Clare Bush¹

 $^{1}\mathrm{Department}$ of Molecular Genetics, University of Louisiana

November 20, 2023

Review

miRNA: A Potential Breast Cancer Treatment? How did we get here?

Clare Bush 1*

- 1. Department of Molecular Genetics, University of Louisiana, Louisiana, USA
- * Corresponding Author: Email: <u>kekmfxlf@telegmail.com</u>

Abstract

Objective

The purpose of this research was to summarize what is currently known about the role of microRNAs in breast cancer and their possible clinical use based on articles published in peer-reviewed publications up to March 2014.

Results

MicroRNAs (miRNAs) are a kind of tiny RNA that typically range in length from 21 to 25 nucleotides. It is already common knowledge that microRNAs (miRNAs) play a crucial role in almost every cellular activity in the body, from development to tumorigenesis. MiRNAs in the bloodstream have been demonstrated to be appealing, readily detectable tumor biomarkers in a number of investigations. Breast cancer is among the most frequent forms of the disease. Different subtypes have been shown to have varying therapeutic responses, metastatic potential, and medication resistance in clinical trials. MicroRNAs may be useful in several aspects of breast cancer care, including diagnosis, prognosis, and treatment.

Conclusion

To choose the most appropriate treatment for each patient, molecular understanding is essential. MicroRNAs have promising use in cancer treatment.

Keywords: Tumorigenesis, miRNA, Breast Cancer, and Biomarkers

1. Why is it so difficult to find a cure for breast cancer?

Breast cancer is one of the most common cancers in the world with more than 1,300,000 cases worldwide.1 This very heterogeneous disease is clinically divided by histological types based on expression of specific receptors: estrogen receptor (ER) positive (the most numerous and diverse), progesterone receptor (PR) and HER2 (ERBB2) receptor or the absence of all of them, named triple negative breast cancer (TNBC).1, 2, 3, 5, 26 Another classification of breast cancer distinguished luminal A, luminal B, basal and HER2 enriched groups.4 It is clinically established that different subtypes may respond differently to therapies, give metastases and present drug resistance.1, 6 For example, TNBC is associated with high risk of recurrence and distant metastases to the brain, compared to other, receptor positive tumours,7 and responses only to chemotherapy.1 Molecular knowledge is crucial for choosing the most suitable therapy for individual patients combined with cost-effectiveness of the treatment.98 Conventional treatment for breast cancer includes wide local excision, sentinel lymph node biopsy or axillary lymph node dissection, adjuvant medical treatment and radiotherapy to the whole breast.25, 99

2. MicroRNAs: A Quick Overview of Their Origin, Mechanism, and Function

MicroRNAs (miRNAs) are a class of short RNA molecules that range in length from 21 to 25 nucleotides8.9 No one anticipated the significant role that this class of molecules would play in gene regulation until it was discovered in 1993 by Victor Ambros et al.10 that the gene lin-4 did not code for a protein. It should come as no surprise that miRNA currently plays a role in tumour initiation and progression given its centrality in all known cellular processes of the organism, including but not limited to development, proliferation, apoptosis, differentiation, and organogenesis8, 9, 10, 11, 12. The biogenesis of miRNAs has been extensively studied.13 miRNAs can be intergenic, intronic, or exonic (in exons of coding or non-coding genes) and can be transcribed as a single miRNA from its own promoter (approximately 50% of the time) or as a cluster of miRNAs from a shared promoter.8, 14, 15 The transcription of miRNA is driven primarily by RNase III. The 5'-cap structure and the 3'-poly(A) tail of the long primary transcript (pri-miRNA) may be larger than 1 kb8, 16. Secondary hairpin structure formed by a region of imperfect complementarity is identified and processed by an RNAse complex. Stem-loop shaped pre-miRNAs are exported from the nucleus into the cytosol by exportin 5.17 The RNase III endonuclease DICER1 cleaves the pre-form, generating a double-stranded miRNA of 18-25 nucleotides in length.16 The double-strand is unwound and the single strands are incorporated into the RNA-Induced Silencing Complex - RISC. MicroRNAs, which are a part of this complex, can regulate gene expression posttranscriptionally by binding partial complementarity to target mRNA, typically leading to mRNA degradation or translation inhibition.19 These interactions typically occur near the 5' terminus of miRNA molecules, contrary to previous beliefs. Computational methods are used to predict mRNA-miRNA interactions, and a number of algorithms are currently available.20 It is known that 3'-UTR mRNA may contain multiple miRNA-binding sites for different miRNAs, and a single miRNA may bind multiple targets.21, 22 The "many targets" hypothesis suggests that 60% of the mRNAs have one or more evolutionarily conserved sequences and that they are able to interact with miRNA.

Studies on chronic lymphocytic leukemia (CLL) have demonstrated that miR-15a and miR-16-1 are knocked down or knocked out in around 69% of CLLs (deletion of chromosome 13q14). This is the first evidence of the involvement of microRNAs in human cancer and was documented in 2002 by Calin and et al.27. The findings prompted the researchers to map all microRNA genes,28 many of which are found at highly variable chromosomal regions that are susceptible to deletions or amplifications. When a transcript encoding a tumor suppressor gene is amplified, it may lead to increased production of the miRNA and, in turn, the silence of the tumour suppressor gene if the miRNA is engaged in the negative regulation of the transcript. However, miRNAs that act to silence oncogenes are often found in regions of the genome called fragile foci, where they are particularly vulnerable to damage from events like deletions or mutations. This means that changes in microRNA expression are the norm rather than the exception in human cancer. Below we provide a few instances of how microRNA is engaged in the cancerogenesis process, as outlined by Hanahan and Weinberg in 2000.

Cancer cells are distinguished by their unchecked capacity to multiply. Multiple studies have shown that miR-34 regulates p53 and Notch pathways, consistent with its tumour suppressor activity. It is well known that Notch signalling maintains the balance between cell proliferation, differentiation, and apoptosis.65 Therefore, alterations in this signalling pathway are linked to tumourigenesis. Notch-1 and Notch-2 are downstream genes of miR-34 in pancreatic cancer cells,67 and Ji et al. reported that miR-34 restoration in human gastric cancer cells decreased expression of target gene Notch66. It is worth noting that pancreatic cancer stem cells are enriched with tumor-initiating cells or cancer stem cells with high levels of Notch-1/2 and loss of miR-34, suggesting the engagement of miR-34 in pancreatic stem cell self-renew

MiR-199 is another miRNA that has a role in controlling the Notch pathway. miR-199b-5p adversely influenced the proliferation of medulloblastoma (MB) cells by inhibiting Hes-1 (a transcription factor). Moreover, over-expression of miR-199b-5p reduces the number of cells with a stem-like phenotype (CD133+) and inhibits the expression of multiple cancer stem-cell genes.68 It is interesting that the level of miR-199b-5p was much greater in the non-metastatic cases

than in the metastatic ones. Patients from first group (high level) have exhibited a greater overall survival. MiR-290 cluster may directly target the Retinoblastoma-like 2 protein (Rbl2), alter telomere integrity and telomere-length homeostasis.69 Small, non-coding RNA plays also a key function in angiogenesis. Some microRNAs, including as let7-f, miR-27b, and miR-130a, have been shown to promote angiogenesis in vitro70, 71. Furthermore, Lee et al. discovered that miR-378 acts as an oncogene by suppressing the expression of tumor suppressors Sufu and Fus-1 to promote tumor cell survival and angiogenesis.

