

Relationship between microRNA-9 and breast cancer

Alex Johannsson ¹, Alex Johannsson², Lakshmi Sharma², Hossein Mossavi², Liam Murphy², Oliver Walsh², Elizabeth Foreman², Anika Patel², and Dev Kumar²

¹Affiliation not available

²Department of biotechnology, University of North Dakota, Grand Forks

November 15, 2023

Relationship between microRNA-9 and breast cancer

Alex Johannsson 1* Lakshmi Sharma 2 Hossein Mossavi 3 Liam Murphy 4 Oliver Walsh 5 Elizabeth foreman 6 Anika Patel 7 Dev Kumar 8

1. Department of biotechnology, University of North Dakota, Grand Forks, ND, USA
2. Department of biotechnology, University of North Dakota, Grand Forks, ND, USA
3. Department of biotechnology, University of North Dakota, Grand Forks, ND, USA
4. Department of biotechnology, University of North Dakota, Grand Forks, ND, USA
5. Department of biotechnology, University of North Dakota, Grand Forks, ND, USA
6. Department of biotechnology, University of North Dakota, Grand Forks, ND, USA
7. Department of biotechnology, University of North Dakota, Grand Forks, ND, USA
8. Department of biotechnology, University of North Dakota, Grand Forks, ND, USA

* Corresponding Author: Email: lcotej@telegmail.com

Abstract

Breast cancer is the most frequent malignancy in women and has devastating effects on both their physical and emotional wellbeing. More cases of it are expected to surface in the future. Mounting data shows that microRNAs (miRNAs) play critical roles in carcinogenesis and development. This study highlights the contrasting roles of miR-9 in breast cancer, which is important since miR-9 plays a crucial role in both the development and the progression of the disease. The major focus of this article is a brief summary of miR-9's functions in the development of breast cancer.

Keyword: Tumorigenesis and development of breast cancer, miR-9, oncogene, tumor suppressor

1. Introduction

Among women, breast cancer has the highest incidence rate and is the second greatest cause of cancer-related mortality [1]. Breast cancer has a complicated and multifaceted etiology. More and more evidence suggests that miRNAs (microRNAs) have a role in controlling breast cancer development and progression [2,3]. Small noncoding RNAs (ncRNAs) called microRNAs (miRNAs) are found in all eukaryotic cells and are responsible for posttranscriptional regulation of gene expression by interacting with the 3' untranslated regions (3'UTRs) of target transcripts. More than 2500 mature miRNAs were found in human to far, which are involved in individual development, cell proliferation, differentiation and apoptosis [4]. Due to their central roles in carcinogenesis, cancer invasion, metastasis, relapse, and medication resistance [5, 6], miRNAs have received considerable interest in cancer research over the last several decades. Researchers found that miR-9 expression is distorted in breast cancer. The roles of miR-9 in controlling breast cancer, however, have been shown to be counterintuitive. In this article, we will examine the methods by which miR-9 influences the onset and progression of breast cancer.

2. Where microRNAs came from

While RNA polymerase II is the primary mediator of miRNA transcription, several other enzymes and proteins are involved in the maturation process [4]. The exact mechanism is as follows: Firstly, the first transcripts of miRNAs with

stem-ring structure are produced, designated as primary miRNAs (pri-miRNAs) which contain the thousands of nt in length. Secondly, the nucleic acid endonuclease Drosha cleaves the stem-ring structure, producing the pre-miRNAs with a hairpin length of around 70 nt. These pre-miRNAs are then exported to the cytoplasm by exportin 5, where they are further cleaved by another endonuclease, Dicer, into approximately 22 nt double-stranded RNAs. A number of cofactor proteins are involved in the cleavage of Drosha and Dicer; for example, DGCR8 is required for Drosha-mediated cleavage of pri-miRNA [6], while TRBP is required for Dicer-mediated cleavage of pre-miRNA [7]. In the end, these dsRNAs are loaded into Ago protein, establishing the RNA-induced silencing complex (RISC) [8], where one chain is kept in RISC to carry out its activities and the other is eliminated. Target 3'UTRs are targeted by miRNA, which directs RISC to block, cleave, or destroy the corresponding mRNA. Cleavage of target transcripts by RISC necessitates full complementary pairing of the miRNAs with target transcripts; however, the matching degree of miRNAs with target transcripts is not great in animals, leading to the unusual cleavage in animals. Given that a single miRNA may control several downstream targets, miRNAs have broad effects in both healthy and diseased states. Professor RA Weinberg of MIT is a renowned oncologist who has written two evaluations on the subject of cancer, "Hallmarks of Cancer" in 2000 and "Hallmarks of Cancer: the next generation" in 2011. He summed up cancer's defining features, discussed every angle of the disease, and is often referenced for his work. According to these two studies, miRNAs control every aspect of malignancies [9,10]. This article summarizes the current understanding of miR-9's functions and associated pathways in breast cancer.

In the research carried out by Mojdeh Mahmoudian and her colleagues, it was shown that the expression of particular microRNAs was higher in BC tumors than it was in the tissues that were next to the tumors. To be more specific, hsa-miR-25-3p, -29a-5p, -105-3p, and -181b1-5p were all found to have an increased expression level, whilst hsa-miR-335-5p and -339-5p were found to have a decreased expression level. With the exception of hsa-miR-339-5p, it was discovered that the TNM phases were related with the upregulation or downregulation of these putative microRNAs. In addition, all of the potential microRNAs linked with the status of HER-2, with the exception of hsa-miR-105-3p. In addition, the results of the analysis of ROC curves demonstrated that the combination of these six microRNAs has the potential to act as a biomarker that differentiates between breast tissue samples that include tumors and those that do not have tumors.

3. miR-9 enhances breast cancer's propensity for metastasis and the development of CSCs (cancer stem cells).

Recent research has overwhelmingly shown that miR-9 has a tumor-promoting function in breast cancer growth, mostly by boosting the disease's metastatic potential (Table 1). According to global miRNA expression profiling done by Grarguard et al. [11] on 47 matched primary and metastatic breast tumor samples from 14 patients with or without lymph node and distant metastases, miR-9 is considerably elevated in the metastatic tumors. Because of their specific expression profiles in cancer cells and tissues, their ability to enter the body's fluid circulation, and their relatively high chemical stability, miRNAs have been identified as a promising and non-invasive tumor marker. For example, Naorem LD et al. [12] used meta-analysis to pool data from ten miRNA profiling studies of triple-negative breast cancer (TNBC) and found that miR-9 is highly expressed; this finding has important implications for TNBC diagnostics.

CSCs have been proven to be the origin of cancer development and therapy resistance in recent investigations [13]. Cancer stem cells (CSCs) are a subset of tumor cells that may self-renew, undergo cell-to-cell differentiation, and react to phenotypic changes in the tumor microenvironment [14]. When polarized, non-migrating epithelial cells undergo epithelial-to-mesenchymal transition (EMT), they acquire the properties of migrating and invading mesenchymal cells as well as the capacity for self-renewal and stem cell-like properties, thereby increasing the generation of CSCs [15]. According to a systematic search of PubMed and Medline databases conducted by Piasecka D et al. [16], four microRNAs (miR-9, miR-221/222, miR-373, and miR-10b) were found to be significantly upregulated and strongly associated with invasion and EMT/CSC properties. Strong evidence was shown by Cheng CW et al. [17] showing overexpression of miR-9 and miR-221 signals a bad result and may promote CSC formation leading to an invasive

phenotype. According to Gwak JM et al. [18], cancers with a CD44+/CD24- phenotype, vimentin expression, and E-cadherin loss are more likely to express miR-9 than luminal tumors, those with a high tumor stage, and those with a high histologic grade. In sum, miR-9 is a perfect predictor of breast cancer start, metastasis, and chemoresistance since CSCs are thought to be at the root of carcinogenesis, development, and drug resistance.

