoporosis (DOP), a diabetes complication, is marked by weakened bone microarchitecture and reduced bone mineral density (BMD) due to elevated glucose levels [158]. Individuals with DOP are significantly more susceptible to fractures, leading to increased disability and mortality [159]. The disorder is linked to hampered bone regeneration and remodeling, along with reduced osteogenic differentiation and angiogenesis capabilities of bone marrow mesenchymal stem cells (BMSCs) [160]. Restoring the compromised osteogenesis and angiogenesis functions of BMSCs is pivotal for treating DOP [161]. Both types 1 and 2 diabetes are associated with an increased risk of osteoporosis and fractures [162]. BMSCs are adult stem cells located in the bone marrow, capable of transforming into various cell types, including osteoblasts (bone-forming cells), chondrocytes (cartilage-forming cells), and adipocytes (fat-storing cells)[163]. NF- κ B, a transcription factor, regulates genes involved in inflammation, bone resorption, and bone formation [164]. Inhibiting NF- κ B activation has shown promise in enhancing bone formation and alleviating osteopenia in osteoporosis animal models [165]. NF- κ B activation is known to exert an anti-anabolic effect on bone formation, suggesting that NF- κ B inhibitors might serve as anabolic agents in bone health [166]. Fan et al. noted that CUR might reverse the hindered osteogenic differentiation and proangiogenic capability of BMSCs in hyperglycemic environments. Their studies reveal that CUR fosters bone reconstruction and vascular development in a DOP mouse framework, highlighting its therapeutic potential for diabetic osteoporosis. The research also suggests CUR enhances BMSC-driven bone formation and angiogenesis in high glucose scenarios, possibly by suppressing the hyperactive NF- κ B signaling. These insights propose curcumin's effectiveness in counteracting DOP by advancing bone reconstruction and vascular growth [160].

Diabetic gastroparesis, marked by sluggish gastric evacuation in diabetes mellitus patients [167], is a prevalent complication, impacting 30-50% of individuals with type 1 or type 2 diabetes [168, 169]. Symptoms include premature fullness, weight reduction, stomach swelling, discomfort, nausea, and vomiting [170]. This condition disrupts glycemic regulation and severely affects life quality [171]. Diabetic gastroparesis's origin involves OS, inflammation, and loss of gastric ICCs crucial for modulating gastric movement [172, 173]. ICCs, specialized cells in the gastrointestinal system, regulate gut motility and coordinate smooth muscle contractions [174]. They generate and propagate electrical slow waves, crucial for the digestive system's rhythmic contractions and regulating the digestive tract's food passage [175]. ICC dysfunction or loss is linked to various gastrointestinal motility disorders, including diabetic gastroparesis [176, 177]. SCF, a growth factor, is vital for ICC development and maintenance, while c-kit serves as a receptor for SCF [178, 179]. SCF and c-kit signaling are essential for ICC survival, differentiation, and function, crucial for regulating gastric movement and emptying [180].

Jin et al. explored curcumin's impact on diabetic gastric motility in rats, noting improvements in gastric

emptying rates, diminished OS, inhibited NF- κ B activation, and increased SCF/c-kit expression in diabetic rat stomach tissues. Curcumin's protective role on ICCs underscores its potential therapeutic application in diabetic gastroparesis. The study also found curcumin's antioxidative and free radical scavenging properties and its ability to restore SCF/Kit protein levels in diabetic rats. Curcumin's supplementation mitigated OS and NF- κ B activation, hinting at its protective potential against diabetic gastroparesis. CUR decreased MDA levels, increased SOD activity, inhibited ROS formation, and prevented ROS-induced apoptosis in diabetic rat stomach tissues, demonstrating its beneficial effects on gastric motility and OS [181].

Table 1 and Figure 3 briefly show curcumin effects on NF- κ B pathway in diabetes mellitus.

Table 1. Curcumin effects on NF- κ B pathway in diabetes mellitus

Figure 3. Targeting of the NF- κ B pathway by curcumin in diabetes mellitus: Focusing on oxidative stress and inflammation

Conclusion

The study underscores curcumin's potential as a therapeutic agent in mitigating DM complications, primarily through modulating the NF- κ B pathway. By demonstrating anti-inflammatory, antioxidative, and anti-fibrotic properties, CUR has been effective in managing multiple diabetic conditions, including nephropathy, cardiomyopathy, and gastroparesis. The alteration of critical signaling pathways such as CaMKII, PPAR- γ , NF- κ B, and TGF- β 1 is pivotal in curcumin's beneficial effects. Additionally, its capacity to diminish OS, curb inflammation, and enhance vascular functionality vindicate CUR as a promising candidate molecule for management of DM complication. These findings highlight the necessity to further explore the potential of CUR as a supplementary treatment option in management of DM complications.

Author Contributions Statement

M.Y.Z & M.G: Writing – review & editing, Validation. A.S, H.O.A & A.Y: Writing – original draft, Conceptualization. S.B.O & A.S: Writing – review & editing, funding acquisition. F.K: Validation. A.M.J & M.K.A: Visualization, Methodology. A.J & A.H.A: Methodology. M.Y.Z & A.S: Supervision and Conceptualization. N.S.J & F.K: Data curation and Software.

Acknowledgments

Not applicable

Declaration of conflicting interests

The authors declare no conflict of interest.

Funding

Not applicable

Ethical statement

Not applicable

Data availability statement

Not applicable

References

1. Atif, M., J. Siddiqui, and F. Talib. *An overview of diabetes mellitus prediction through machine learning approaches*. in *2019 6th International Conference on Computing for Sustainable Global Development (INDIACom)*. 2019. IEEE.
2. Alam, S., et al., *Diabetes Mellitus: insights from epidemiology, biochemistry, risk factors, diagnosis, complications and comprehensive management*. *Diabetology*, 2021. **2** (2): p. 36-

50.