Some microRNAs were shown to have higher expression in BC tumor relative to surrounding tissues, as found in a research by Mojdeh Mahmoudian and colleagues. There was an upregulation of hsa-miR-25-3p, -29a-5p, -105-3p, and -181b1-5p and a downregulation of hsa-miR-335-5p and -339-5p. Except for hsa-miR-339-5p, these putative microRNAs were shown to have an up or downregulation related with TNM stages. All potential microRNAs linked with HER-2 status, with the exception of hsa-miR-105-3p. Further, ROC analysis indicated that the combination of these six microRNAs has the potential to act as a biomarker to distinguish between tumor and non-tumor breast tissue samples.

In human tumors, metastasis is the leading cause of death. There is currently strong evidence that microRNAs coordinate and play important roles in tumour invasion and metastasis.74, 75 Si et al. found that miR-21 was highly overexpressed in breast tumors compared to the matched normal breast tissues among 157 human miRNAs analyzed. MiR-21 has been demonstrated to inhibit tumor cell invasion and decrease expression of Pdcd4 in cultured colon cancer cells, according to research by Asangani et al.

4. Clinical methods for miRNA detection

Biomarkers must be used in clinical practice for detection and early diagnosis. Because of their compact size, miRNAs are very stable, making them an ideal diagnostic tool. It is important to further investigate the miRNA topic because of its potential use in tumor diagnosis, prognosis, and cure.29 Numerous studies have been developed on the global expression of miRNAs, and they can be detected and isolated from frozen and paraffin-embedded tissues, blood,37, 38 and various biologic fluids like urine,39 sputum40 or saliva41. MiRNA expression profiles (miRNome) have been shown to be superior than global mRNA profiling in discriminating between cancer types32, 35. It's also important to note that miRNA expression profile is tissue-specific, which might be useful in clinical diagnosis and treatment.36

Kong et al.24 provide a review of methods and strategies for profiling miRNA expression, and 46 the most pressing question is whether or not the miRNAs changing in a given disease are representative of, and unique to, that disease. Some miRNAs are already being used in clinical practice; for instance, the FDA has approved a miRNA-based diagnostic assay.47 The most widely used techniques for detecting miRNA are quantitative-reverse transcription polymerase chain reaction (qRT-PCR), hybridization-based methods, and next generation sequencing [review 47]. Using the Solexa deep-sequencing technology, Wei et al. systematically screened the miRNA expression profile of T-47D breast cancer cells treated with and without prolactin.62 They found a number of miRNAs significantly differentially expressed between these two panels and also detected several new miRNAs associated with the prolactin receptor signaling pathway in breast cancer. Several cutting-edge technological approaches have been proposed for miRNA research and analysis; these include a nanopore sensor based on the -haemolysin protein49 and an electrochemical genosensor that can readily detect miRNA in the serum or other biological samples.48 However, there are still significant limitations to use miRNA expression as routine in the clinic; thus, more clinical studies are needed.

5. The role of microRNA in breast cancer: a potential roadblock to curative treatment?

Iorio et al. (2005) were the first to report changes in miRNA expression in human breast cancer.50 They examined 76 breast tumors, 10 normal breast samples, and 14 breast cancer cell lines to determine which miRNAs are substantially

downregulated in cancerous tissues compared to normal ones. Microarray and Northern blot analysis found 29 miRNAs in this panel to have abnormal expression. The most often altered microRNAs in breast cancer were microRNA-10b, microRNA-125b, microRNA-145, and microRNA-21 and microRNA-155. This study shows that they may have a role in breast cancer tumourogenesis as either a tumour suppressor (down-regulated) or an oncogene (upregulated). In 2007, Sempere et al. published a paper comparing the distribution of miRNas in breast tumor tissue and normal tissue from more than 100 patients using hybridization in situ.52 miR-125b expression is known to be high in differentiated cells and tissues51, so its decreased level in breast cancer suggests impaired differentiation capabilities of cancer cells. As was previously noted, miRNAs may suppress oncogenes, hence preventing tumorigenesis. In breast cancer, members of the ErbB family are frequently amplified or overexpressed (20-30%) and are significantly associated with a worse prognosis53, 54. Scott et al. examined miR-125a and miR-125b overexpression in SKBR3 cells, which decreased ErbB2 protein level by approximately 40-65% and ErbB3 level by around 60-80%.54 Wang et al.55 provided the first experimental evidence that miR-125a, miR-15b, and miR-205 collaborate to control the expression of ErbB2/ErbB3 in breast cancer cells.56 ErbB family is a promising target for selective anticancer medicines due to its involvement in tumourogenesis. There are two main types of ErbB-targeted medicines now in use in the clinic: tyrosine kinase inhibitors (such aslapatinib against EGFR and ErbB) and blocking antibodies (like transtuzumab targeting ErbB2). Antibody56 is the only known method of blocking the ErbB3 receptor, and it is now being examined in human clinical trials (http://www.clinical-trials.gov/). This is because to the poor or absent kinase activity of this receptor. Several studies have shown that ErbB2 requires ErbB3 to promote breast cancer cell proliferation57 and that ErbB3 plays an important role in ErbB2-altered breast cancers58. The same research team also found that ErbB3 contributes to ErbB2-mediated therapeutic resistance to tamoxifen59 and paclitaxel60, and they proposed a novel strategy to target ErbB2/ErbB3 by reducing their protein levels by microRNA rather than inhibiting only the

Two miRNAs, miR-10b* and miR-139-5p, were down-regulated and three, miR-425, miR-454, and miR-301a, were upregulated for all three subtypes; they were shown to be differently expressed by Biagioni et al. Importantly, the soft agar colony formation experiment in MCF7 breast cancer cells demonstrated that lower levels of miR-10b* related to larger tumor growth, suggesting that this miRNA acts as a master regulator of breast cancer cell proliferation. Exploring the therapeutic potential of miR-10b* in breast cancer may need further in vivo research in a xenograft model, which demonstrated a vital function for miR-10b* in breast cancer cell proliferation64. Zhong et al. found that an altered miRNA expression pattern is important in gaining resistance to adriamycin and docetaxel which are two chemotherapeutic drugs routinely used in the treatment of breast cancer. Targeting PTEN.79 may be involved in this control.

As was previously established, miR-21 is crucial in the development of tumors. Wang et al. looked at how miR-21 expression correlated with doxorubicin sensitivity in breast cancer cells. They demonstrated that miR-21 dysregulation is crucial to doxorubicin resistance in breast cancer by using TaqMan RT-PCR and Western blot assay to identify mature miR-21 and tumour suppressor gene (PTEN) protein expression.80

Deregulation of miRNA-200c is associated with treatment resistance in breast cancer. Researchers found that miRNA-200c was down-regulated in non-responders compared to responders after analyzing miRNA expression in tumor samples from 39 breast cancer patients who had received neoadjuvant chemotherapy.81 Similarly, miRNA-30c was found to play a critical role in chemoresistance by directly targeting the actin-binding protein Twinfilin 1, which is responsible for promoting epithelial-to-mesenchymal transition.82 Both in vitro and in vivo, miR-19 inhibition sensitized MDR cells to chemotherapeutic agents.83 Although radiotherapy is a tried-and-true method of treating cancer, nothing is known about how microRNA may affect radiation resistance [revived in 85] in breast cancer at the present time. Interestingly, analyses have been performed on different cell lines that differed in their TP53 status, and the results suggest that miR-95 promotes radiation resistance independently of the TP53 function, as shown by Liu et al.85 in prostate and breast cancer. Experiments using xenograft tumor models showed that tumors overexpressing

miR-95 underwent reduced necrosis and increased proliferation while being irradiated. The production of miRNA-21 in breast cancer cells leads to radiation resistance by interfering with cell cycle progression (radiation-induced G2/M arrest).86 miRNA-21 has a function in radioresistance as a radioresistant miRNA. Antisense targeting of miR-155 by Chen et al.87 and miRNA-302 as a possible radiotherapy sensitizer by Liang et al.89 provide evidence for this hypothesis.