4. MiR-9 is significantly associated with breast cancer outcome and recurrence.

Furthermore, miR-9 is an independent risk factor for shorter times without illness. There is a strong correlation between miR-9 and recurrence and survival in breast cancer (Table 1). MiR-9 is emerging as a biomarker for diagnosis and prognosis, and miR-9-based medication is a potential therapeutic for breast cancer, as revealed by a study of miRNAs implicated in the onset and course of the disease by Bertoli et al. [19]. Furthermore, miR-9 was included in a summary of miRNAs identified as possible diagnostic miRNAs in breast cancer by Bertoli et al. [20]. Low miR-9 expression appears to have a protective effect and is associated with improved overall survival (OS), smaller tumors, and ER (estrogen receptor, ER)-positive (ER+) cancers, as determined by an analysis of 985 breast cancers from The Cancer Genome Atlas (TCGA) by Sporn JC et al. [21]. High miR-9 levels may be a prognostic marker in TNBC, as demonstrated by Jang MH et al. [22]. This is because they are substantially correlated with poor disease-free survival and distant metastasis-free survival. In ER+ breast cancer cases, Zhou X et al. [23] showed that miR-9 level predicts LR, and that patients with a lower miR-9 level had a better 10-year LR-free survival.

Taken together, miR-9 might boost breast cancer metastasis and CSCs-like features notably in highly aggressive breast cancer such as TNBC. Furthermore, miR-9 might be a potential biomarker for breast cancer detection and prognosis, since it is strongly linked to breast cancer recurrence and a poor prognosis. So, how does miR-9 function?

5. The mode of action of miR-9

5.1. The post-transcriptional regulation of gene expression by miR-9 occurs by direct binding to target genes.

By binding to the 3'-untranslated regions of target genes, miRNAs suppress their expression after transcription has already taken place. This mechanism (Fig. 1) is another way in which miR-9 promotes the onset and progression of breast cancer. Breast cancer cells' migration and invasion may be encouraged by miR-9 because it targets forkhead box O1 (FOXO1). This was validated by Liu DZ et al. Ma L et al. [25,26] revealed that the production of miR-9 is triggered by MYC and MYCN, and its overexpression leads to enhanced cell motility and invasiveness by directly binding to the critical metastasis-suppressing protein E-cadherin. Cancer cells become more metastatic when E-cadherin is downregulated, which accomplishes two things: (a) it prepares the cells for epithelial-mesenchymal transition (EMT) and invasion; and (b) it activates -catenin signaling, which increases VEGF expression and thus tumor angiogenesis. Ma L et al. [27] further examined E-cadherin-independent effects of miR-9 and found leukemia inhibitory factor receptor (LIFR) as a miR-9 target in E-cadherin-negative tumor cells and a novel metastasis suppressor, which inhibits metastasis by activating the Hippo-YAP pathway. As a result, LIFR depletion is associated with a dismal prognosis in breast cancer. Previous research from our lab showed that miR-9 promotes EMT and metastasis in breast cancer by focusing on STARD13 [28]. Our findings are supported by those of D'Ippolito E et al. [29], who found that ligand-dependent activation of PDGFR signaling induced endogenous miR-9 expression, which in turn repressed STARD13 and promoted the creation of vascular-like structures in TNBC cells.

5.2. MiR-9 mediates interactions between competing endogenous RNAs (ceRNAs) that control the development of breast cancer.

RNAs with the same miRNA binding sites might cross-talk by competing for miRNAs; this phenomenon was termed ceRNA and added to post-transcriptional suppression of targets by miRNAs. One of the hottest topics in recent years is the ceRNA mechanism, which explains how coding and non-coding genes may mutually control expression by

binding competitively to the same miRNAs [30]. The CeRNA mechanism introduces a novel paradigm for regulating gene expression and a fresh approach to studying the transcriptome, allowing for a more in-depth and thorough explanation of a variety of biological events. As can be shown in Fig. 1, miR-9 may play a mediating role in ceRNA interactions that drive breast cancer growth. 2. Inhibiting the metastasis of breast cancer cells by inducing E-cadherin expression through the miR-9-mediated ceRNA mechanism [31] is one such example. Our previous study confirmed that miR-9 could bind to both the FOXO1- and E-cadherin-3'UTR, and that there was competition of miR-9 between FOXO1- and E-cadherin-3'UTR. Additionally, our earlier findings revealed that CYP4Z1 3'UTR limits the migration and EMT of breast cancer cells by functioning as a ceRNA for E-cadherin, since CYP4Z1 is a possible target of miR-9 too [32]. We further confirmed that the 3'UTRs of CDH5, HOXD1, and HOXD10 suppress breast cancer metastasis by acting as STARD13 ceRNAs, which prevents the miR-9, miR-10b, and miR-125b from doing their jobs [28]. These findings point to a crucial function for miR-9-mediated ceRNA regulatory networks in the development of breast cancer.

5.3. Exosomal miR-9 promotes breast cancer development.

Importantly, miRNA may be transported via exosomes to exercise its effects. Exosomes, which have dimensions between 50 and 150 nm, are a kind of membrane-bound extracellular vesicle. Some molecules, such microRNAs (miRNAs), are packaged within exosomes. More and more data suggests that tumor cells secrete and transmit exosomal miRNAs to interact with both other tumor cells and cells present in the tumor microenvironment. More critically, exosomal miRNAs are shown to act as signaling molecules to control tumor development, angiogenesis, metastasis, susceptibility to chemotherapy, and immune evasion [33]. Tumor-secreted miR-9 may be transported by exosomes to recipient normal fibroblasts (NFs), as shown by S Baroni et al. [34], and this absorption contributes to the conversion of NFs into CAFs, which in turn increases cell motility in breast cancer. Treatment of MCF-7 cells with exosomes from MDA-MB-231 cells leads to the downregulation of target gene PTEN of MCF-7 cells, hence making MCF-7 cells more metastatic, as previously reported by Kia V et al. [35]. Inducing CSC traits and chemotherapy resistance, Shen M et al. [36] found that breast cancer cells secrete multiple microRNAs (miRNAs) into extracellular vesicles (EVs), including miR-9, miR-195, and miR-203a-3p. These miRNAs all target the transcription factor One Cut Homeobox 2 (ONECUT2).