3. Cd, M., *Projections of global mortality and burden of disease from 2002 to 2030*. PLoS Med, 2006. **3** : p. 2011-2030.
4. Zhu, B.T., *Παθογενής μεσηνιασμός οφ αυτοϊμμυνοε διαβητες μελλιτυς ιν ηυμανς: ποτεντιαλ ρολε οφ στρεπτοζοτοσιν-ινδυσεδ σελεςτιε αυτοϊμμυνοιψ αγαινιστ ηυμαν ιολετ β-σελλς*. Cells, 2022. **11** (3): p. 492.
5. Li, Z., et al., *Elevated glucose metabolism driving pro-inflammatory response in B cells contributes to the progression of type 1 diabetes*. Clinical Immunology, 2023. **255** : p. 109729.
6. Hameed, I., et al., *Type 2 diabetes mellitus: from a metabolic disorder to an inflammatory condition*. World journal of diabetes, 2015.**6** (4): p. 598.
7. Zheng, Y., S.H. Ley, and F.B. Hu, *Global aetiology and epidemiology of type 2 diabetes mellitus and its complications*. Nature reviews endocrinology, 2018. **14** (2): p. 88-98.
8. Ben-Haroush, A., Y. Yogev, and M. Hod, *Epidemiology of gestational diabetes mellitus and its association with Type 2 diabetes*. Diabetic Medicine, 2004. **21** (2): p. 103-113.
9. Kearney, P., et al., *Efficacy of cholesterol-lowering therapy in 18,686 people with diabetes in 14 randomised trials of statins: a meta-analysis*. Lancet (London, England), 2008. **371** (9607): p. 117-125.
10. Misra, S. and K.R. Owen, *Genetics of monogenic diabetes: present clinical challenges*. Current diabetes reports, 2018.**18** : p. 1-11.
11. Rendra, E., et al., *Reactive oxygen species (ROS) in macrophage activation and function in diabetes*. Immunobiology, 2019.**224** (2): p. 242-253.
12. Maruhashi, T. and Y. Higashi, *Pathophysiological association between diabetes mellitus and endothelial dysfunction*. Antioxidants, 2021. **10** (8): p. 1306.
13. Kolluru, G.K., S.C. Bir, and C.G. Kevil, *Endothelial dysfunction and diabetes: effects on angiogenesis, vascular remodeling, and wound healing*. International journal of vascular medicine, 2012.**2012** .
14. Volpe, C.M.O., et al., *Cellular death, reactive oxygen species (ROS) and diabetic complications*. Cell death & disease, 2018.**9** (2): p. 119.
15. Pricci, F., et al., *Oxidative stress in diabetes-induced endothelial dysfunction involvement of nitric oxide and protein kinase C*. Free Radical Biology and Medicine, 2003. **35** (6): p. 683-694.
16. Rungseesantivanon, S., et al., *Έρςυμιν ιμπροες προστανοιδ ρατιο ιν διαβητις μεσευτερις αρτεριες ασσοσιατεδ ωπη ςψςλοοξψγενασε-2 ανδ ΝΦ-κΒ συππρεσσιον*. Diabetes, metabolic syndrome and obesity: targets and therapy, 2010: p. 421-429.
17. Albensi, B.C., *Ωηατ ις νυςλεαρ φαστορ καππα Β (ΝΦ-κΒ) δοιγγ ιν ανδ το τηε μιτοςηονδριον*; Frontiers in cell and developmental biology, 2019. **7** : p. 154.
18. Liu, T., et al., *ΝΦ-κΒ σιγγαλιγγ ιν ινφλαμματαιον*. Signal transduction and targeted therapy, 2017. **2** (1): p. 1-9.
19. Huang, D., et al., *Φοσαλ αδηρεσιον κινασε μεδιατες σελλ συριαλ ια ΝΦ-κΒ ανδ ΕΡΚ σιγγαλιγγ πατηωαψς*. American Journal of Physiology-Cell Physiology, 2007. **292** (4): p. C1339-C1352.
20. Zhong, L., M.J. Simard, and J. Huot, *Endothelial microRNAs regulating the NF- κ B pathway and cell adhesion molecules during inflammation*. The FASEB Journal, 2018. **32** (8): p. 4070-4084.
21. Mollah, Z.U., et al., *Αβνορμαλ ΝΦ-κΒ φυνςτιον ςηαραςτεριζες ηυμαν τψπε 1 διαβητες δευδριτις σελλς ανδ μονοςψτες*. The Journal of Immunology, 2008. **180** (5): p. 3166-3175.

22. Liu, J.-j., et al., *Ινhibιτιον οφ ΝΦ-κΒ ανδ Ωντ/β-ζατενιν/ΓΣΚ3β σιγναλινγ πατηωαψς αμειορατες ζαρδιομοψοψτε ηψπερτροπηψ ανδ φιβροοις ιν στρεπτοζοτοσιν (ΣΤΖ)-ινδυσεδ τψπε 1 διαβετις ρατς*. Current Medical Science, 2020. **40** : p. 35-47.
23. Akash, M.S.H., K. Rehman, and S. Chen, *Role of inflammatory mechanisms in pathogenesis of type 2 diabetes mellitus*. Journal of cellular biochemistry, 2013. **114** (3): p. 525-531.
24. Zhou, J., et al., *ἀταλπολ αμειορατες ηγη-φατ διετ-ινδυσεδ ινσυλιν ρεοιστανζε ανδ αδιοοσε τιοσυε ινφλαμματοιον βψ συππρεοσινγ τηε ΘΝΚ ανδ ΝΦ-κΒ πατηωαψς*. Biochemical and Biophysical Research Communications, 2015. **467** (4): p. 853-858.
25. Wunderlich, F.T., et al., *Ηεπατις ΝΦ-κΒ εοοοεντιαλ μοδυλατορ δεφιοιενψψ πρεεντς οβεοιτψ-ινδυσεδ ινσυλιν ρεοιστανζε βυτ οψνεργιζεο ωιτη ηγη-φατ φεεδινγ ιν τυμορηγενεοις*. Proceedings of the national academy of sciences, 2008. **105** (4): p. 1297-1302.
26. Hayden, M.S. and S. Ghosh. *Πεγυλατιον οφ ΝΦ-κΒ βψ ΤΝΦ φαμιλψ ζψτοκινεο* . in *Seminars in immunology* . 2014. Elsevier.
27. Ganesan, K., K.M. Ramkumar, and B. Xu, *ϊτεξιν ρεοοτορεο πανζρεατιο β-ζελλ φυνζτιον ανδ ινσυλιν σιγναλινγ τηρουγη Νρφ2 ανδ ΝΦ-κΒ σιγναλινγ πατηωαψς*. European Journal of Pharmacology, 2020.**888** : p. 173606.
28. ALTamimi, J.Z., et al., *υρζυμιν ρεεοοεοο διαβετιο νεπηροπατηψ ιν στρεπτοζοτοσιν-ινδυσεδ διαβετεο ιν ρατς βψ ινhibιτιον οφ ΠΚ^β/π66Σηο αξιο ανδ αοτιατιον οφ ΦΟΕΟ-3α*. The Journal of nutritional biochemistry, 2021. **87** : p. 108515.
29. Pan, Y., et al., *Inhibition of high glucose-induced inflammatory response and macrophage infiltration by a novel curcumin derivative prevents renal injury in diabetic rats*. British journal of pharmacology, 2012. **166** (3): p. 1169-1182.
30. Jiménez-Flores, L.M., et al., *Α ΠΠΑΡγ, ΝΦ-κΒ ανδ ΑΜΠΚ-δεπενδεντ μεζηανιομ μαψ βε ινολεδ ιν τηε βενεφιοιαλ εφφεοις οφ ζυρζυμιν ιν τηε διαβετιο δβ/δβ μιζε λιερ*. Molecules, 2014.**19** (6): p. 8289-8302.
31. Singh, K., et al., *Impact of green extraction on curcuminoid content, antioxidant activities and anti-cancer efficiency (in vitro) from turmeric rhizomes (Curcuma longa L.)*. Foods, 2022.**11** (22): p. 3633.
32. Moutabian, H., et al., *The cardioprotective effects of nano-curcumin against doxorubicin-induced cardiotoxicity: A systematic review*. Biofactors, 2022. **48** (3): p. 597-610.
33. Shen, W., et al., *Chitosan nanoparticles embedded with curcumin and its application in pork antioxidant edible coating*.International Journal of Biological Macromolecules, 2022. **204** : p. 410-418.
34. Saifi, B., et al., *An overview of the therapeutic effects of curcumin in reproductive disorders with a focus on the antiinflammatory and immunomodulatory activities*. Phytotherapy Research, 2022.**36** (2): p. 808-823.
35. Meng, B., J. Li, and H. Cao, *Antioxidant and antiinflammatory activities of curcumin on diabetes mellitus and its complications*.Current pharmaceutical design, 2013. **19** (11): p. 2101-2113.
36. Weisberg, S.P., R. Leibel, and D.V. Tortoriello, *Dietary curcumin significantly improves obesity-associated inflammation and diabetes in mouse models of diabetes*. Endocrinology, 2008.**149** (7): p. 3549-3558.
37. Oh, D.-Y. and H.-S. Kim, *Physicochemical properties of the turmeric (Curcuma longa L.) in Jindo Korea*. Journal of Environmental Science International, 2019. **28** (4): p. 403-412.
38. Priyadarsini, K.I., *The chemistry of curcumin: from extraction to therapeutic agent*. Molecules, 2014. **19** (12): p. 20091-20112.