Important microRNAs in metastasis are called metastamiRs. It was shown in a poor prognosis phenotype, migration, and invasion. When comparing tumors with and without lymph node metastasis, the relative expression of miR-155 was significantly higher in tumors with lymph node metastasis.87 Similarly, Petrovi et al. found that miR-21 expression levels were significantly elevated in invasive with non-invasive component and pure invasive cancers compared with normal tissu. miR-21 is a major particular element to the process of invasion, since it shows the greatest difference between non-invasive and pure invasive cancer samples compared to other studied group pairs.92 Resistance to trastuzumab is a serious problem in the treatment of HER2+ breast tumors. Gong et al. looked for miRNAs that were differently expressed in the trastuzumab-resistant breast cancer cells, and observed that miRNA-21 was up-regulated among the reported PTEN-targeting miRNAs. They demonstrated that re-sensitizing resistant cells to trastuzumab's therapeutic activities-growth arrest, proliferation suppression, and G1-S cell cycle check-could be achieved by inhibiting miR-21 using antisense oligonucleotides.93

Using a polyamidoamine (PAMAM) dendrimers vector, Mei et al. showed that combining taxol chemotherapy with miR-21 inhibitor treatment reduced cell viability and invasiveness, suggesting that this may be a promising novel therapeutic approach for the treatment of breast malignancies.94 Tamoxifen (the selective oestrogen receptor modulator) resistance is a clinical issue in the treatment of HER2 positive breast cancer. Using tamoxifen-resistant MCF-7 cell models, Cittelly et al.95 investigated the role of microRNAs (miRNAs) in this phenomenon and concluded that miR-342 regulates tamoxifen response in breast tumor cell lines by modulating the expression of genes involved in apoptosis and cell cycle progression in tumor cells. Thus, restoring miR-342 expression may be a potential therapeutic strategy to sensitizing and reducing the development of tamoxifen resistant breast cancers. Another research team has linked MiR-221/222.96 to tamoxifen resistance in breast cancer. Overexpression of miR-145 reduces levels of cancer cell survival factors and inhibits cancer cell growth and metastasis, according to data published in 2011 by Kim and colleagues.97 They showed that miR-145 delivered via an adenoviral vector system significantly suppressed tumor growth in mice bearing breast tumors.

U.S. The function of microRNAs in breast cancer resistance and sensitivity is being studied in a clinical study (NCT01612871) authorized by the National Institutes of Health. Women with metastatic invasive breast cancer or locally advanced breast cancer for whom tamoxifen or anti aromatase medication is appropriate are eligible to participate in the trial. Participants are being sought for this research at this time [http://www.clinicaltrials.gov]. You may find online descriptions of several more clinical studies. Deregulation of microRNAs has a role in carcinogenesis, invasion, metastasis, and resistance to therapy. Different forms of human tumors have been linked to aberrant miRNA expression, suggesting that miRNAs may act as tumour-suppressor genes or oncogenes. Due to its various roles in cellular homeostasis and one-hit-many-target route, miRNA shows considerable promise as a therapeutic tool. MiRNA has been suggested in the literature as a possible biomarker for use in personalized medicine as a therapeutic tool.

Funding: N/A

Conflicts of Interest: The authors declare that they have no competing interests

References

- 1) Mahmoudian M, Razmara E, Mahmud Hussen B, Simiyari M, Lotfizadeh N, Motaghed H, Khazraei Monfared A, Montazeri M, Babashah S. Identification of a six-microRNA signature as a potential diagnostic biomarker in breast cancer tissues. J Clin Lab Anal. 2021 Nov;35(11):e24010. https://doi.org/10.1002/jcla.24010 PMID: 34528314; PMCID: PMC8605139.
- 2) Adams BD, Wali VB, Cheng CJ, et al., 2016. miR-34a silences c-SRC to attenuate tumor growth in triple-negative breast cancer. Cancer Res, 76(4):927-939. https://doi.org/10.1158/0008-5472.CAN-15-2321
- 3) Amorim M, Salta S, Henrique R, et al., 2016. Decoding the usefulness of non-coding RNAs as breast cancer markers. J Transl Med, 14:265. https://doi.org/10.1186/s12967-016-1025-3
- 4) Anfossi S, Fu X, Nagvekar R, et al., 2018. MicroRNAs, regulatory messengers inside and outside cancer cells. In: Mettinger KL, Rameshwar P, Kumar V (Eds.), Exosomes, Stem Cells and MicroRNA. Springer, Cham, p.87-108. https://doi.org/10.1007/978-3-319-74470-4 6
- 5) Atkinson SR, Marguerat S, Bähler J, 2012. Exploring long non-coding RNAs through sequencing. Semin Cell Dev Biol, 23(2):200-205. https://doi.org/10.1016/j.semcdb.2011.12.003
- 6) Bai XD, Han GH, Liu Y, et al., 2018. MiRNA-20a-5p promotes the growth of triple-negative breast cancer cells through targeting RUNX3. Biomed Pharmacother, 103: 1482-1489. https://doi.org/10.1016/j.biopha.2018.04.165
- 7) Bayraktar R, Pichler M, Kanlikilicer P, et al., 2017. MicroRNA 603 acts as a tumor suppressor and inhibits triple-negative breast cancer tumorigenesis by targeting elongation factor 2 kinase. Oncotarget, 8(7):11641-11658. https://doi.org/10.18632/oncotarget.14264
- 8) Bhardwaj A, Singh H, Rajapakshe K, et al., 2017. Regulation of miRNA-29c and its downstream pathways in preneoplastic progression of triple-negative breast cancer. Oncotarget, 8(12):19645-19660. https://doi.org/10.18632/oncotarget.14902
- 9) Biswas T, Efird JT, Prasad S, et al., 2017. The survival benefit of neoadjuvant chemotherapy and PCR among patients with advanced stage triple negative breast cancer. Oncotarget, 8(68):112712-112719. https://doi.org/10.18632/oncotarget.22521
- 10) Boon RA, Jaé N, Holdt L, et al., 2016. Long noncoding RNAs: from clinical genetics to therapeutic targets? J Am Coll Cardiol, 67(10):1214-1226. https://doi.org/10.1016/j.jacc.2015.12.051
- 11) Browne G, Dragon JA, Hong DL, et al., 2016. MicroRNA- 378-mediated suppression of Runx1 alleviates the aggressive phenotype of triple-negative MDA-MB-231 human breast cancer cells. Tumour Biol, 37(7):8825-8839. https://doi.org/10.1007/s13277-015-4710-6
- 12) Catalanotto C, Cogoni C, Zardo G, 2016. MicroRNA in control of gene expression: an overview of nuclear functions. Int J Mol Sci, 17(10):1712. https://doi.org/10.3390/ijms17101712
- 13) Chadwick BP, Scott KC, 2013. Molecular versatility: the many faces and functions of noncoding RNA. Chromosome Res, 21(6-7):555-559. https://doi.org/10.1007/s10577-013-9397-1
- 14) Chen H, Pan H, Qian Y, et al., 2018. MiR-25-3p promotes the proliferation of triple negative breast cancer by targeting BTG2. Mol Cancer, 17:4. https://doi.org/10.1186/s12943-017-0754-0
- 15) Chen J, Wang BC, Tang JH, 2012. Clinical significance of microRNA-155 expression in human breast cancer. J Surg Oncol, 106(3):260-266. https://doi.org/10.1002/jso.22153
- 16) Chen JW, Shin VY, Siu MT, et al., 2016. miR-199a-5p confers tumor-suppressive role in triple-negative breast cancer. BMC Cancer, 16:887. https://doi.org/10.1186/s12885-016-2916-7
- 17) Chen QN, Wei CC, Wang ZX, et al., 2017. Long non-coding RNAs in anti-cancer drug resistance. Oncotarget, 8(1): 1925-1936. https://doi.org/10.18632/oncotarget.12461
- 18) Chen XW, Zhao M, Huang J, et al., 2018. microRNA-130a suppresses breast cancer cell migration and invasion by targeting FOSL1 and upregulating ZO-1. J Cell Biochem, 119(6):4945-4956. https://doi.org/10.1002/jcb.26739
- 19) Collignon J, Lousberg L, Schroeder H, et al., 2016. Triple- negative breast cancer: treatment challenges and solutions. Breast Cancer (Dove Med Press), 8:93-107. https://doi.org/10.2147/BCTT.S69488