6. The potential of miR-9-based pharmaceuticals

Due to its intricate regulation mechanism, miRNA has the potential to be used as a therapeutic target in a wide range of pathological diseases. A single miRNA may control many target genes, and a single target gene can be targeted by numerous miRNAs. Meanwhile, advancements in RNA molecular delivery technologies have raised the feasibility of treating illness using miRNAs. Just over 20 years after its discovery, miRNA research has moved out of the lab and into the clinic. Phase I clinical studies have been completed successfully, and phase II trials are now under way. The mi34 (MRX34) miRNA analogue is one of the most rapidly expanding areas of cancer therapy. It has shown promising therapeutic results in phase I clinical trials for the treatment of patients with primary liver cancer, small cell lung cancer, lymphoma, melanoma, multiple myeloma, and renal cell carcinoma. In addition, there have been phase I clinical trials of miR-16 analogues in patients with malignant pleural mesothelioma or non-small cell lung cancer, where EGFR antibody-coated EDV nanoparticles are used to deliver miR-16 to cancer tissues and five patients recruited in the first batch achieve some extent of therapeutic effects [37]. Based on the observation that miR-9 is overexpressed in breast cancer, Yu Y et al. developed a cascade miRNA recognition system to selectively kill cancer cells by inducing the expression of a reporter gene (fluorescent protein) or the anticancer gene p53. Nude mice have tumors that are 58% smaller in volume and weight than control mice, indicating that miRNAs may be employed as biomarkers and have

significant promise as a cancer treatment [38]. Given that miR-9 has been shown to sensitize acute myeloid leukemia (AML) to Daunorubicin [39] and that chemotherapy induces breast cancer cells to secrete miR-9-based EV resulting in chemoresistance [36], this raises the intriguing possibility that miR-9 could be combined with other drugs to improve the efficacy of treatment and attenuate drug resistance, or be a predictor of response to chemotherapy in breast cancer.

7. Proliferation of breast cancer cells is inhibited by miR-9.

It is intriguing that miR-9 may have a tumor-suppressing function in breast cancer proliferation regulation. The tumor suppressor-like action of miR-9 in breast cancer cells was shown by Selcuklu SD et al. [40], who found that miR-9 may decrease cell proliferation and increase cell death by directly targeting MTHFD2. Supporting Selcuklu SD's position, Zhao X et al. [41] discovered that the lncRNA taurine-upregulated gene 1 (TUG1) promotes cell proliferation by inhibiting miR-9 and that TUG1 positively regulates the expression of MTHFD2 in MCF-7 cells. Baldassari F et al. [42] also demonstrated that the combination of specific miRNAs, including miR-181a, miR-326, miR-9, and miR-9-2-3p, may be used to better understand the differences between tumoral and peritumoral tissues in breast cancer patients. G Sun et al. [43] found a profile of miRNAs (miR-9, miR-148a, miR-31, miR-375, miR-21, miR-135b, miR-196a, and miR-196b). By directly targeting Her-2, ectopic expression of miR-9 suppressed colony formation and tumor engraftment in breast cancer cells and enhanced their sensitivity to docetaxel (DOC) and cyclophosphamide.

8. Prospectives

Breast cancer is a deadly illness that poses a major risk to human health due to its widespread prevalence among women across the globe. microRNAs are very important in breast cancer progression and survival. MicroRNAs (miRNAs) have the properties of multi-target and efficient regulation since a single miRNA may affect numerous target genes. Using miRNAs as a therapeutic target has the potential to open up new avenues in cancer therapy. An increasing body of research confirms that miR-9 is abnormally expressed in breast cancer. Studies to date have suggested that miR-9 acts as an anti-oncogene in breast cancer proliferation, preventing the disease in its earliest stages, but as an oncogene in the dissemination of more malignant breast cancers. Therefore, the stage of breast cancer determines the miR-9 level and its function in breast cancer.

Cancer biomarker research has increasingly focused on circulating miRNAs due to their small size, stability, and non-invasive access, making them ideal candidates for usage as inexpensive, readily available, and therapeutically applicable molecular biomarkers for diagnosis and prognosis. With the advancement of sequencing technology, miR-9 may be used as a non-invasive tumor marker of breast cancer by measuring miR-9 level in human blood, urine, or bile due to its specificity in expression in breast cancer and its ability to enter the humoral circulation and stability. Further investigation into exosomes may one day allow for their application in the delivery of miRNA-9 inhibitors to breast cancer tissues. Finally, miR-9-knockout mice combined with the spontaneous breast cancer model (MMTV-PyMT) mice can be better used for evaluating the function of miR-9 in breast cancer, and other genetically-modified mice models with miR-9 knockout should be used for this analysis as well.

Funding: N/A

Conflicts of Interest: The authors declare that they have no competing interests

References

- 1) Mahmoudian M, Razmara E, Mahmud Hussen B, Simiyari M, Lotfizadeh N, Motaghd H, Khazraei Monfared A, Montazeri M, Babashah S. Identification of a six-microRNA signature as a potential diagnostic biomarker in breast cancer tissues. *J Clin Lab Anal.* 2021 Nov;35(11):e24010. <https://doi.org/10.1002/jcla.24010> PMID: 34528314; PMCID: PMC8605139.
- 2) Adams BD, Wali VB, Cheng CJ, et al., 2016. miR-34a silences c-SRC to attenuate tumor growth in triple-negative breast cancer. *Cancer Res*, 76(4):927-939. <https://doi.org/10.1158/0008-5472.CAN-15-2321>
- 3) Amorim M, Salta S, Henrique R, et al., 2016. Decoding the usefulness of non-coding RNAs as breast cancer markers. *J Transl Med*, 14:265. <https://doi.org/10.1186/s12967-016-1025-3>
- 4) Anfossi S, Fu X, Nagvekar R, et al., 2018. MicroRNAs, regulatory messengers inside and outside cancer cells. In: Mettinger KL, Rameshwar P, Kumar V (Eds.), *Exosomes, Stem Cells and MicroRNA*. Springer, Cham, p.87-108. https://doi.org/10.1007/978-3-319-74470-4_6
- 5) Atkinson SR, Marguerat S, Bähler J, 2012. Exploring long non-coding RNAs through sequencing. *Semin Cell Dev Biol*, 23(2):200-205. <https://doi.org/10.1016/j.semcdb.2011.12.003>
- 6) Bai XD, Han GH, Liu Y, et al., 2018. MiRNA-20a-5p promotes the growth of triple-negative breast cancer cells through targeting RUNX3. *Biomed Pharmacother*, 103: 1482-1489. <https://doi.org/10.1016/j.biopha.2018.04.165>
- 7) Bayraktar R, Pichler M, Kanlikilicer P, et al., 2017. MicroRNA 603 acts as a tumor suppressor and inhibits triple-negative breast cancer tumorigenesis by targeting elongation factor 2 kinase. *Oncotarget*, 8(7):11641-11658. <https://doi.org/10.18632/oncotarget.14264>
- 8) Bhardwaj A, Singh H, Rajapakshe K, et al., 2017. Regulation of miRNA-29c and its downstream pathways in preneoplastic progression of triple-negative breast cancer. *Oncotarget*, 8(12):19645-19660. <https://doi.org/10.18632/oncotarget.14902>
- 9) Biswas T, Efird JT, Prasad S, et al., 2017. The survival benefit of neoadjuvant chemotherapy and PCR among patients with advanced stage triple negative breast cancer. *Oncotarget*, 8(68):112712-112719. <https://doi.org/10.18632/oncotarget.22521>
- 10) Boon RA, Jaé N, Holdt L, et al., 2016. Long noncoding RNAs: from clinical genetics to therapeutic targets? *J Am Coll Cardiol*, 67(10):1214-1226. <https://doi.org/10.1016/j.jacc.2015.12.051>
- 11) Browne G, Dragon JA, Hong DL, et al., 2016. MicroRNA- 378-mediated suppression of Runx1 alleviates the aggressive phenotype of triple-negative MDA-MB-231 human breast cancer cells. *Tumour Biol*, 37(7):8825-8839. <https://doi.org/10.1007/s13277-015-4710-6>
- 12) Catalanotto C, Cogoni C, Zardo G, 2016. MicroRNA in control of gene expression: an overview of nuclear functions. *Int J Mol Sci*, 17(10):1712. <https://doi.org/10.3390/ijms17101712>
- 13) Chadwick BP, Scott KC, 2013. Molecular versatility: the many faces and functions of noncoding RNA. *Chromosome Res*, 21(6-7):555-559. <https://doi.org/10.1007/s10577-013-9397-1>
- 14) Chen H, Pan H, Qian Y, et al., 2018. MiR-25-3p promotes the proliferation of triple negative breast cancer by targeting BTG2. *Mol Cancer*, 17:4. <https://doi.org/10.1186/s12943-017-0754-0>
- 15) Chen J, Wang BC, Tang JH, 2012. Clinical significance of microRNA-155 expression in human breast cancer. *J Surg Oncol*, 106(3):260-266. <https://doi.org/10.1002/jso.22153>
- 16) Chen JW, Shin VY, Siu MT, et al., 2016. miR-199a-5p confers tumor-suppressive role in triple-negative breast cancer. *BMC Cancer*, 16:887. <https://doi.org/10.1186/s12885-016-2916-7>
- 17) Chen QN, Wei CC, Wang ZX, et al., 2017. Long non-coding RNAs in anti-cancer drug resistance. *Oncotarget*, 8(1): 1925-1936. <https://doi.org/10.18632/oncotarget.12461>
- 18) Chen XW, Zhao M, Huang J, et al., 2018. microRNA-130a suppresses breast cancer cell migration and invasion by targeting FOSL1 and upregulating ZO-1. *J Cell Biochem*, 119(6):4945-4956. <https://doi.org/10.1002/jcb.26739>
- 19) Collignon J, Lousberg L, Schroeder H, et al., 2016. Triple- negative breast cancer: treatment challenges and solutions. *Breast Cancer* (Dove Med Press), 8:93-107. <https://doi.org/10.2147/BCTT.S69488>