39. Yang, H., et al., *Structure–activity relationship of curcumin: Role of the methoxy group in anti-inflammatory and anticarcinogenic effects of curcumin*. Journal of agricultural and food chemistry, 2017.**65** (22): p. 4509-4515.
40. Joe, B., M. Vijaykumar, and B. Lokesh, *Biological properties of curcumin-cellular and molecular mechanisms of action*. Critical reviews in food science and nutrition, 2004. **44** (2): p. 97-111.
41. Prasad, S., A.K. Tyagi, and B.B. Aggarwal, *Recent developments in delivery, bioavailability, absorption and metabolism of curcumin: the golden pigment from golden spice*. Cancer research and treatment: official journal of Korean Cancer Association, 2014. **46** (1): p. 2-18.
42. Dei Cas, M. and R. Ghidoni, *Dietary curcumin: correlation between bioavailability and health potential*. Nutrients, 2019.**11** (9): p. 2147.
43. Kotha, R.R. and D.L. Luthria, *Curcumin: biological, pharmaceutical, nutraceutical, and analytical aspects*. Molecules, 2019.**24** (16): p. 2930.
44. Hassaninasab, A., et al., *Discovery of the curcumin metabolic pathway involving a unique enzyme in an intestinal microorganism*. Proceedings of the National Academy of Sciences, 2011. **108** (16): p. 6615-6620.
45. Pandey, A., et al., *Reductive metabolites of curcumin and their therapeutic effects*. Heliyon, 2020. **6** (11).
46. Ireson, C.R., et al., *Metabolism of the cancer chemopreventive agent curcumin in human and rat intestine*. Cancer Epidemiology Biomarkers & Prevention, 2002. **11** (1): p. 105-111.
47. Heger, M., et al., *The molecular basis for the pharmacokinetics and pharmacodynamics of curcumin and its metabolites in relation to cancer*. Pharmacological reviews, 2014. **66** (1): p. 222-307.
48. Bengmark, S., *Ψρυσμιν, Αντοξισ αντοξιδαντ ανδ νατυραλ ΝφκΒ, ζψςλοοξψγενασε-2, λιποοξψγενασε, ανδ ινδυσβλε νιτρις οξιδε σφντσηασε ινηβιτορ: Α σφιελδ αγαϊνιστ αςυτε ανδ σφηρονις δισεασες*. Θουρνάλ οφ Παρεντεραλ ανδ Εντεραλ Νυτριτιον, 2006. **30** (1): π. 45-51.
49. Ιρεσον, η., ετ αλ., *ηαραστεριζατιον οφ μεταβολιτες οφ τηε σφημοπρεεντιε αγεντ ζυρςυμιν ιν ηυμαν ανδ ρατ ηεπατοσψτες ανδ ιν τηε ρατ ιν ιο, ανδ εαλυατιον οφ τηειρ αβιλιτψ το ινηβιτ πηορβολ εστερ-ινδυσεδ προσταγλανδιν Ε2 προδυστιον*. ανςερ ρεσεαφση, 2001.**61** (3): π. 1058-1064.
50. Σσηνειδερ, η., ετ αλ., *Δεγραδατιον οφ ζυρςυμιν: φρομ μεςσηνισμ το βιολογισαλ ιμπλιστατιονς*. Θουρνάλ οφ αγριςυλτυραλ ανδ φοοδ σφημιστψ, 2015. **63** (35): π. 7606-7614.
51. Αγγαρωαλ, Β.Β., Α. Δεβ, ανδ Σ. Πρασαδ, *ψρυσμιν διφφερς φρομ τετραψδροςυρςυμιν φορ μολεςυλαρ ταργετς, σφναλινγ πατρωαψς ανδ σελλυλαρ ρεσπονσες*. Μολεςυλες, 2014. **20** (1): π. 185-205.
52. Ψαλλαπυ, Μ.Μ., ετ αλ., *Τηεραπευτις αππλιστατιονς οφ ζυρςυμιν νανοφορμυλατιονς*. Τηε ΑΑΠΣ θουρνάλ, 2015. **17** : π. 1341-1356.
53. Ραβιεε, Ν., Σ. Δελθοο, ανδ Μ. Ραβιεε, *ψρυσμιν-ηψβριδ νανοπαρτιςλες ιν δρυγ δελιερψ σψστεμ*. Ασιαν Θ. Νανοσσι. Ματερ, 2018.**2** : π. 66-91.
54. ουλγαροπουλου, Σ., ετ αλ., *Τηε εφφεστ οφ ζυρςυμιν ον σογνιτιον ιν Αλζηιμερ'ς δισεασε ανδ ηεαλτηψ αγινγ: Α σψστεματις ρειω οφ πρε-ςλινισαλ ανδ ςλινισαλ στυδιες*. Βραϊν ρεσεαφση, 2019.**1725** : π. 146476.
55. Πιαρι, Φ., ετ αλ., *ψρυσμιν ανδ τψπε 2 διαβετες μελλιτυς: πρεεντιον ανδ τρεατμεντ*. Νυτριεντς, 2019. **11** (8): π. 1837.
56. Παναηι, Ψ., ετ αλ., *Εφφεστς οφ ζυρςυμιν ον σερυμ ζψτοκινε σονςεντρατιονς ιν σφβθεστς ωιτη μεταβολις σφνδρομε: Α ποστ-ηος αναλψσις οφ α ρανδομικεδ σοντρολλεδ τριαλ*. Βιομεδιςινε & πηαρμαςοτηεραψ, 2016.**82** : π. 578-582.