- 20) Costa FF, 2005. Non-coding RNAs: new players in eukaryotic biology. Gene, 357(2):83-94. https://doi.org/10.1016/j.gene.2005.06.019
- 21) De S, Das S, Mukherjee S, et al., 2017. Establishment of twist-1 and TGFBR2 as direct targets of microRNA-20a in mesenchymal to epithelial transition of breast cancer cell-line MDA-MB-231. Exp Cell Res, 361(1):85-92. https://doi.org/10.1016/j.yexcr.2017.10.005
- 22) Delás MJ, Hannon GJ, 2017. lncRNAs in development and disease: from functions to mechanisms. Open Biol, 7(7): 170121. https://doi.org/10.1098/rsob.170121
- 23) Deng H, Zhang J, Shi JJ, et al., 2016. Role of long non-coding RNA in tumor drug resistance. Tumor Biol, 37(9):11623-11631. https://doi.org/10.1007/s13277-016-5125-8
- 24) Eades G, Wolfson B, Zhang YS, et al., 2015. lincRNA-RoR and miR-145 regulate invasion in triple-negative breast cancer via targeting ARF6. Mol Cancer Res, 13(2):330-338. https://doi.org/10.1158/1541-7786.MCR-14-0251
- 25) Eades GL, Zhou Q, 2014. Abstract 1463: long non-coding RNA RoR and microRNA-145 regulate tumor cell invasion in triple-negative breast cancer via targeting of ADP- ribosylation factor 6. Cancer Res, 74(S19):1463. https://doi.org/10.1158/1538-7445.AM2014-1463
- 26) Evans JR, Feng FY, Chinnaiyan AM, 2016. The bright side of dark matter: lncRNAs in cancer. J Clin Invest, 126(8): 2775-2782. https://doi.org/10.1172/JCI84421
- 27) Fang H, Xie JP, Zhang M, et al., 2017. miRNA-21 promotes proliferation and invasion of triple-negative breast cancer cells through targeting PTEN. Am J Transl Res, 9(3): 953-961.
- 28) Ferlay J, Héry C, Autier P, et al., 2010. Global burden of breast cancer. In: Li C (Ed.), Breast Cancer Epidemiology. Springer, New York, p.1-19. https://doi.org/10.1007/978-1-4419-0685-4 1
- 29) Fu PF, Zheng X, Fan X, et al., 2019. Role of cytoplasmic lncRNAs in regulating cancer signaling pathways. J Zhejiang Univ-Sci B (Biomed & Biotechnol), 20(1):1-8. https://doi.org/10.1631/jzus.B1800254
- 30) Gebert LFR, MacRae IJ, 2019. Regulation of microRNA function in animals. Nat Rev Mol Cell Biol, 20(1):21-37. https://doi.org/10.1038/s41580-018-0045-7
- 31) Gilam A, Conde J, Weissglas-Volkov D, et al., 2016. Local microRNA delivery targets Palladin and prevents metastatic breast cancer. Nat Commun, 7:12868. https://doi.org/10.1038/ncomms12868
- 32) Gu J, Wang YP, Wang XD, et al., 2018. Downregulation of lncRNA GAS5 confers tamoxifen resistance by activating miR-222 in breast cancer. Cancer Lett, 434:1-10. https://doi.org/10.1016/j.canlet.2018.06.039
- 33) Gülben K, Berberoglu U, Kinaş V, et al., 2014. Breast cancer subtypes can be a predictor of pathologic complete response and survival in the neoadjuvant setting for T4 noninflammatory breast cancer. Acta Chir Belg, 114(3): 153-159. https://doi.org/10.1080/00015458.2014.11681001
- 34) Gupta RA, Shah N, Wang KC, et al., 2010. Long non-coding RNA HOTAIR reprograms chromatin state to promote cancer metastasis. Nature, 464(7291):1071-1076. https://doi.org/10.1038/nature08975
- 35) Han JG, Han BJ, Wu XY, et al., 2018. Knockdown of lncRNA H19 restores chemo-sensitivity in paclitaxel-resistant triple-negative breast cancer through triggering apoptosis and regulating Akt signaling pathway. Toxicol Appl Pharmacol, 359:55-61. https://doi.org/10.1016/j.taap.2018.09.018
- 36) Han JJ, Yu JJ, Dai YN, et al., 2018. Overexpression of miR- 361-5p in triple-negative breast cancer (TNBC) inhibits migration and invasion by targeting RQCD1 and inhibiting the EGFR/PI3K/Akt pathway. Bosn J Basic Med Sci, 19(1):52-59. https://doi.org/10.17305/bjbms.2018.3399
- 37) Harrow J, Frankish A, Gonzalez JM, et al., 2012. GENCODE: the reference human genome annotation for the encode project. Genome Res, 22(9):1760-1774. https://doi.org/10.1101/gr.135350.111
- 38) Hata A, Kashima R, 2016. Dysregulation of microRNA biogenesis machinery in cancer. Crit Rev Biochem Mol Biol, 51(3):121-134. https://doi.org/10.3109/10409238.2015.1117054
- 39) Hiatt RA, Brody JG, 2018. Environmental determinants of breast cancer. Annu Rev Public Health, 39:113-133. https://doi.org/10.1146/annurev-publhealth-040617-014101
- 40) Hong LQ, Pan F, Jiang HF, et al., 2016. MiR-125b inhibited epithelial-mesenchymal transition of triple-negative breast cancer by targeting MAP2K7. Onco Targets Ther, 9: 2639-2648. https://doi.org/10.2147/OTT.S102713