- 20) Costa FF, 2005. Non-coding RNAs: new players in eukaryotic biology. *Gene*, 357(2):83-94. <https://doi.org/10.1016/j.gene.2005.06.019>
- 21) De S, Das S, Mukherjee S, et al., 2017. Establishment of twist-1 and TGFBR2 as direct targets of microRNA-20a in mesenchymal to epithelial transition of breast cancer cell-line MDA-MB-231. *Exp Cell Res*, 361(1):85-92. <https://doi.org/10.1016/j.yexcr.2017.10.005>
- 22) Delás MJ, Hannon GJ, 2017. lncRNAs in development and disease: from functions to mechanisms. *Open Biol*, 7(7): 170121. <https://doi.org/10.1098/rsob.170121>
- 23) Deng H, Zhang J, Shi JJ, et al., 2016. Role of long non-coding RNA in tumor drug resistance. *Tumor Biol*, 37(9):11623- 11631. <https://doi.org/10.1007/s13277-016-5125-8>
- 24) Eades G, Wolfson B, Zhang YS, et al., 2015. lincRNA-RoR and miR-145 regulate invasion in triple-negative breast cancer via targeting ARF6. *Mol Cancer Res*, 13(2):330- 338. <https://doi.org/10.1158/1541-7786.MCR-14-0251>
- 25) Eades GL, Zhou Q, 2014. Abstract 1463: long non-coding RNA RoR and microRNA-145 regulate tumor cell invasion in triple-negative breast cancer via targeting of ADP- ribosylation factor 6. *Cancer Res*, 74(S19):1463. <https://doi.org/10.1158/1538-7445.AM2014-1463>
- 26) Evans JR, Feng FY, Chinnaiyan AM, 2016. The bright side of dark matter: lncRNAs in cancer. *J Clin Invest*, 126(8): 2775-2782. <https://doi.org/10.1172/JCI84421>
- 27) Fang H, Xie JP, Zhang M, et al., 2017. miRNA-21 promotes proliferation and invasion of triple-negative breast cancer cells through targeting PTEN. *Am J Transl Res*, 9(3): 953-961.
- 28) Ferlay J, Héry C, Autier P, et al., 2010. Global burden of breast cancer. In: Li C (Ed.), *Breast Cancer Epidemiology*. Springer, New York, p.1-19. https://doi.org/10.1007/978-1-4419-0685-4_1
- 29) Fu PF, Zheng X, Fan X, et al., 2019. Role of cytoplasmic lncRNAs in regulating cancer signaling pathways. *J Zhejiang Univ-Sci B (Biomed & Biotechnol)*, 20(1):1-8. <https://doi.org/10.1631/jzus.B1800254>
- 30) Gebert LFR, MacRae IJ, 2019. Regulation of microRNA function in animals. *Nat Rev Mol Cell Biol*, 20(1):21-37. <https://doi.org/10.1038/s41580-018-0045-7>
- 31) Gilam A, Conde J, Weissglas-Volkov D, et al., 2016. Local microRNA delivery targets Palladin and prevents metastatic breast cancer. *Nat Commun*, 7:12868. <https://doi.org/10.1038/ncomms12868>
- 32) Gu J, Wang YP, Wang XD, et al., 2018. Downregulation of lncRNA GAS5 confers tamoxifen resistance by activating miR-222 in breast cancer. *Cancer Lett*, 434:1-10. <https://doi.org/10.1016/j.canlet.2018.06.039>
- 33) Gülben K, Berberoglu U, Kinaş V, et al., 2014. Breast cancer subtypes can be a predictor of pathologic complete response and survival in the neoadjuvant setting for T4 noninflammatory breast cancer. *Acta Chir Belg*, 114(3): 153-159. <https://doi.org/10.1080/00015458.2014.11681001>
- 34) Gupta RA, Shah N, Wang KC, et al., 2010. Long non-coding RNA HOTAIR reprograms chromatin state to promote cancer metastasis. *Nature*, 464(7291):1071-1076. <https://doi.org/10.1038/nature08975>
- 35) Han JG, Han BJ, Wu XY, et al., 2018. Knockdown of lncRNA H19 restores chemo-sensitivity in paclitaxel-resistant triple-negative breast cancer through triggering apoptosis and regulating Akt signaling pathway. *Toxicol Appl Pharmacol*, 359:55-61. <https://doi.org/10.1016/j.taap.2018.09.018>
- 36) Han JJ, Yu JJ, Dai YN, et al., 2018. Overexpression of miR- 361-5p in triple-negative breast cancer (TNBC) inhibits migration and invasion by targeting RQCD1 and inhibiting the EGFR/PI3K/Akt pathway. *Bosn J Basic Med Sci*, 19(1):52-59. <https://doi.org/10.17305/bjbm.2018.3399>
- 37) Harrow J, Frankish A, Gonzalez JM, et al., 2012. GENCODE: the reference human genome annotation for the encode project. *Genome Res*, 22(9):1760-1774. <https://doi.org/10.1101/gr.135350.111>
- 38) Hata A, Kashima R, 2016. Dysregulation of microRNA biogenesis machinery in cancer. *Crit Rev Biochem Mol Biol*, 51(3):121-134. <https://doi.org/10.3109/10409238.2015.1117054>
- 39) Hiatt RA, Brody JG, 2018. Environmental determinants of breast cancer. *Annu Rev Public Health*, 39:113-133. <https://doi.org/10.1146/annurev-publhealth-040617-014101>
- 40) Hong LQ, Pan F, Jiang HF, et al., 2016. MiR-125b inhibited epithelial-mesenchymal transition of triple-negative breast cancer by targeting MAP2K7. *Oncotargets Ther*, 9: 2639-2648. <https://doi.org/10.2147/OTT.S102713>