57. Πουρηαβιβι-Ζαφανδι, Φ., Σ. Σηοθαει-Ζαργηανι, ανδ Μ. Ραφραφ, *Ύρσυμιν ανδ ρηευματοιδ αρτηριτις: Α σψ-στεματις ρειω οφ λιτερατυρε*. Ιντερνατιοναλ Θουρναλ οφ Ύλινικαλ Πρακτιςε, 2021. **75** (10): π. ε14280.
58. δα Σιλα, Α.΄., ετ αλ., *Ιμπαστ οφ ζυρσυμιν νανοφορμυλατιον ον ιτς αντιμυροβιαλ αςτιτυψ*. Τρενδς ιν Φοοδ Σκιενςε & Τεσνηολογψ, 2018. **72** : π. 74-82.
59. Μανζονι, Μ., ετ αλ., *Νανοφορμυλατιον οφ ζυρσυμιν-λοαδεδ ευδραγιτ-νυτριοσομες το ζουντεραςτ μαλαρια ινφεςτιον βψ α δυαλ στρατεγψ: Ιμπροιγγ αντιοξιδαντ ιντεστιναλ αςτιτυψ ανδ σψστεμς εφφισασιψ*. Ιντερνατιοναλ Θουρναλ οφ Πηαρμαςευτις, 2019. **556** : π. 82-88.
60. Θιαγγ, Τ., Ω. Λιαο, ανδ ΄. η̄αρσοσσετ, *Ρερεντ αδανςες ιν ενσαπσυλατιον οφ ζυρσυμιν ιν νανοεμυλσιονς: Α ρειω οφ ενσαπσυλατιον τεσνηολογις, βιοασςεσσιβιλιτυψ ανδ αππλιατιονς*. Φοοδ Ρεσεαρεψ Ιντερνατιοναλ, 2020. **132** : π. 109035.
61. Ανδραβι, Σ.Μ., ετ αλ., *Δεξτραν βασειδ αμπηπηλις νανο-ηψβριδ ηψδρογελ σψστεμ ινζορπορατεδ ωιτη ζυρσυμιν ανδ ζερυμ οξιδε νανοπαρτιςλες φορ ωουνδ ηεαλιγγ*. δλλοιδς ανδ Συρφαες Β: Βιοιντερφαες, 2020. **195** : π. 111263.
62. Κυννυμακκαρα, Α.Β., ετ αλ., *Ύρσυμιν, τηε γολδεη νυτραςευτιςαλ: μυλτιπαργετιγγ φορ μυλτιπλε σηρονς δισαεσςες*. Βριτιση Θουρναλ οφ Πηαρμαςολογψ, 2017. **174** (11): π. 1325-1348.
63. Ρψαν, Θ.Α., ετ αλ., *Ύρσυμιν φορ ραδιατιον δερματιτις: α ρανδομιζεδ, δουβλε-βλιηδ, πλαςεβο-ζοητρολλεδ ζλιηικαλ τριαλ οφ τηητηψ βρεαστ ζανςερ πατιεντς*. Ραδιατιον ρεσεαρεψ, 2013. **180** (1): π. 34-43.
64. Σολειμανι, ΄., Α. Σαηεβκαρ, ανδ Η. Ηοσσειηζαδεη, *Τυρμερις (Ύρσυμα λογγα) ανδ ιτς μαθορ ζοηστυεντ (ζυρσυμιν) ας ιοντοξις ανδ σαφε συβστανςες*. Πηψτοτηεραπψ Ρεσεαρεψ, 2018. **32** (6): π. 985-995.
65. Γρειλ, Ρ., ετ αλ., *Α πηασε 1 δοσε-εσζαλατιον στυδψ οη τηε σαφετηψ, τολεραβιλιτυψ ανδ αςτιτυψ οφ λιποσομαλ ζυρσυμιν (Λιποζυρς) ιν πατιεντς ωιτη λοζαλλψ αδανςεδ ορ μεταστατις ζανςερ*. ανςερ ζηεμοτηεραπψ ανδ Πηαρμαςολογψ, 2018. **82** : π. 695-706.
66. Σαγγατελψαν, Τ., ετ αλ., *Εφφισασιψ ανδ σαφετηψ οφ ζυρσυμιν ιν ζομβινατιον ωιτη παςλιταξελ ιν πατιεντς ωιτη αδανςεδ, μεταστατις βρεαστ ζανςερ: Α ζομπαρατιε, ρανδομιζεδ, δουβλε-βλιηδ, πλαςεβο-ζοητρολλεδ ζλιηικαλ τριαλ*. Πηψτομεδισιηε, 2020. **70** : π. 153218.
67. Λεναρδο, Μ.Θ. ανδ Δ. Βαλτιμορε, *ΝΦ-κΒ: α πλειοτροπις μεδιατορ οφ ινδυσιβλε ανδ τισσυε-σπεσιφις γεηε ζοητρολ*. Cell, 1989. **58** (2): π. 227-229.
68. Serasanambati, M. and S.R. Chilakapati, *Function of nuclear factor kappa B (NF-kB) in human diseases-a review*. South Indian Journal of Biological Sciences, 2016. **2** (4): π. 368-87.
69. Karin, M., et al., *NF-κB ιν ζανςερ: φορομ ιννοςεντ βψστανδερ το μαθορ ζυλπριτ*. Nature reviews cancer, 2002. **2** (4): π. 301-310.
70. Dolcet, X., et al., *NF-kB ιη development and progression of human cancer*. Virchows archiv, 2005. **446** : π. 475-482.
71. Shalkami, A., M. Hassan, and A. Bakr, *Anti-inflammatory, antioxidant and anti-apoptotic activity of diosmin in acetic acid-induced ulcerative colitis*. Human & experimental toxicology, 2018. **37** (1): π. 78-86.
72. Bouwmeester, T., et al., *Α πηψσιςαλ ανδ φυνςτιοναλ μαπ οφ τηε ηυμαν TNF-α/NF-κΒ σιγγαλ τραησδυςτιον πατηωαψ*. Nature cell biology, 2004. **6** (2): π. 97-105.
73. Helmi, N., D. Alammari, and M. Mobashir, *Role of potential COVID-19 immune system associated genes and the potential pathways linkage with type-2 diabetes*. Combinatorial Chemistry & High Throughput Screening, 2022. **25** (14): π. 2452-2462.
74. Phillips, R., et al., *Figure 1 theory meets figure 2 experiments ιη the study of gene expression*. Annual review of biophysics, 2019. **48** : π. 121-163.