- 41) Hu JH, Xu J, Wu YQ, et al., 2015. Identification of microRNA- 93 as a functional dysregulated miRNA in triple-negative breast cancer. Tumour Biol, 36(1):251-258. https://doi.org/10.1007/s13277-014-2611-8
- 42) Huang J, Zhou N, Watabe K, et al., 2014. Long non-coding RNA UCA1 promotes breast tumor growth by suppression of p27 (Kip1). Cell Death Dis, 5:e1008. https://doi.org/10.1038/cddis.2013.541
- 43) Huarte M, 2015. The emerging role of lncRNAs in cancer. Nat Med, 21(11):1253-1261. https://doi.org/10.1038/nm.3981
- 44) Jia ZM, Liu Y, Gao Q, et al., 2016. miR-490-3p inhibits the growth and invasiveness in triple-negative breast cancer by repressing the expression of TNKS2. Gene, 593(1):41-47. https://doi.org/10.1016/j.gene.2016.08.014
- 45) Karagoz K, Sinha R, Arga KY, 2015. Triple negative breast cancer: a multi-omics network discovery strategy for candidate targets and driving pathways. OMICS, 19(2):115-130. https://doi.org/10.1089/omi.2014.0135
- 46) Khaled N, Bidet Y, 2019. New insights into the implication of epigenetic alterations in the EMT of triple negative breast cancer. Cancers (Basel), 11(4):559. https://doi.org/10.3390/cancers11040559
- 47) Kim SY, Kawaguchi T, Yan L, et al., 2017. Clinical relevance of microRNA expressions in breast cancer validated using The Cancer Genome Atlas (TCGA). Ann Surg Oncol, 24(10):2943-2949. https://doi.org/10.1245/s10434-017-5984-2
- 48) Kolesnikov NN, Veryaskina YA, Titov SE, et al., 2019. Expression of microRNAs in molecular genetic breast cancer subtypes. Cancer Treat Res Commun, 20:100026. https://doi.org/10.1016/j.ctarc.2016.08.006
- 49) Kunej T, Obsteter J, Pogacar Z, et al., 2014. The decalog of long non-coding RNA involvement in cancer diagnosis and monitoring. Crit Rev Clin Lab Sci, 51(6):344-357. https://doi.org/10.3109/10408363.2014.944299
- 50) Lee J, Jung JH, Chae YS, et al., 2016. Long noncoding RNA snaR regulates proliferation, migration and invasion of triple-negative breast cancer cells. Anticancer Res, 36(12): 6289-6295. https://doi.org/10.21873/anticanres.11224
- 51) Lehmann BD, Bauer JA, Chen X, et al., 2011. Identification of human triple-negative breast cancer subtypes and preclinical models for selection of targeted therapies. J Clin Invest, 121(7):2750-2767. https://doi.org/10.1172/JCI45014
- 52) Li HY, Liang JL, Kuo YL, et al., 2017. miR-105/93-3p promotes chemoresistance and circulating miR-105/93-3p acts as a diagnostic biomarker for triple negative breast cancer. Breast Cancer Res, 19:133. https://doi.org/10.1186/s13058-017-0918-2
- 53) Li J, Chen CC, Ma XC, et al., 2016. Long noncoding RNA NRON contributes to HIV-1 latency by specifically inducing TAT protein degradation. Nat Commun, 7:11730. https://doi.org/10.1038/ncomms11730
- 54) Li J, Cui ZG, Li H, et al., 2018. Clinicopathological and prognostic significance of long noncoding RNA MALAT1 in human cancers: a review and meta-analysis. Cancer Cell Int, 18:109. https://doi.org/10.1186/s12935-018-0606-z
- 55) Li N, Deng YJ, Zhou LH, et al., 2019. Global burden of breast cancer and attributable risk factors in 195 countries and territories, from 1990 to 2017: results from the global burden of disease study 2017. J Hematol Oncol, 12:140. https://doi.org/10.1186/s13045-019-0828-0
- 56) Li SQ, Zhou J, Wang ZX, et al., 2018. Long noncoding RNA GAS5 suppresses triple negative breast cancer progression through inhibition of proliferation and invasion by competitively binding miR-196a-5p. Biomed Pharmacother, 104:451-457. https://doi.org/10.1016/j.biopha.2018.05.056
- 57) Li WT, Liu CL, Zhao CL, et al., 2016. Downregulation of β3 integrin by miR-30a-5p modulates cell adhesion and invasion by interrupting Erk/Ets-1 network in triple- negative breast cancer. Int J Mol Sci, 48(3):1155-1164. https://doi.org/10.3892/ijo.2016.3319
- 58) Li XH, Hou LL, Yin L, et al., 2020. LncRNA XIST interacts with miR-454 to inhibit cells proliferation, epithelial mesenchymal transition and induces apoptosis in triple- negative breast cancer. J Biosci, 45:45. https://doi.org/10.1007/s12038-020-9999-7
- 59) Li XN, Wu YM, Liu AH, et al., 2016. Long non-coding RNA UCA1 enhances tamoxifen resistance in breast cancer cells through a miR-18a-HIF1α feedback regulatory loop. Tumor Biol, 37(11):14733-14743. https://doi.org/10.1007/s13277-016-5348-8

- 60) Li Z, Li Y, Li Y, et al., 2017. Long non-coding RNA H19 promotes the proliferation and invasion of breast cancer through upregulating DNMT1 expression by sponging miR-152. J Biochem Mol Toxicol, 31(9):e21933. https://doi.org/10.1002/jbt.21933
- 61) Li ZS, Meng QY, Pan AF, et al., 2017. MicroRNA-455-3p promotes invasion and migration in triple negative breast cancer by targeting tumor suppressor EI24. Oncotarget, 8(12):19455-19466. https://doi.org/10.18632/oncotarget.14307
- 62) Li ZX, Qian J, Li J, et al., 2019. Knockdown of lncRNA- HOTAIR downregulates the drug-resistance of breast cancer cells to doxorubicin via the PI3K/AKT/mTOR signaling pathway. Exp Ther Med, 18(1):435-442. https://doi.org/10.3892/etm.2019.7629
- 63) Liang YJ, Hu J, Li JT, et al., 2015. Epigenetic activation of TWIST1 by MTDH promotes cancer stem-like cell traits in breast cancer. Cancer Res, 75(17):3672-3680. https://doi.org/10.1158/0008-5472.CAN-15-0930
- 64) Liedtke C, Mazouni C, Hess K, et al., 2008. Response to neoadjuvant therapy and long-term survival in patients with triple-negative breast cancer. J Clin Oncol, 26(8):1275- 1281. https://doi.org/10.1200/JCO.2007.14.4147
- 65) Lin AF, Li CL, Xing Z, et al., 2016. The LINK-A lncRNA activates normoxic HIF1α signalling in triplenegative breast cancer. Nat Cell Biol, 18(2):213-224. https://doi.org/10.1038/ncb3295
- 66) Liu AN, Qu HJ, Gong WJ, et al., 2019. LncRNA AWPPH and miRNA-21 regulates cancer cell proliferation and chemosensitivity in triple-negative breast cancer by interacting with each other. J Cell Biochem, 120(9):14860-14866. https://doi.org/10.1002/jcb.28747
- 67) Liu HY, Wang G, Yang LL, et al., 2016. Knockdown of long non-coding RNA UCA1 increases the tamoxifen sensitivity of breast cancer cells through inhibition of Wnt/β-catenin pathway. PLoS ONE, 11(12):e0168406. https://doi.org/10.1371/journal.pone.0168406
- 68) Liu L, He J, Wei X, et al., 2017a. MicroRNA-20a-mediated loss of autophagy contributes to breast tumorigenesis by promoting genomic damage and instability. Oncogene, 36(42):5874-5884. https://doi.org/10.1038/onc.2017.193
- 69) Liu L, Yu DH, Shi H, et al., 2017b. Reduced IncRNA Aim enhances the malignant invasion of triple-negative breast cancer cells mainly by activating Wnt/β-catenin/mTOR/ PI3K signaling. Pharmazie, 72(10):599-603. https://doi.org/10.1691/ph.2017.7547
- 70) Liu M, Xing LQ, Liu YJ, 2017. A three-long noncoding RNA signature as a diagnostic biomarker for differentiating between triple-negative and non-triple-negative breast cancers. Medicine (Baltimore), 96(9):e6222. https://doi.org/10.1097/MD.00000000000000222
- 71) Liu XP, Tang HL, Chen JP, et al., 2015. MicroRNA-101 inhibits cell progression and increases paclitaxel sensitivity by suppressing MCL-1 expression in human triple- negative breast cancer. Oncotarget, 6(24):20070-20083. https://doi.org/10.18632/oncotarget.4039
- 72) Luan T, Zhang XM, Wang SY, et al., 2017. Long non-coding RNA MIAT promotes breast cancer progression and functions as ceRNA to regulate DUSP7 expression by sponging miR-155-5p. Oncotarget, 8(44):76153-76164. https://doi.org/10.18632/oncotarget.19190
- 73) Luo LY, Tang HL, Ling L, et al., 2018. LINC01638 lncRNA activates MTDH-Twist1 signaling by preventing SPOP- mediated c-Myc degradation in triple-negative breast cancer. Oncogene, 37(47):6166-6179. https://doi.org/10.1038/s41388-018-0396-8
- 74) Luo N, Zhang KJ, Li X, et al., 2020. ZEB1 induced-upregulation of long noncoding RNA ZEB1-AS1 facilitates the progression of triple negative breast cancer by binding with ELAVL1 to maintain the stability of ZEB1 mRNA. J Cell Biochem, online. https://doi.org/10.1002/jcb.29572
- 75) Lv ZD, Kong B, Liu XP, et al., 2016. miR-655 suppresses epithelial-to-mesenchymal transition by targeting Prrx1 in triple-negative breast cancer. J Cell Mol Med, 20(5): 864-873. https://doi.org/10.1111/jcmm.12770
- 76) Ma DC, Chen C, Wu J, et al., 2019. Up-regulated lncRNA AFAP1-AS1 indicates a poor prognosis and promotes carcinogenesis of breast cancer. Breast Cancer, 26(1):74-83. https://doi.org/10.1007/s12282-018-0891-3