- 41) Hu JH, Xu J, Wu YQ, et al., 2015. Identification of microRNA- 93 as a functional dysregulated miRNA in triple-negative breast cancer. *Tumour Biol*, 36(1):251-258. <https://doi.org/10.1007/s13277-014-2611-8>
- 42) Huang J, Zhou N, Watabe K, et al., 2014. Long non-coding RNA UCA1 promotes breast tumor growth by suppression of p27 (Kip1). *Cell Death Dis*, 5:e1008. <https://doi.org/10.1038/cddis.2013.541>
- 43) Huarte M, 2015. The emerging role of lncRNAs in cancer. *Nat Med*, 21(11):1253-1261. <https://doi.org/10.1038/nm.3981>
- 44) Jia ZM, Liu Y, Gao Q, et al., 2016. miR-490-3p inhibits the growth and invasiveness in triple-negative breast cancer by repressing the expression of TNKS2. *Gene*, 593(1):41-47. <https://doi.org/10.1016/j.gene.2016.08.014>
- 45) Karagoz K, Sinha R, Arga KY, 2015. Triple negative breast cancer: a multi-omics network discovery strategy for candidate targets and driving pathways. *OMICS*, 19(2):115- 130. <https://doi.org/10.1089/omi.2014.0135>
- 46) Khaled N, Bidet Y, 2019. New insights into the implication of epigenetic alterations in the EMT of triple negative breast cancer. *Cancers (Basel)*, 11(4):559. <https://doi.org/10.3390/cancers11040559>
- 47) Kim SY, Kawaguchi T, Yan L, et al., 2017. Clinical relevance of microRNA expressions in breast cancer validated using The Cancer Genome Atlas (TCGA). *Ann Surg Oncol*, 24(10):2943-2949. <https://doi.org/10.1245/s10434-017-5984-2>
- 48) Kolesnikov NN, Veryaskina YA, Titov SE, et al., 2019. Expression of microRNAs in molecular genetic breast cancer subtypes. *Cancer Treat Res Commun*, 20:100026. <https://doi.org/10.1016/j.ctarc.2016.08.006>
- 49) Kunej T, Obsteter J, Pogacar Z, et al., 2014. The decalog of long non-coding RNA involvement in cancer diagnosis and monitoring. *Crit Rev Clin Lab Sci*, 51(6):344-357. <https://doi.org/10.3109/10408363.2014.944299>
- 50) Lee J, Jung JH, Chae YS, et al., 2016. Long noncoding RNA snaR regulates proliferation, migration and invasion of triple-negative breast cancer cells. *Anticancer Res*, 36(12): 6289-6295. <https://doi.org/10.21873/anticancer.11224>
- 51) Lehmann BD, Bauer JA, Chen X, et al., 2011. Identification of human triple-negative breast cancer subtypes and preclinical models for selection of targeted therapies. *J Clin Invest*, 121(7):2750-2767. <https://doi.org/10.1172/JCI45014>
- 52) Li HY, Liang JL, Kuo YL, et al., 2017. miR-105/93-3p promotes chemoresistance and circulating miR-105/93-3p acts as a diagnostic biomarker for triple negative breast cancer. *Breast Cancer Res*, 19:133. <https://doi.org/10.1186/s13058-017-0918-2>
- 53) Li J, Chen CC, Ma XC, et al., 2016. Long noncoding RNA NRON contributes to HIV-1 latency by specifically inducing TAT protein degradation. *Nat Commun*, 7:11730. <https://doi.org/10.1038/ncomms11730>
- 54) Li J, Cui ZG, Li H, et al., 2018. Clinicopathological and prognostic significance of long noncoding RNA MALAT1 in human cancers: a review and meta-analysis. *Cancer Cell Int*, 18:109. <https://doi.org/10.1186/s12935-018-0606-z>
- 55) Li N, Deng YJ, Zhou LH, et al., 2019. Global burden of breast cancer and attributable risk factors in 195 countries and territories, from 1990 to 2017: results from the global burden of disease study 2017. *J Hematol Oncol*, 12:140. <https://doi.org/10.1186/s13045-019-0828-0>
- 56) Li SQ, Zhou J, Wang ZX, et al., 2018. Long noncoding RNA GAS5 suppresses triple negative breast cancer progression through inhibition of proliferation and invasion by competitively binding miR-196a-5p. *Biomed Pharmacother*, 104:451-457. <https://doi.org/10.1016/j.biopha.2018.05.056>
- 57) Li WT, Liu CL, Zhao CL, et al., 2016. Downregulation of β 3 integrin by miR-30a-5p modulates cell adhesion and invasion by interrupting Erk/Ets-1 network in triple- negative breast cancer. *Int J Mol Sci*, 48(3):1155-1164. <https://doi.org/10.3892/ijo.2016.3319>
- 58) Li XH, Hou LL, Yin L, et al., 2020. LncRNA XIST interacts with miR-454 to inhibit cells proliferation, epithelial mesenchymal transition and induces apoptosis in triple- negative breast cancer. *J Biosci*, 45:45. <https://doi.org/10.1007/s12038-020-9999-7>
- 59) Li XN, Wu YM, Liu AH, et al., 2016. Long non-coding RNA UCA1 enhances tamoxifen resistance in breast cancer cells through a miR-18a-HIF1 α feedback regulatory loop. *Tumor Biol*, 37(11):14733-14743. <https://doi.org/10.1007/s13277-016-5348-8>