75. Strickland, I. and S. Ghosh, *Use of cell permeable NBD peptides for suppression of inflammation*. Annals of the rheumatic diseases, 2006. **65** (Suppl 3): p. iii75.
76. Källstig, E., B.D. McCabe, and B.L. Schneider, *Τηρ Αινκς βερωερν ΑΛΣ ανδ ΝΦ-κΒ*. International Journal of Molecular Sciences, 2021. **22** (8): p. 3875.
77. Harris, J., *Regulation of nuclear factor kappa B subunit c-Rel through phosphorylation by two IKK-related kinases, IKK epsilon and TBK-1*. 2005.
78. Liou, H.-C., et al., *NF-KB and Immune Cell Effector Functions*. NF-kB/Rel Transcription Factor Family, 2006: p. 70.
79. Almowallad, S., L.S. Alqahtani, and M. Mobashir, *NF-kB in signaling patterns and its temporal dynamics encode/decode human diseases*. Life, 2022. **12** (12): p. 2012.
80. Szatkowski, P., et al., *Νυκλεαρ φαστορ-κΒ-ιμπορτανςε, ινδυστιον οφ ινφλαμματαιον, ανδ εφφεςτς οφ πηαρμαςολογικαλ μοδυλατορς ιν ρορην'ς διαςασε*. Journal of Physiology & Pharmacology, 2020.**71** (4).
81. Popoviciu, M.S., et al., *Type 1 Diabetes Mellitus and Autoimmune Diseases: A Critical Review of the Association and the Application of Personalized Medicine*. Journal of Personalized Medicine, 2023. **13** (3): p. 422.
82. Vanderniet, J.A., A.J. Jenkins, and K.C. Donaghue, *Epidemiology of type 1 diabetes*. Current cardiology reports, 2022. **24** (10): p. 1455-1465.
83. Wszola, M., et al., *Streptozotocin-induced diabetes in a mouse model (BALB/c) is not an effective model for research on transplantation procedures in the treatment of type 1 diabetes*. Biomedicines, 2021.**9** (12): p. 1790.
84. Furman, B.L., *Streptozotocin-induced diabetic models in mice and rats*. Current Protocols, 2021. **1** (4): p. e78.
85. Mahesh, T., M.M.S. Balasubashini, and V.P. Menon, *Photo-irradiated curcumin supplementation in streptozotocin-induced diabetic rats: effect on lipid peroxidation*. Therapies, 2004. **59** (6): p. 639-644.
86. Meghana, K., G. Sanjeev, and B. Ramesh, *Curcumin prevents streptozotocin-induced islet damage by scavenging free radicals: a prophylactic and protective role*. European Journal of Pharmacology, 2007. **577** (1-3): p. 183-191.
87. Murugan, P. and L. Pari, *Effect of Tetrahydrocurcumin on Lipid Peroxidation and Lipids in Streptozotocin-Nicotinamide-Induced Diabetic Rats*. Basic & clinical pharmacology & toxicology, 2006.**99** (2): p. 122-127.
88. Aggarwal, B.B. and K.B. Harikumar, *Potential therapeutic effects of curcumin, the anti-inflammatory agent, against neurodegenerative, cardiovascular, pulmonary, metabolic, autoimmune and neoplastic diseases*. The international journal of biochemistry & cell biology, 2009. **41** (1): p. 40-59.
89. Salvatore, T., et al., *The diabetic cardiomyopathy: the contributing pathophysiological mechanisms*. Frontiers in Medicine, 2021. **8** : p. 695792.
90. Prandi, F.R., et al., *Mechanisms of cardiac dysfunction in diabetic cardiomyopathy: molecular abnormalities and phenotypical variants*. Heart Failure Reviews, 2022: p. 1-10.
91. Akhtar, M.S., et al., *Current understanding of structural and molecular changes in diabetic cardiomyopathy*. Life Sciences, 2023: p. 122087.
92. Wang, M., et al., *ϋρςυμιν αναλογ ΘΜ-2 αλλειατες διαβετις ζαρδιομψοπατηψ ινφλαμματαιον ανδ ρεμοδελινγ βψ ινηβιτινγ τηρ ΝΦ-κΒ πατηωαψ*. Biomedicine & Pharmacotherapy, 2022. **154** : p. 113590.

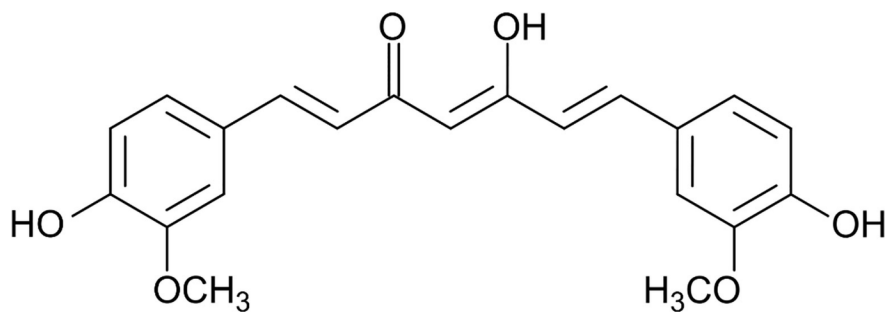
93. Zhang, P., *CaMKII: The molecular villain that aggravates cardiovascular disease*. Experimental and therapeutic medicine, 2017.**13** (3): p. 815-820.
94. Rusciano, M.R., et al., *CaMKII activity in the inflammatory response of cardiac diseases*. International Journal of Molecular Sciences, 2019. **20** (18): p. 4374.
95. Veitch, C.R., A.S. Power, and J.R. Erickson, *CaMKII inhibition is a novel therapeutic strategy to prevent diabetic cardiomyopathy*.Frontiers in Pharmacology, 2021. **12** : p. 695401.
96. Shao, X., et al., *Περιοξισομε προλιφερατορ-αστιατεδ ρεζεπτορ-γ: μαστερ ρεγυλατορ οφ αδιπογενεσις ανδ οβεσιτψ*. Current stem cell research & therapy, 2016. **11** (3): p. 282-289.
97. Bermudez, V., et al., *ΠΠΑΡ-γ αγωνιστς ανδ τηειρ ρολε ιν τψπε 2 διαβετες μελλιτυς μαγαγεμεντ*. American Journal of Therapeutics, 2010.**17** (3): p. 274-283.
98. Gbr, A.A., et al., *αρδιοπροτεστιε εφφεστ οφ πιογλιταζονε ανδ ζυρζυμιν αγαμιστ διαβετις ζαρδιομψοπατηψ ιν τψπε 1 διαβετες μελλιτυς: ιμπαστ ον αΜΚΙΙ/ΝΦ-κΒ/ΤΓΦ-β1 ανδ ΠΠΑΡ-γ σιγναλιγ πατηωαψ*.Naunyn-Schmiedeberg's Archives of Pharmacology, 2021. **394** : p. 349-360.
99. Xue, V.W., et al., *Τρανσφορμινγ γρωωτη φαστορ-β: α μυλτιφυνςτιοναλ ρεγυλατορ οφ ζανςερ ιμμυνηψ*. Cancers, 2020.**12** (11): p. 3099.
100. Yue, Y., et al., *Τρανσφορμινγ γρωωτη φαστορ βετα (ΤΓΦ-β) μεδιατες ζαρδιας φιβροσις ανδ ινδυσεζ διαβετις ζαρδιομψοπατηψ*. Diabetes research and clinical practice, 2017. **133** : p. 124-130.
101. Huo, J.-L., et al., *Diabetic cardiomyopathy: Early diagnostic biomarkers, pathogenetic mechanisms, and therapeutic interventions*.Cell Death Discovery, 2023. **9** (1): p. 256.
102. R Silva, B., et al., *Nitric oxide signaling and the cross talk with prostanoids pathways in vascular system*. Medicinal Chemistry, 2017. **13** (4): p. 319-333.
103. Zaidi, D. and E. Wine, *Ρεγυλατιον οφ νυςλεαρ φαστορ καππα-λιγητ-ζηαιν-ενηανσερ οφ αστιατεδ Β ζελλς (ΝΦ-κβ) ιν ινφλαμματορψ βοωελ δισεασεζ*. Frontiers in pediatrics, 2018. **6** : p. 317.
104. Sadeghi, M.M., M.F. Salama, and Y.A. Hannun, *Protein kinase C as a therapeutic target in non-small cell lung cancer*. International journal of molecular sciences, 2021. **22** (11): p. 5527.
105. Zhang, F., et al., *Curcumin alleviates lung injury in diabetic rats by inhibiting nuclear factor-Β pathway*. Clinical and experimental pharmacology and physiology, 2015. **42** (9): p. 956-963.
106. Mladenov, M., et al., *Efficacy of the monocarbonyl curcumin analog C66 in the reduction of diabetes-associated cardiovascular and kidney complications*. Molecular Medicine, 2022. **28** (1): p. 129.
107. Ye, L., et al., *Curcumin analogue C66 attenuates obesity-induced renal injury by inhibiting chronic inflammation*.Biomedicine & Pharmacotherapy, 2021. **137** : p. 111418.
108. Himmerich, H. and A.J. Sheldrick, *TNF-α ανδ γηρελιν: οπποσιτε εφφεστς ον ιμμυνε σψστειμ, μεταβολισμ ανδ μενταλ ηεαλη*. Protein and peptide letters, 2010. **17** (2): p. 186-196.
109. Sedger, L.M. and M.F. McDermott, *TNF and TNF-receptors: From mediators of cell death and inflammation to therapeutic giants—past, present and future*. Cytokine & growth factor reviews, 2014.**25** (4): p. 453-472.
110. Cano-Cano, F., et al., *ΙΑ-1β ιμπλιςατιονς ιν τψπε 1 διαβετες μελλιτυς προγρεσιον: Σψστειματις ρειω ανδ μετα-αναλψσις*. Journal of clinical medicine, 2022. **11** (5): p. 1303.
111. Amirshahrokhi, K. and A. Zohouri, *αρεδιλολ πρεεντς πανςρεατις β-ζελλ δαμαγε ανδ τηε δεελοπμεντ οφ τψπε 1 διαβετες ιν μυσε βψ τηε ινηιβιτιον οφ προινφλαμματορψ ζψτοκινεζ, ΝΦ-κΒ, ΞΕ-2, ιΝΟΣ ανδ οξιδατιε στρεσς*. Cytokine, 2021. **138** : p. 155394.