- 77) Matamala N, Vargas MT, González-Cámpora R, et al., 2015. Tumor microRNA expression profiling identifies circulating microRNAs for early breast cancer detection. Clin Chem, 61(8):1098-1106. https://doi.org/10.1373/clinchem.2015.238691
- 78) Mathe A, Scott RJ, Avery-Kiejda K, 2015. miRNAs and other epigenetic changes as biomarkers in triple negative breast cancer. Int J Mol Sci, 16(12):28347-28376. https://doi.org/10.3390/ijms161226090
- 79) Mattick JS, 2011. The central role of RNA in human development and cognition. FEBS Lett, 585(11):1600-1616. https://doi.org/10.1016/j.febslet.2011.05.001
- 80) Mattick JS, Makunin IV, 2006. Non-coding RNA. Hum Mol Genet, 15(1):R17-R29. https://doi.org/10.1093/hmg/ddl046 Mattick JS, Amaral PP, Dinger ME, et al., 2009. RNA regulation of epigenetic processes. BioEssays, 31(1):51-59. https://doi.org/10.1002/bies.080099
- 81) Mayer IA, Abramson VG, Lehmann BD, et al., 2014. New strategies for triple-negative breast cancer—deciphering the heterogeneity. Clin Cancer Res, 20(4):782-790. https://doi.org/10.1158/1078-0432.CCR-13-0583
- 82) Miao YF, Fan RG, Chen LG, et al., 2016. Clinical significance of long non-coding RNA MALAT1 expression in tissue and serum of breast cancer. Ann Clin Lab Sci, 46(4):418-424.
- 83) Mou EX, Wang H, 2019. LncRNA LUCAT1 facilitates tumorigenesis and metastasis of triple-negative breast cancer through modulating miR-5702. Biosci Rep, 39(9): BSR20190489. https://doi.org/10.1042/BSR20190489
- 84) Niu LM, Fan QX, Yan M, et al., 2019. LncRNA NRON down- regulates lncRNA snaR and inhibits cancer cell proliferation in TNBC. Biosci Rep, 39(5):BSR20190468. https://doi.org/10.1042/BSR20190468
- 85) O'Brien K, Lowry MC, Corcoran C, et al., 2015. MiR-134 in extracellular vesicles reduces triple-negative breast cancer aggression and increases drug sensitivity. Oncotarget, 6(32):32774-32789. https://doi.org/10.18632/oncotarget.5192
- 86) Onyeagucha B, Rajamanickam S, Subbarayalu P, et al., 2016. Abstract P2-03-04: down-regulation of Bcl2-related ovarian killer (BOK) by miR-296-5p protects breast cancer cells from paclitaxel-induced apoptosis. Cancer Res, 76(S4): P2-03-04. https://doi.org/10.1158/1538-7445.SABCS15-P2-03-04
- 87) Paraskevopoulou MD, Hatzigeorgiou AG, 2016. Analyzing miRNA-lncRNA interactions. In: Feng Y, Zhang L (Eds.), Long Non-Coding RNAs: Methods and Protocols. Humana Press, New York, p.271-286. https://doi.org/10.1007/978-1-4939-3378-5 21
- 88) Phan B, Majid S, Ursu S, et al., 2016. Tumor suppressor role of microRNA-1296 in triple-negative breast cancer. Oncotarget, 7(15):19519-19530. https://doi.org/10.18632/oncotarget.6961
- 89) Piasecka D, Braun M, Kordek R, et al., 2018. MicroRNAs in regulation of triple-negative breast cancer progression. J Cancer Res Clin Oncol, 144(8):1401-1411. https://doi.org/10.1007/s00432-018-2689-2
- 90) Prensner JR, Chinnaiyan AM, 2011. The emergence of lncRNAs in cancer biology. Cancer Discov, 1(5):391-407. https://doi.org/10.1158/2159-8290.CD-11-0209
- 91) Razaviyan J, Hadavi R, Tavakoli R, et al., 2018. Expression of miRNAs targeting mTOR and S6K1 genes of mTOR signaling pathway including miR-96, miR-557, and miR-3182 in triple-negative breast cancer. Appl Biochem Biotechnol, 186(4):1074-1089. https://doi.org/10.1007/s12010-018-2773-8
- 92) Ren Y, Han XD, Yu K, et al., 2014. microRNA-200c downregulates XIAP expression to suppress proliferation and promote apoptosis of triple-negative breast cancer cells. Mol Med Rep, 10(1):315-321. https://doi.org/10.3892/mmr.2014.2222
- 93) Reshetnikova G, Troyanovsky S, Rimm DL, 2007. Definition of a direct extracellular interaction between Met and E- cadherin. Cell Biol Int, 31(4):366-373. https://doi.org/10.1016/j.cellbi.2007.01.022
- 94) Rhodes LV, Martin EC, Segar HC, et al., 2015. Dual regulation by microRNA-200b-3p and microRNA-200b-5p in the inhibition of epithelial-to-mesenchymal transition in triple- negative breast cancer. Oncotarget, 6(18):16638-16652. https://doi.org/10.18632/oncotarget.3184
- 95) Romero-Cordoba SL, Rodriguez-Cuevas S, Rebollar-Vega R, et al., 2016. A microRNA signature identifies subtypes of triple-negative breast cancer and reveals miR-342-3p as regulator of a lactate metabolic