- 60) Li Z, Li Y, Li Y, et al., 2017. Long non-coding RNA H19 promotes the proliferation and invasion of breast cancer through upregulating DNMT1 expression by sponging miR-152. *J Biochem Mol Toxicol*, 31(9):e21933. <https://doi.org/10.1002/jbt.21933>
- 61) Li ZS, Meng QY, Pan AF, et al., 2017. MicroRNA-455-3p promotes invasion and migration in triple negative breast cancer by targeting tumor suppressor EI24. *Oncotarget*, 8(12):19455-19466. <https://doi.org/10.18632/oncotarget.14307>
- 62) Li ZX, Qian J, Li J, et al., 2019. Knockdown of lncRNA- HOTAIR downregulates the drug-resistance of breast cancer cells to doxorubicin via the PI3K/AKT/mTOR signaling pathway. *Exp Ther Med*, 18(1):435-442. <https://doi.org/10.3892/etm.2019.7629>
- 63) Liang YJ, Hu J, Li JT, et al., 2015. Epigenetic activation of TWIST1 by MTDH promotes cancer stem-like cell traits in breast cancer. *Cancer Res*, 75(17):3672-3680. <https://doi.org/10.1158/0008-5472.CAN-15-0930>
- 64) Liedtke C, Mazouni C, Hess K, et al., 2008. Response to neoadjuvant therapy and long-term survival in patients with triple-negative breast cancer. *J Clin Oncol*, 26(8):1275- 1281. <https://doi.org/10.1200/JCO.2007.14.4147>
- 65) Lin AF, Li CL, Xing Z, et al., 2016. The LINK-A lncRNA activates normoxic HIF1 α signalling in triple-negative breast cancer. *Nat Cell Biol*, 18(2):213-224. <https://doi.org/10.1038/ncb3295>
- 66) Liu AN, Qu HJ, Gong WJ, et al., 2019. LncRNA AWPPH and miRNA-21 regulates cancer cell proliferation and chemosensitivity in triple-negative breast cancer by interacting with each other. *J Cell Biochem*, 120(9):14860-14866. <https://doi.org/10.1002/jcb.28747>
- 67) Liu HY, Wang G, Yang LL, et al., 2016. Knockdown of long non-coding RNA UCA1 increases the tamoxifen sensitivity of breast cancer cells through inhibition of Wnt/ β -catenin pathway. *PLoS ONE*, 11(12):e0168406. <https://doi.org/10.1371/journal.pone.0168406>
- 68) Liu L, He J, Wei X, et al., 2017a. MicroRNA-20a-mediated loss of autophagy contributes to breast tumorigenesis by promoting genomic damage and instability. *Oncogene*, 36(42):5874-5884. <https://doi.org/10.1038/onc.2017.193>
- 69) Liu L, Yu DH, Shi H, et al., 2017b. Reduced lncRNA Aim enhances the malignant invasion of triple-negative breast cancer cells mainly by activating Wnt/ β -catenin/mTOR/ PI3K signaling. *Pharmazie*, 72(10):599-603. <https://doi.org/10.1691/ph.2017.7547>
- 70) Liu M, Xing LQ, Liu YJ, 2017. A three-long noncoding RNA signature as a diagnostic biomarker for differentiating between triple-negative and non-triple-negative breast cancers. *Medicine (Baltimore)*, 96(9):e6222. <https://doi.org/10.1097/MD.00000000000006222>
- 71) Liu XP, Tang HL, Chen JP, et al., 2015. MicroRNA-101 inhibits cell progression and increases paclitaxel sensitivity by suppressing MCL-1 expression in human triple- negative breast cancer. *Oncotarget*, 6(24):20070-20083. <https://doi.org/10.18632/oncotarget.4039>
- 72) Luan T, Zhang XM, Wang SY, et al., 2017. Long non-coding RNA MIAT promotes breast cancer progression and functions as ceRNA to regulate DUSP7 expression by sponging miR-155-5p. *Oncotarget*, 8(44):76153-76164. <https://doi.org/10.18632/oncotarget.19190>
- 73) Luo LY, Tang HL, Ling L, et al., 2018. LINC01638 lncRNA activates MTDH-Twist1 signaling by preventing SPOP- mediated c-Myc degradation in triple-negative breast cancer. *Oncogene*, 37(47):6166-6179. <https://doi.org/10.1038/s41388-018-0396-8>
- 74) Luo N, Zhang KJ, Li X, et al., 2020. ZEB1 induced-upregulation of long noncoding RNA ZEB1-AS1 facilitates the progression of triple negative breast cancer by binding with ELAVL1 to maintain the stability of ZEB1 mRNA. *J Cell Biochem*, online. <https://doi.org/10.1002/jcb.29572>
- 75) Lv ZD, Kong B, Liu XP, et al., 2016. miR-655 suppresses epithelial-to-mesenchymal transition by targeting Prrx1 in triple-negative breast cancer. *J Cell Mol Med*, 20(5): 864-873. <https://doi.org/10.1111/jcmm.12770>
- 76) Ma DC, Chen C, Wu J, et al., 2019. Up-regulated lncRNA AFAP1-AS1 indicates a poor prognosis and promotes carcinogenesis of breast cancer. *Breast Cancer*, 26(1):74-83. <https://doi.org/10.1007/s12282-018-0891-3>

- 77) Matamala N, Vargas MT, González-Cámpora R, et al., 2015. Tumor microRNA expression profiling identifies circulating microRNAs for early breast cancer detection. *Clin Chem*, 61(8):1098-1106. <https://doi.org/10.1373/clinchem.2015.238691>
- 78) Mathe A, Scott RJ, Avery-Kiejda K, 2015. miRNAs and other epigenetic changes as biomarkers in triple negative breast cancer. *Int J Mol Sci*, 16(12):28347-28376. <https://doi.org/10.3390/ijms161226090>
- 79) Mattick JS, 2011. The central role of RNA in human development and cognition. *FEBS Lett*, 585(11):1600-1616. <https://doi.org/10.1016/j.febslet.2011.05.001>
- 80) Mattick JS, Makunin IV, 2006. Non-coding RNA. *Hum Mol Genet*, 15(1):R17-R29. <https://doi.org/10.1093/hmg/ddl046> Mattick JS, Amaral PP, Dinger ME, et al., 2009. RNA regulation of epigenetic processes. *BioEssays*, 31(1):51-59. <https://doi.org/10.1002/bies.080099>
- 81) Mayer IA, Abramson VG, Lehmann BD, et al., 2014. New strategies for triple-negative breast cancer—deciphering the heterogeneity. *Clin Cancer Res*, 20(4):782-790. <https://doi.org/10.1158/1078-0432.CCR-13-0583>
- 82) Miao YF, Fan RG, Chen LG, et al., 2016. Clinical significance of long non-coding RNA MALAT1 expression in tissue and serum of breast cancer. *Ann Clin Lab Sci*, 46(4):418- 424.
- 83) Mou EX, Wang H, 2019. LncRNA LUCAT1 facilitates tumorigenesis and metastasis of triple-negative breast cancer through modulating miR-5702. *Biosci Rep*, 39(9): BSR20190489. <https://doi.org/10.1042/BSR20190489>
- 84) Niu LM, Fan QX, Yan M, et al., 2019. LncRNA NRON down- regulates lncRNA snaR and inhibits cancer cell proliferation in TNBC. *Biosci Rep*, 39(5):BSR20190468. <https://doi.org/10.1042/BSR20190468>
- 85) O'Brien K, Lowry MC, Corcoran C, et al., 2015. MiR-134 in extracellular vesicles reduces triple-negative breast cancer aggression and increases drug sensitivity. *Oncotarget*, 6(32):32774-32789. <https://doi.org/10.18632/oncotarget.5192>
- 86) Onyeagucha B, Rajamanickam S, Subbarayalu P, et al., 2016. Abstract P2-03-04: down-regulation of Bcl2-related ovarian killer (BOK) by miR-296-5p protects breast cancer cells from paclitaxel-induced apoptosis. *Cancer Res*, 76(S4): P2-03-04. <https://doi.org/10.1158/1538-7445.SABCS15-P2-03-04>
- 87) Paraskevopoulou MD, Hatzigeorgiou AG, 2016. Analyzing miRNA-lncRNA interactions. In: Feng Y, Zhang L (Eds.), *Long Non-Coding RNAs: Methods and Protocols*. Humana Press, New York, p.271-286. https://doi.org/10.1007/978-1-4939-3378-5_21
- 88) Phan B, Majid S, Ursu S, et al., 2016. Tumor suppressor role of microRNA-1296 in triple-negative breast cancer. *Oncotarget*, 7(15):19519-19530. <https://doi.org/10.18632/oncotarget.6961>
- 89) Piasecka D, Braun M, Kordek R, et al., 2018. MicroRNAs in regulation of triple-negative breast cancer progression. *J Cancer Res Clin Oncol*, 144(8):1401-1411. <https://doi.org/10.1007/s00432-018-2689-2>
- 90) Prensner JR, Chinnaiyan AM, 2011. The emergence of lncRNAs in cancer biology. *Cancer Discov*, 1(5):391-407. <https://doi.org/10.1158/2159-8290.CD-11-0209>
- 91) Razaviyan J, Hadavi R, Tavakoli R, et al., 2018. Expression of miRNAs targeting mTOR and S6K1 genes of mTOR signaling pathway including miR-96, miR-557, and miR-3182 in triple-negative breast cancer. *Appl Biochem Biotechnol*, 186(4):1074-1089. <https://doi.org/10.1007/s12010-018-2773-8>
- 92) Ren Y, Han XD, Yu K, et al., 2014. microRNA-200c downregulates XIAP expression to suppress proliferation and promote apoptosis of triple-negative breast cancer cells. *Mol Med Rep*, 10(1):315-321. <https://doi.org/10.3892/mmr.2014.2222>
- 93) Reshetnikova G, Troyanovsky S, Rimm DL, 2007. Definition of a direct extracellular interaction between Met and E- cadherin. *Cell Biol Int*, 31(4):366-373. <https://doi.org/10.1016/j.cellbi.2007.01.022>
- 94) Rhodes LV, Martin EC, Segar HC, et al., 2015. Dual regulation by microRNA-200b-3p and microRNA-200b-5p in the inhibition of epithelial-to-mesenchymal transition in triple- negative breast cancer. *Oncotarget*, 6(18):16638-16652. <https://doi.org/10.18632/oncotarget.3184>
- 95) Romero-Cordoba SL, Rodriguez-Cuevas S, Rebollar-Vega R, et al., 2016. A microRNA signature identifies subtypes of triple-negative breast cancer and reveals miR-342-3p as regulator of a lactate metabolic