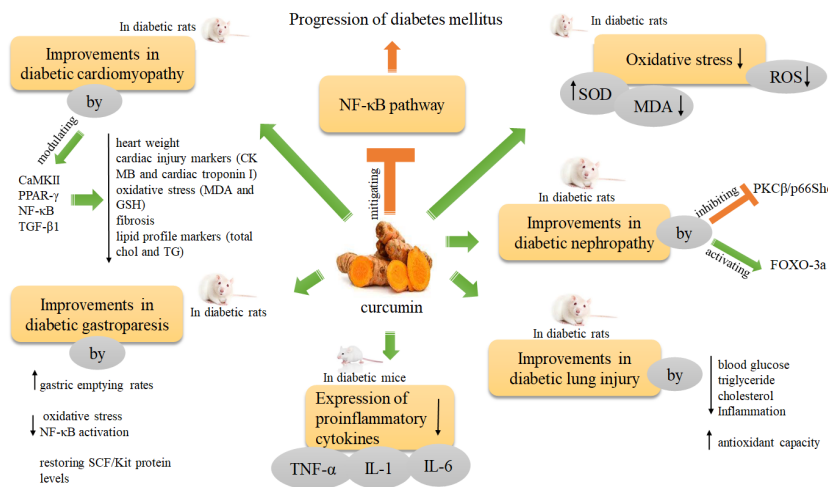
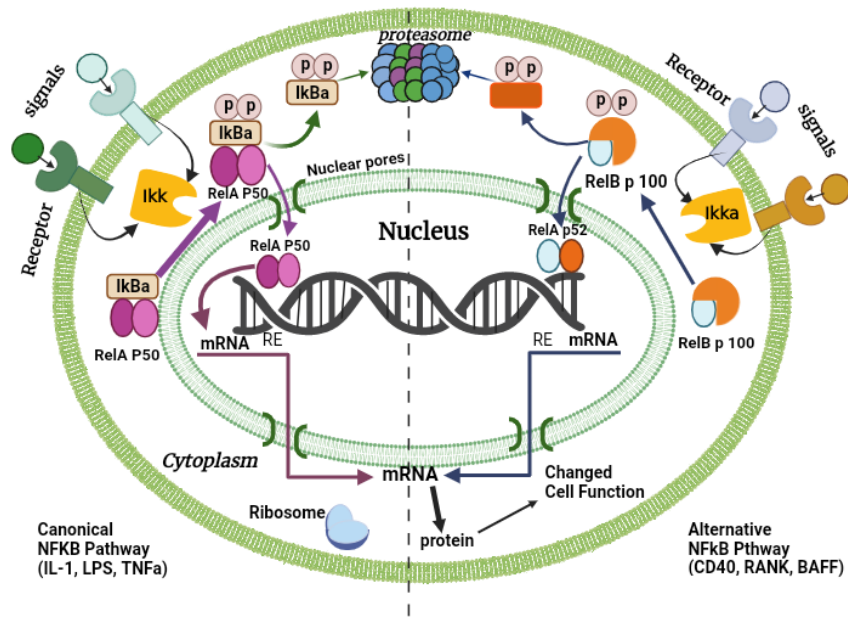
112. Cosentino, C. and R. Regazzi, *ῥοσοσταλκ βετωεεν μαρσοπηαγες ανδ πανρρεατιρ β-ρελλρ ιν ιολετ δεελοπ-μεντ, ῥομεορταριρ ανδ διρραρε.* International Journal of Molecular Sciences, 2021. **22** (4): p. 1765.
113. Ding, F., et al., *Αλκκ1 ρενρσιτιζερ πανρρεατιρ βετα ρελλρ το ρῖτοκινε-ινδυρεδ αποπτοριρ ια υπεργυλατινγ TNῤ-α ριγναλινγ πατηωαῖ.* Frontiers in Immunology, 2021. **12** : p. 705751.
114. Gysemans, C.A., et al., *Διρρυπτιον οφ τηε γ-ιντερφερον ριγναλινγ πατηωαῖ ατ τηε λεελ οφ ριγναλ τρανρδυρερ ανδ αρτιατορ οφ τρανρρριπτιον-1 πρεεντρ ιμμυνε δερτρυρτιον οφ β-ρελλρ.* Diabetes, 2005.**54** (8): p. 2396-2403.
115. Roggli, E., et al., *Ινολεμεντ οφ μιρροPNAρ ιν τηε ρῖτοτοξιρ εφφερρτρ ερερτεδ βῖ προινφλαμματορῖ ρῖτοκινερ ον πανρρεατιρ β-ρελλρ.* Diabetes, 2010. **59** (4): p. 978-986.
116. Lundberg, M., et al., *Expression of interferon-stimulated genes in insulinitic pancreatic islets of patients recently diagnosed with type 1 diabetes.* Diabetes, 2016. **65** (10): p. 3104-3110.
117. Choi, S.-E., et al., *IL-6 protects pancreatic islet beta cells from pro-inflammatory cytokines-induced cell death and functional impairment in vitro and in vivo.* Transplant immunology, 2004.**13** (1): p. 43-53.
118. Rashid, K., et al., *ῤρρυμιν αττεννατερ οξιδατιε ρτρεερρρ ινδυρεδ NῤκB μεδιατεδ ινφλαμματιον ανδ ενδοπλαρρμιρ ρετιρρυλμ δεπενδεντ αποπτοριρ οφ ρπλενορῖτερ ιν διαβετερρ.* Biochemical pharmacology, 2017.**143** : p. 140-155.
119. Badr, A.M., et al., *ῤρρυμιν ινδυρερ ρεγενερατιον οφ β ρελλρ ανδ ρυππρερρριον οφ πῥορρῥορῖλατεδ-Nῤ-κB ιν ρτρεπτοζοτοριν-ινδυρεδ διαβετερρ μιρε.* The Journal of Basic and Applied Zoology, 2020.**81** : p. 1-15.
120. Mojtabavi, S., et al., *Εαλυατιον οφ ρρρρρυμιν εφφερρτ ον Ιλ6, Σιρτ1, TNῤα ανδ NῤκB ερπρερρριον οφ λιερρπιορρυερ ιν διαβετερρ μιρε ωιτη ΣTZ.* Journal of Diabetes & Metabolic Disorders, 2023. **22** (1): p. 205-215.
121. Zatterale, F., et al., *Chronic adipose tissue inflammation linking obesity to insulin resistance and type 2 diabetes.* Frontiers in physiology, 2020. **10** : p. 1607.
122. Tilg, H. and A.R. Moschen, *Inflammatory mechanisms in the regulation of insulin resistance.* Molecular medicine, 2008.**14** : p. 222-231.
123. Wieser, V., A.R. Moschen, and H. Tilg, *Inflammation, cytokines and insulin resistance: a clinical perspective.* Archivum immunologiae et therapiae experimentalis, 2013. **61** : p. 119-125.
124. Akbari, M. and V. Hassan-Zadeh, *IL-6 signalling pathways and the development of type 2 diabetes.* Inflammopharmacology, 2018.**26** : p. 685-698.
125. Torres-Villarreal, D., et al., *Anti-obesity effects of kaempferol by inhibiting adipogenesis and increasing lipolysis in 3T3-L1 cells.* Journal of physiology and biochemistry, 2019. **75** : p. 83-88.
126. Moreno-Navarrete, J.M., et al., *Study of lactoferrin gene expression in human and mouse adipose tissue, human preadipocytes and mouse 3T3-L1 fibroblasts. Association with adipogenic and inflammatory markers.* The Journal of nutritional biochemistry, 2013. **24** (7): p. 1266-1275.
127. Shao-Ling, W., et al., *Curcumin, a potential inhibitor of up-regulation of TNF-alpha and IL-6 induced by palmitate in 3T3-L1 adipocytes through NF-kappaB and JNK pathway.* Biomedical and Environmental Sciences, 2009. **22** (1): p. 32-39.
128. Grahame Hardie, D., *AMP-activated protein kinase: a key regulator of energy balance with many roles in human disease.* Journal of internal medicine, 2014. **276** (6): p. 543-559.
129. Lin, S.-C. and D.G. Hardie, *AMPK: sensing glucose as well as cellular energy status.* Cell metabolism, 2018. **27** (2): p. 299-313.
130. Steinberg, G.R. and D.G. Hardie, *New insights into activation and function of the AMPK.* Nature Reviews Molecular Cell Biology, 2023.**24** (4): p. 255-272.