- pathway through silencing monocarboxylate transporter 1. Cancer Res, 76(6):A47. https://doi.org/10.1158/1538-7445.NONRNA15-A47
- 96) Sha S, Yuan DY, Liu YJ, et al., 2017. Targeting long non-coding RNA DANCR inhibits triple negative breast cancer progression. Biol Open, 6(9):1310-1316. https://doi.org/10.1242/bio.023135
- 97) Shen X, Zhong JX, Yu P, et al., 2019. YY1-regulated LINC00152 promotes triple negative breast cancer progression by affecting on stability of PTEN protein. Biochem Biophys Res Commun, 509(2):448-454. https://doi.org/10.1016/j.bbrc.2018.12.074
- 98) Shin VY, Siu MT, Ho JC, et al., 2014. Abstract 531: miR- 199a-5p is a biomarker for and regulator of epithelial- mesenchymal transition in triple-negative breast cancer patients. Cancer Res, 74(S19):531. https://doi.org/10.1158/1538-7445.AM2014-531
- 99) Shin VY, Chen JW, Cheuk IWY, et al., 2019. Long non-coding RNA NEAT1 confers oncogenic role in triplenegative breast cancer through modulating chemoresistance and cancer stemness. Cell Death Dis, 10(4):270. https://doi.org/10.1038/s41419-019-1513-5
- 100) Shukla GC, Singh J, Barik S, 2011. MicroRNAs: processing, maturation, target recognition and regulatory functions. Mol Cell Pharmacol, 3(3):83-92. Siegel RL, Miller KD, Jemal A, 2019. Cancer statistics, 2019. CA Cancer J Clin, 69(1):7-34. https://doi.org/10.3322/caac.21551
- 101) Smith MA, Mattick JS, 2017. Structural and functional annotation of long noncoding RNAs. In: Keith JM (Ed.), Bioinformatics: Volume II: Structure, Function, and Applications. Humana Press, New York, p.65-85. https://doi.org/10.1007/978-1-4939-6613-4
- 102) Song GQ, Zhao Y, 2015. MicroRNA-211, a direct negative regulator of CDC25B expression, inhibits triple-negative breast cancer cells' growth and migration. Tumor Biol, 36(7):5001-5009. https://doi.org/10.1007/s13277-015-3151-6
- 103) Song X, Liu ZY, Yu ZY, 2019. LncRNA NEF is downregulated in triple negative breast cancer and correlated with poor prognosis. Acta Biochim Biophys Sin (Shanghai), 51(4):386-392. https://doi.org/10.1093/abbs/gmz021
- 104) Sørlie T, 2004. Molecular portraits of breast cancer: tumour subtypes as distinct disease entities. Eur J Cancer, 40(18): 2667-2675. https://doi.org/10.1016/j.ejca.2004.08.021
- 105) St. Laurent G, Wahlestedt C, Kapranov P, 2015. The landscape of long noncoding RNA classification. Trends Genet, 31(5):239-251. https://doi.org/10.1016/j.tig.2015.03.007
- 106) Sun WL, Yang YB, Xu CJ, et al., 2017. Regulatory mechanisms of long noncoding RNAs on gene expression in cancers. Cancer Genet, 216-217:105-110. https://doi.org/10.1016/j.cancergen.2017.06.003
- 107) Sun X, Li YQ, Zheng MZ, et al., 2016. MicroRNA-223 increases the sensitivity of triple-negative breast cancer stem cells to TRAIL-induced apoptosis by targeting HAX-1. PLoS ONE, 11(9):e0162754. https://doi.org/10.1371/journal.pone.0162754
- 108) Taft RJ, Pang KC, Mercer TR, et al., 2010. Non-coding RNAs: regulators of disease. J Pathol, 220(2):126-139. https://doi.org/10.1002/path.2638
- 109) Tian T, Wang M, Lin S, et al., 2018. The impact of lncRNA dysregulation on clinicopathology and survival of breast cancer: a systematic review and meta-analysis. Mol Ther Nucleic Acids, 12:359-369. https://doi.org/10.1016/j.omtn.2018.05.018
- 110) Tse JC, Kalluri R, 2007. Mechanisms of metastasis: epithelial- to-mesenchymal transition and contribution of tumor microenvironment. J Cell Biochem, 101(4):816-829. https://doi.org/10.1002/jcb.21215
- 111) Tsouko E, Wang J, Frigo DE, et al., 2015. miR-200a inhibits migration of triple-negative breast cancer cells through direct repression of the EPHA2 oncogene. Carcinogenesis, 36(9):1051-1060. https://doi.org/10.1093/carcin/bgv087
- 112) Verma A, Kaur J, Mehta K, 2019. Molecular oncology update: breast cancer gene expression profiling. Asian J Oncol, 1(2):65-72. https://doi.org/10.4103/2454-6798.173282
- 113) Wang B, Zhang QY, 2012. The expression and clinical significance of circulating microRNA-21 in serum of five solid tumors. J Cancer Res Clin Oncol, 138(10):1659-1666. https://doi.org/10.1007/s00432-012-1244-9