- pathway through silencing monocarboxylate transporter 1. *Cancer Res*, 76(6):A47. <https://doi.org/10.1158/1538-7445.NONRNA15-A47>
- 96) Sha S, Yuan DY, Liu YJ, et al., 2017. Targeting long non-coding RNA DANCR inhibits triple negative breast cancer progression. *Biol Open*, 6(9):1310-1316. <https://doi.org/10.1242/bio.023135>
- 97) Shen X, Zhong JX, Yu P, et al., 2019. YY1-regulated LINC00152 promotes triple negative breast cancer progression by affecting on stability of PTEN protein. *Biochem Biophys Res Commun*, 509(2):448-454. <https://doi.org/10.1016/j.bbrc.2018.12.074>
- 98) Shin VY, Siu MT, Ho JC, et al., 2014. Abstract 531: miR-199a-5p is a biomarker for and regulator of epithelial-mesenchymal transition in triple-negative breast cancer patients. *Cancer Res*, 74(S19):531. <https://doi.org/10.1158/1538-7445.AM2014-531>
- 99) Shin VY, Chen JW, Cheuk IWY, et al., 2019. Long non-coding RNA NEAT1 confers oncogenic role in triple-negative breast cancer through modulating chemoresistance and cancer stemness. *Cell Death Dis*, 10(4):270. <https://doi.org/10.1038/s41419-019-1513-5>
- 100) Shukla GC, Singh J, Barik S, 2011. MicroRNAs: processing, maturation, target recognition and regulatory functions. *Mol Cell Pharmacol*, 3(3):83-92. Siegel RL, Miller KD, Jemal A, 2019. *Cancer statistics, 2019*. *CA Cancer J Clin*, 69(1):7-34. <https://doi.org/10.3322/caac.21551>
- 101) Smith MA, Mattick JS, 2017. Structural and functional annotation of long noncoding RNAs. In: Keith JM (Ed.), *Bioinformatics: Volume II: Structure, Function, and Applications*. Humana Press, New York, p.65-85. https://doi.org/10.1007/978-1-4939-6613-4_4
- 102) Song GQ, Zhao Y, 2015. MicroRNA-211, a direct negative regulator of CDC25B expression, inhibits triple-negative breast cancer cells' growth and migration. *Tumor Biol*, 36(7):5001-5009. <https://doi.org/10.1007/s13277-015-3151-6>
- 103) Song X, Liu ZY, Yu ZY, 2019. LncRNA NEF is downregulated in triple negative breast cancer and correlated with poor prognosis. *Acta Biochim Biophys Sin (Shanghai)*, 51(4):386-392. <https://doi.org/10.1093/abbs/gmz021>
- 104) Sørlie T, 2004. Molecular portraits of breast cancer: tumour subtypes as distinct disease entities. *Eur J Cancer*, 40(18): 2667-2675. <https://doi.org/10.1016/j.ejca.2004.08.021>
- 105) St. Laurent G, Wahlestedt C, Kapranov P, 2015. The landscape of long noncoding RNA classification. *Trends Genet*, 31(5):239-251. <https://doi.org/10.1016/j.tig.2015.03.007>
- 106) Sun WL, Yang YB, Xu CJ, et al., 2017. Regulatory mechanisms of long noncoding RNAs on gene expression in cancers. *Cancer Genet*, 216-217:105-110. <https://doi.org/10.1016/j.cancergen.2017.06.003>
- 107) Sun X, Li YQ, Zheng MZ, et al., 2016. MicroRNA-223 increases the sensitivity of triple-negative breast cancer stem cells to TRAIL-induced apoptosis by targeting HAX-1. *PLoS ONE*, 11(9):e0162754. <https://doi.org/10.1371/journal.pone.0162754>
- 108) Taft RJ, Pang KC, Mercer TR, et al., 2010. Non-coding RNAs: regulators of disease. *J Pathol*, 220(2):126-139. <https://doi.org/10.1002/path.2638>
- 109) Tian T, Wang M, Lin S, et al., 2018. The impact of lncRNA dysregulation on clinicopathology and survival of breast cancer: a systematic review and meta-analysis. *Mol Ther Nucleic Acids*, 12:359-369. <https://doi.org/10.1016/j.omtn.2018.05.018>
- 110) Tse JC, Kalluri R, 2007. Mechanisms of metastasis: epithelial-to-mesenchymal transition and contribution of tumor microenvironment. *J Cell Biochem*, 101(4):816-829. <https://doi.org/10.1002/jcb.21215>
- 111) Tsouko E, Wang J, Frigo DE, et al., 2015. miR-200a inhibits migration of triple-negative breast cancer cells through direct repression of the EPHA2 oncogene. *Carcinogenesis*, 36(9):1051-1060. <https://doi.org/10.1093/carcin/bgv087>
- 112) Verma A, Kaur J, Mehta K, 2019. Molecular oncology update: breast cancer gene expression profiling. *Asian J Oncol*, 1(2):65-72. <https://doi.org/10.4103/2454-6798.173282>
- 113) Wang B, Zhang QY, 2012. The expression and clinical significance of circulating microRNA-21 in serum of five solid tumors. *J Cancer Res Clin Oncol*, 138(10):1659-1666. <https://doi.org/10.1007/s00432-012-1244-9>