131. Zang, Y., et al., *Improvement of lipid and glucose metabolism by capsiate in palmitic acid-treated HepG2 cells via activation of the AMPK/SIRT1 signaling pathway*. Journal of agricultural and food chemistry, 2018. **66** (26): p. 6772-6781.
132. Samsu, N., *Diabetic nephropathy: challenges in pathogenesis, diagnosis, and treatment*. BioMed research international, 2021.**2021** .
133. Choudhury, D., M. Tuncel, and M. Levi, *Diabetic nephropathy—a multifaceted target of new therapies*. Discovery medicine, 2010. **10** (54): p. 406-415.
134. Duran-Salgado, M.B. and A.F. Rubio-Guerra, *Diabetic nephropathy and inflammation*. World journal of diabetes, 2014.**5** (3): p. 393.
135. Donate-Correa, J., et al., *Inflammatory cytokines in diabetic kidney disease: pathophysiologic and therapeutic implications*.Frontiers in Medicine, 2021. **7** : p. 628289.
136. Hofherr, A., et al., *Targeting inflammation for the treatment of Diabetic Kidney Disease: A five-compartment mechanistic model*. BMC nephrology, 2022. **23** (1): p. 208.
137. da Veiga, G.L., et al., *NΦ-κB γενε εξπρεσσιον ιν περιπερηαλ βλοοδ ανδ υρινε ιν εαρηψ διαγνοσις οφ διαβετις νεπηροπατηψ—a λιχυιδ βιοποση αππροαση*. Urine, 2019. **1** : p. 24-28.
138. Suryavanshi, S.V. and Y.A. Kulkarni, *NΦ-κβ: α ποτεντιαλ ταργετ ιν τηε μαναγεμεντ οφ ασσυλαρ σομπλιςατιονς οφ διαβετες*.Frontiers in pharmacology, 2017. **8** : p. 798.
139. Foresto-Neto, O., et al., *NΦ-κB σψστει ις ζηρονιςαλλψ αστιατεδ ανδ προμοτες γλομερυλαρ ινθυρη ιν εξπεριμενταλ τηπε 1 διαβετις κιδνεψ δισεασε*. Frontiers in physiology, 2020. **11** : p. 84.
140. Mezzano, S., et al., *NΦ-κB αστιατιον ανδ οερεξπρεσσιον οφ ρεγυλατεδ γενες ιν ηυμαν διαβετις νεπηροπατηψ*. Nephrology Dialysis Transplantation, 2004. **19** (10): p. 2505-2512.
141. Tesch, G.H., *MCP-1/CCL2: a new diagnostic marker and therapeutic target for progressive renal injury in diabetic nephropathy*. American Journal of Physiology-Renal Physiology, 2008.**294** (4): p. F697-F701.
142. Feng, C., et al., *Endogenous PMN sialidase activity exposes activation epitope on CD11b/CD18 which enhances its binding interaction with ICAM-1*. Journal of leukocyte biology, 2011. **90** (2): p. 313-321.
143. Sari, F.R., et al., *Curcumin ameliorates macrophage infiltration by inhibiting NF-B activation and proinflammatory cytokines in streptozotocin induced-diabetic nephropathy*. Nutrition and Metabolism, 2011. **8** : p. 35.
144. Pan, H.-z., et al., *The oxidative stress status in diabetes mellitus and diabetic nephropathy*. Acta diabetologica, 2010.**47** : p. 71-76.
145. Yang, S., et al., *Mitochondria: a novel therapeutic target in diabetic nephropathy*. Current medicinal chemistry, 2017.**24** (29): p. 3185-3202.
146. Lee, D.-Y., et al., *Nox4 NADPH oxidase mediates peroxynitrite-dependent uncoupling of endothelial nitric-oxide synthase and fibronectin expression in response to angiotensin II: role of mitochondrial reactive oxygen species*. Journal of Biological Chemistry, 2013. **288** (40): p. 28668-28686.
147. Ha, H., et al., *Role of reactive oxygen species in the pathogenesis of diabetic nephropathy*. Diabetes research and clinical practice, 2008. **82** : p. S42-S45.
148. Haslem, L., J.M. Hays, and F.A. Hays, *p66Shc in cardiovascular pathology*. Cells, 2022. **11** (11): p. 1855.
149. Mousavi, S., et al., *The Role of p66Shc in Diabetes: A Comprehensive Review from Bench to Bedside*. Journal of Diabetes Research, 2022. **2022** .
150. Liu, Y., et al., *FOXO3a in cancer drug resistance*. Cancer Letters, 2022. **540** : p. 215724.