- 114) Wang C, Zheng XQ, Shen CY, et al., 2012. MicroRNA-203 suppresses cell proliferation and migration by targeting BIRC5 and LASP1 in human triple-negative breast cancer cells. J Exp Clin Cancer Res, 31:58. https://doi.org/10.1186/1756-9966-31-58
- 115) Wang H, Tan ZQ, Hu H, et al., 2019. microRNA-21 promotes breast cancer proliferation and metastasis by targeting LZTFL1. BMC Cancer, 19:738. https://doi.org/10.1186/s12885-019-5951-3
- 116) Wang J, Tsouko E, Jonsson P, et al., 2014. miR-206 inhibits cell migration through direct targeting of the actin-binding protein Coronin 1C in triple-negative breast cancer. Mol Oncol, 8(8):1690-1702. https://doi.org/10.1016/j.molonc.2014.07.006
- 117) Wang L, Liu DQ, Wu XR, et al., 2018. Long non-coding RNA (LncRNA) RMST in triple-negative breast cancer (TNBC): expression analysis and biological roles research. J Cell Physiol, 233(10):6603-6612. https://doi.org/10.1002/jcp.26311
- 118) Wang LH, Luan T, Zhou SH, et al., 2019. LncRNA HCP5 promotes triple negative breast cancer progression as a ceRNA to regulate BIRC3 by sponging miR-219a-5p. Cancer Med, 8(9):4389-4403. https://doi.org/10.1002/cam4.2335
- 119) Wang N, Hou MS, Zhan Y, et al., 2019a. LncRNA PTCSC3 inhibits triple-negative breast cancer cell proliferation by downregulating lncRNA H19. J Cell Biochem, 120(9): 15083-15088. https://doi.org/10.1002/jcb.28769
- 120) Wang N, Zhong CC, Fu MT, et al., 2019b. Long non-coding RNA HULC promotes the development of breast cancer through regulating LYPD1 expression by sponging miR- 6754-5p. Onco Targets Ther, 12:10671-10679. https://doi.org/10.2147/OTT.S226040
- 121) Wang OC, Yang F, Liu YH, et al., 2017. C-MYC-induced upregulation of lncRNA SNHG12 regulates cell proliferation, apoptosis and migration in triple-negative breast cancer. Am J Transl Res, 9(2):533-545.
- 122) Wang PS, Chou CH, Lin CH, et al., 2018. A novel long non-coding RNA linc-ZNF469-3 promotes lung metastasis through miR-574-5p-ZEB1 axis in triple negative breast cancer. Oncogene, 37(34):4662-4678. https://doi.org/10.1038/s41388-018-0293-1
- 123) Wang SW, Ke H, Zhang HL, et al., 2018. LncRNA MIR100HG promotes cell proliferation in triple-negative breast cancer through triplex formation with p27 loci. Cell Death Dis, 9(8):805. https://doi.org/10.1038/s41419-018-0869-2
- 124) Wang XL, Chen T, Zhang Y, et al., 2019. Long noncoding RNA Linc00339 promotes triple-negative breast cancer progression through miR-377-3p/HOXC6 signaling pathway. J Cell Physiol, 234(8):13303-13317. https://doi.org/10.1002/jcp.28007
- 125) Wang XS, Zhang Z, Wang HC, et al., 2006. Rapid identification of UCA1 as a very sensitive and specific unique marker for human bladder carcinoma. Clin Cancer Res, 12(16):4851-4858. https://doi.org/10.1158/1078-0432.CCR-06-0134
- 126) Wang YX, Zhang ZY, Wang JQ, 2018. MicroRNA-384 inhibits the progression of breast cancer by targeting ACVR1. Oncol Rep, 39(6):2563-2574. https://doi.org/10.3892/or.2018.6385
- 127) Winton MJ, Igaz LM, Wong MM, et al., 2008. Disturbance of nuclear and cytoplasmic TAR DNA-binding protein (TDP-43) induces disease-like redistribution, sequestration, and aggregate formation. J Biol Chem, 283(19): 13302-13309. https://doi.org/10.1074/jbc.M800342200
- 128) Wu CH, Luo J, 2016. Long non-coding RNA (IncRNA) urothelial carcinoma-associated 1 (UCA1) enhances tamoxifen resistance in breast cancer cells via inhibiting mtor signaling pathway. Med Sci Monit, 22:3860-3867. https://doi.org/10.12659/msm.900689
- 129) Wu JL, Shuang ZY, Zhao JF, et al., 2018. Linc00152 promotes tumorigenesis by regulating DNMTs in triplenegative breast cancer. Biomed Pharmacother, 97:1275-1281. https://doi.org/10.1016/j.biopha.2017.11.055
- 130) Xiong HP, Yan T, Zhang WJ, et al., 2018. miR-613 inhibits cell migration and invasion by downregulating Daam1 in triple-negative breast cancer. Cell Signal, 44:33-42. https://doi.org/10.1016/j.cellsig.2018.01.013
- 131) Xu ST, Xu JH, Zheng ZR, et al., 2017. Long non-coding RNA ANRIL promotes carcinogenesis via sponging miR-199a in triple-negative breast cancer. Biomed Pharmacother, 96:14-21. https://doi.org/10.1016/j.biopha.2017.09.107

- 132) Yang CF, Humphries B, Li YF, et al., 2017. Abstract 1468: miR-200b targets ARHGAP18 and suppresses triple negative breast cancer metastasis. Cancer Res, 77(S13):1468. https://doi.org/10.1158/1538-7445.AM2017-1468
- 133) Yang F, Liu YH, Dong SY, et al., 2016a. Co-expression networks revealed potential core lncRNAs in the triple- negative breast cancer. Gene, 591(2):471-477. https://doi.org/10.1016/j.gene.2016.07.002
- 134) Yang F, Dong SY, Lv L, et al., 2016b. Long non-coding RNA AFAP1-AS1 was up-regulated in triple-negative breast cancer and regulated proliferation and invasion. Int J Clin Exp Pathol, 9(6):6378-6384.
- 135) Yang J, Meng XL, Yu Y, et al., 2019. LncRNA POU3F3 promotes proliferation and inhibits apoptosis of cancer cells in triple-negative breast cancer by inactivating caspase 9. Biosci Biotechnol Biochem, 83(6):1117-1123. https://doi.org/10.1080/09168451.2019.1588097
- 136) Yoon MK, Mitrea DM, Ou L, et al., 2012. Cell cycle regulation by the intrinsically disordered proteins p21 and p27. Biochem Soc Trans, 40(5):981-988. https://doi.org/10.1042/bst20120092
- 137) Youness RA, Hafez HM, Khallaf E, et al., 2019. The long noncoding RNA sONE represses triple-negative breast cancer aggressiveness through inducing the expression of miR-34a, miR-15a, miR-16, and let-7a. J Cell Physiol, 234(11):20286-20297. https://doi.org/10.1002/jcp.28629
- 138) Yu FS, Wang L, Zhang BW, 2019. Long non-coding RNA DRHC inhibits the proliferation of cancer cells in triple negative breast cancer by downregulating long non-coding RNA HOTAIR. Oncol Lett, 18(4):3817-3822. https://doi.org/10.3892/ol.2019.10683
- 139) Zhang H, Li BW, Zhao HB, et al., 2015. The expression and clinical significance of serum miR-205 for breast cancer and its role in detection of human cancers. Int J Clin Exp Med, 8(2):3034-3043.
- 140) Zhang KJ, Luo ZL, Zhang Y, et al., 2016. Circulating lncRNA H19 in plasma as a novel biomarker for breast cancer. Cancer Biomark, 17(2):187-194. https://doi.org/10.3233/CBM-160630
- 141) Zhang KM, Liu P, Tang HL, et al., 2018. AFAP1-AS1 promotes epithelial-mesenchymal transition and tumorigenesis through Wnt/β-catenin signaling pathway in triple- negative breast cancer. Front Pharmacol, 9:1248. https://doi.org/10.3389/fphar.2018.01248
- 142) Zhang R, Xia LQ, Lu WW, et al., 2016. LncRNAs and cancer. Oncol Lett, 12(2):1233-1239. https://doi.org/10.3892/ol.2016.4770
- 143) Zhang YY, He Q, Hu ZY, et al., 2016. Long noncoding RNA LINP1 regulates repair of DNA double-strand breaks in triple-negative breast cancer. Nat Struct Mol Biol, 23(6): 522-530. https://doi.org/10.1038/nsmb.3211
- 144) Zhao D, Besser AH, Wander SA, et al., 2015. Cytoplasmic p27 promotes epithelial-mesenchymal transition and tumor metastasis via STAT3-mediated TWIST1 upregulation. Oncogene, 34(43):5447-5459. https://doi.org/10.1038/onc.2014.473
- 145) Zhao M, Ding XF, Shen JY, et al., 2017. Use of liposomal doxorubicin for adjuvant chemotherapy of breast cancer in clinical practice. J Zhejiang Univ-Sci B (Biomed & Biotechnol), 18(1):15-26. https://doi.org/10.1631/jzus.B1600303
- 146) Zhao ZT, Li L, Du PN, et al., 2019. Transcriptional downregulation of miR-4306 serves as a new therapeutic target for triple negative breast cancer. Theranostics, 9(5):1401-1416. https://doi.org/10.7150/thno.30701
- 147) Zheng LH, Zhang YH, Fu YJ, et al., 2019. Long non-coding RNA MALAT1 regulates BLCAP mRNA expression through binding to miR-339-5p and promotes poor prognosis in breast cancer. Biosci Rep, 39(2):BSR20181284. https://doi.org/10.1042/BSR20181284
- 148) Zuo YG, Li Y, Zhou ZY, et al., 2017. Long non-coding RNA MALAT1 promotes proliferation and invasion via targeting miR-129-5p in triple-negative breast cancer. Biomed Pharmacother, 95:922-928. https://doi.org/10.1016/j.biopha.2017.09.005