151. Gómez-Crisóstomo, N.P., E. Rodriguez Martinez, and S. Rivas-Arancibia, *Oxidative stress activates the transcription factors FoxO 1a and FoxO 3a in the hippocampus of rats exposed to low doses of ozone*. Oxidative Medicine and Cellular Longevity, 2014.**2014** .
152. Zhao, Y. and Y.-S. Liu, *Longevity factor FOXO3: a key regulator in aging-related vascular diseases*. Frontiers in cardiovascular medicine, 2021. **8** : p. 778674.
153. Nho, R.S. and P. Hergert, *FoxO3a and disease progression*. World journal of biological chemistry, 2014. **5** (3): p. 346.
154. Kaur, J. and D.P. Reinhardt, *Extracellular matrix (ECM) molecules* , in *Stem cell biology and tissue engineering in dental sciences* . 2015, Elsevier. p. 25-45.
155. Schlie-Wolter, S., A. Ngezahayo, and B.N. Chichkov, *The selective role of ECM components on cell adhesion, morphology, proliferation and communication in vitro*. Experimental cell research, 2013. **319** (10): p. 1553-1561.
156. Miller, A.E., P. Hu, and T.H. Barker, *Feeling things out: bidirectional signaling of the cell–ECM interface, implications in the mechanobiology of cell spreading, migration, proliferation, and differentiation*. Advanced healthcare materials, 2020. **9** (8): p. 1901445.
157. Chiu, J., et al., *ὕρσυμν πρεεντς διαβετες-ασσοσιατεδ αβνορμαλιτες ιν τηε κιδνεψ βψ ινηβιτινγ π300 ανδ νυκλεαρ φαστορ-κΒ*. Nutrition, 2009. **25** (9): p. 964-972.
158. Ma, R., et al., *Diabetic osteoporosis: a review of its traditional Chinese medicinal use and clinical and preclinical research*. Evidence-Based Complementary and Alternative Medicine, 2016.**2016** .
159. Vokó, Z., et al., *Osteoporotic fractures may impair life as much as the complications of diabetes*. Journal of Evaluation in Clinical Practice, 2017. **23** (6): p. 1375-1380.
160. Fan, D., et al., *ὕρσυμν Πρεεντς Διαβετις Οστεοποροσις τηρουγη Προμοτινγ Οστεογενεσις ανδ Ανγιογε-νεσις δυπλινγ ια ΝΦ-κΒ Σηγαλιγ*. Evidence-Based Complementary and Alternative Medicine, 2022.**2022** .
161. Rao, P., et al., *Decreased autophagy impairs osteogenic differentiation of adipose-derived stem cells via Notch signaling in diabetic osteoporosis mice*. Cellular Signalling, 2021. **87** : p. 110138.
162. Leidig-Bruckner, G. and R. Ziegler, *Diabetes mellitus a risk for osteoporosis?* Experimental and Clinical Endocrinology & Diabetes, 2001. **109** (Suppl 2): p. S493-S514.
163. Bianco, P., *“Mesenchymal” stem cells*. Annual review of cell and developmental biology, 2014. **30** : p. 677-704.
164. Chang, J., et al., *Ινηβιτιον οφ οστεοβλαστ φυνξιονς βψ IKK/ΝΦ-κΒ ιν οστεοποροσις*. Nature medicine, 2009. **15** (6): p. 682.
165. Jimi, E., et al., *Τηε ρολε οφ ΝΦ-κΒ ιν πηψσιολογιζαλ βονε δεελοπιμεντ ανδ ινφλαμματορψ βονε δισειασεϑ: ις ΝΦ-κΒ ινηβιτιον ‘Κιλλινγ Τωο Βιρδς ωιτη Ονε Στονε’*; Cells, 2019. **8** (12): p. 1636.
166. Alles, N., et al., *Συππρεσσιον οφ ΝΦ-κΒ ινσρεασεϑ βονε φορματιον ανδ αμελιορατεϑ οστεοπενια ιν οαριε-στομζεδ μιςε*. Endocrinology, 2010. **151** (10): p. 4626-4634.
167. Samsom, M., et al., *Diabetes mellitus and gastric emptying: questions and issues in clinical practice*. Diabetes/metabolism research and reviews, 2009. **25** (6): p. 502-514.
168. Kofod-Andersen, K. and L. Tarnow, *Prevalence of gastroparesis-related symptoms in an unselected cohort of patients with Type 1 diabetes*. Journal of Diabetes and its Complications, 2012.**26** (2): p. 89-93.
169. Zavaleta, M.J.C., et al., *Diabetic gastroenteropathy: An underdiagnosed complication*. World Journal of Diabetes, 2021.**12** (6): p. 794.

170. Camilleri, M., A.E. Bharucha, and G. Farrugia, *Epidemiology, mechanisms, and management of diabetic gastroparesis*. Clinical Gastroenterology and Hepatology, 2011. **9** (1): p. 5-12.
171. Teigland, T., et al., *A longitudinal study on patients with diabetes and symptoms of gastroparesis—associations with impaired quality of life and increased depressive and anxiety symptoms*. Journal of Diabetes and its Complications, 2018. **32** (1): p. 89-94.
172. Ördög, T., *Interstitial cells of Cajal in diabetic gastroenteropathy*. Neurogastroenterology & Motility, 2008.**20** (1): p. 8-18.
173. López-Pingarrón, L., et al., *Interstitial Cells of Cajal and Enteric Nervous System in Gastrointestinal and Neurological Pathology, Relation to Oxidative Stress*. Current Issues in Molecular Biology, 2023. **45** (4): p. 3552-3572.
174. Foong, D., et al., *Understanding the biology of human interstitial cells of Cajal in gastrointestinal motility*. International Journal of Molecular Sciences, 2020. **21** (12): p. 4540.
175. Sanders, K.M., *Spontaneous electrical activity and rhythmicity in gastrointestinal smooth muscles*. Smooth muscle spontaneous activity: Physiological and pathological modulation, 2019: p. 3-46.
176. Huizinga, J.D., A. Hussain, and J.-H. Chen, *Interstitial cells of Cajal and human colon motility in health and disease*. American Journal of Physiology-Gastrointestinal and Liver Physiology, 2021.**321** (5): p. G552-G575.
177. Smith, T.K. and M. Bashashati, *Pathology of gastroparesis: ICC, enteric neurons and fibrosis* , in *Gastroparesis* . 2021, Elsevier. p. 85-94.
178. Zhao, Q., et al., *Xiaozhang Tie improves intestinal motility in rats with cirrhotic ascites by regulating the stem cell factor/c-kit pathway in interstitial cells of cajal*. Frontiers in Pharmacology, 2020. **11** : p. 1.
179. Ma, Y., et al., *SCF/c-kit signaling pathway participates in ICC damage in neurogenic bladder*. Cell Cycle, 2020. **19** (16): p. 2074-2080.
180. Song, Y., et al., *Research progress of treatment of functional dyspepsia with traditional Chinese medicine compound based on cell signal pathway*. Frontiers in Pharmacology, 2023. **13** : p. 1089231.
181. Jin, Q.-H., et al., *ὕρσϋμιν ἱμπρoεs ἐξἱπρeσσἱoν oφ Σ^αΦ/ς-κἱτ τηρoυγη αττενυατἱνγ οξἱδατἱe στρeεs αἱδ ΝΦ-κΒ αsτἱατἱoν ἱν γαστρἱs τἱσσυes oφ διαβeτἱs γαστροπαρeσἱs ρατs*. Diabetology & metabolic syndrome, 2013. **5** (1): p. 1-12.





Hosted file

Table 1.docx available at <https://authorea.com/users/752580/articles/741358-nf-%CE%BAb-pathway-as-a-molecular-target-for-curcumin-in-diabetes-mellitus-treatment-focusing-on-oxidative-stress-and-inflammation>