- 114) Wang C, Zheng XQ, Shen CY, et al., 2012. MicroRNA-203 suppresses cell proliferation and migration by targeting BIRC5 and LASP1 in human triple-negative breast cancer cells. *J Exp Clin Cancer Res*, 31:58. <https://doi.org/10.1186/1756-9966-31-58>
- 115) Wang H, Tan ZQ, Hu H, et al., 2019. microRNA-21 promotes breast cancer proliferation and metastasis by targeting LZTFL1. *BMC Cancer*, 19:738. <https://doi.org/10.1186/s12885-019-5951-3>
- 116) Wang J, Tsouko E, Jonsson P, et al., 2014. miR-206 inhibits cell migration through direct targeting of the actin-binding protein Coronin 1C in triple-negative breast cancer. *Mol Oncol*, 8(8):1690-1702. <https://doi.org/10.1016/j.molonc.2014.07.006>
- 117) Wang L, Liu DQ, Wu XR, et al., 2018. Long non-coding RNA (LncRNA) RMST in triple-negative breast cancer (TNBC): expression analysis and biological roles research. *J Cell Physiol*, 233(10):6603-6612. <https://doi.org/10.1002/jcp.26311>
- 118) Wang LH, Luan T, Zhou SH, et al., 2019. LncRNA HCP5 promotes triple negative breast cancer progression as a ceRNA to regulate BIRC3 by sponging miR-219a-5p. *Cancer Med*, 8(9):4389-4403. <https://doi.org/10.1002/cam4.2335>
- 119) Wang N, Hou MS, Zhan Y, et al., 2019a. LncRNA PTCSC3 inhibits triple-negative breast cancer cell proliferation by downregulating lncRNA H19. *J Cell Biochem*, 120(9): 15083-15088. <https://doi.org/10.1002/jcb.28769>
- 120) Wang N, Zhong CC, Fu MT, et al., 2019b. Long non-coding RNA HULC promotes the development of breast cancer through regulating LYPD1 expression by sponging miR-6754-5p. *Onco Targets Ther*, 12:10671-10679. <https://doi.org/10.2147/OTT.S226040>
- 121) Wang OC, Yang F, Liu YH, et al., 2017. C-MYC-induced upregulation of lncRNA SNHG12 regulates cell proliferation, apoptosis and migration in triple-negative breast cancer. *Am J Transl Res*, 9(2):533-545.
- 122) Wang PS, Chou CH, Lin CH, et al., 2018. A novel long non-coding RNA linc-ZNF469-3 promotes lung metastasis through miR-574-5p-ZEB1 axis in triple negative breast cancer. *Oncogene*, 37(34):4662-4678. <https://doi.org/10.1038/s41388-018-0293-1>
- 123) Wang SW, Ke H, Zhang HL, et al., 2018. LncRNA MIR100HG promotes cell proliferation in triple-negative breast cancer through triplex formation with p27 loci. *Cell Death Dis*, 9(8):805. <https://doi.org/10.1038/s41419-018-0869-2>
- 124) Wang XL, Chen T, Zhang Y, et al., 2019. Long noncoding RNA Linc00339 promotes triple-negative breast cancer progression through miR-377-3p/HOXC6 signaling pathway. *J Cell Physiol*, 234(8):13303-13317. <https://doi.org/10.1002/jcp.28007>
- 125) Wang XS, Zhang Z, Wang HC, et al., 2006. Rapid identification of UCA1 as a very sensitive and specific unique marker for human bladder carcinoma. *Clin Cancer Res*, 12(16):4851-4858. <https://doi.org/10.1158/1078-0432.CCR-06-0134>
- 126) Wang YX, Zhang ZY, Wang JQ, 2018. MicroRNA-384 inhibits the progression of breast cancer by targeting ACVR1. *Oncol Rep*, 39(6):2563-2574. <https://doi.org/10.3892/or.2018.6385>
- 127) Winton MJ, Igaz LM, Wong MM, et al., 2008. Disturbance of nuclear and cytoplasmic TAR DNA-binding protein (TDP-43) induces disease-like redistribution, sequestration, and aggregate formation. *J Biol Chem*, 283(19): 13302-13309. <https://doi.org/10.1074/jbc.M800342200>
- 128) Wu CH, Luo J, 2016. Long non-coding RNA (lncRNA) urothelial carcinoma-associated 1 (UCA1) enhances tamoxifen resistance in breast cancer cells via inhibiting mtor signaling pathway. *Med Sci Monit*, 22:3860-3867. <https://doi.org/10.12659/msm.900689>
- 129) Wu JL, Shuang ZY, Zhao JF, et al., 2018. Linc00152 promotes tumorigenesis by regulating DNMTs in triple-negative breast cancer. *Biomed Pharmacother*, 97:1275-1281. <https://doi.org/10.1016/j.biopha.2017.11.055>
- 130) Xiong HP, Yan T, Zhang WJ, et al., 2018. miR-613 inhibits cell migration and invasion by downregulating Daam1 in triple-negative breast cancer. *Cell Signal*, 44:33-42. <https://doi.org/10.1016/j.cellsig.2018.01.013>
- 131) Xu ST, Xu JH, Zheng ZR, et al., 2017. Long non-coding RNA ANRIL promotes carcinogenesis via sponging miR-199a in triple-negative breast cancer. *Biomed Pharmacother*, 96:14-21. <https://doi.org/10.1016/j.biopha.2017.09.107>

- 132) Yang CF, Humphries B, Li YF, et al., 2017. Abstract 1468: miR-200b targets ARHGAP18 and suppresses triple negative breast cancer metastasis. *Cancer Res*, 77(S13):1468. <https://doi.org/10.1158/1538-7445.AM2017-1468>
- 133) Yang F, Liu YH, Dong SY, et al., 2016a. Co-expression networks revealed potential core lncRNAs in the triple-negative breast cancer. *Gene*, 591(2):471-477. <https://doi.org/10.1016/j.gene.2016.07.002>
- 134) Yang F, Dong SY, Lv L, et al., 2016b. Long non-coding RNA AFAP1-AS1 was up-regulated in triple-negative breast cancer and regulated proliferation and invasion. *Int J Clin Exp Pathol*, 9(6):6378-6384.
- 135) Yang J, Meng XL, Yu Y, et al., 2019. LncRNA POU3F3 promotes proliferation and inhibits apoptosis of cancer cells in triple-negative breast cancer by inactivating caspase 9. *Biosci Biotechnol Biochem*, 83(6):1117-1123. <https://doi.org/10.1080/09168451.2019.1588097>
- 136) Yoon MK, Mitrea DM, Ou L, et al., 2012. Cell cycle regulation by the intrinsically disordered proteins p21 and p27. *Biochem Soc Trans*, 40(5):981-988. <https://doi.org/10.1042/bst20120092>
- 137) Youness RA, Hafez HM, Khallaf E, et al., 2019. The long noncoding RNA sONE represses triple-negative breast cancer aggressiveness through inducing the expression of miR-34a, miR-15a, miR-16, and let-7a. *J Cell Physiol*, 234(11):20286-20297. <https://doi.org/10.1002/jcp.28629>
- 138) Yu FS, Wang L, Zhang BW, 2019. Long non-coding RNA DRHC inhibits the proliferation of cancer cells in triple negative breast cancer by downregulating long non-coding RNA HOTAIR. *Oncol Lett*, 18(4):3817-3822. <https://doi.org/10.3892/ol.2019.10683>
- 139) Zhang H, Li BW, Zhao HB, et al., 2015. The expression and clinical significance of serum miR-205 for breast cancer and its role in detection of human cancers. *Int J Clin Exp Med*, 8(2):3034-3043.
- 140) Zhang KJ, Luo ZL, Zhang Y, et al., 2016. Circulating lncRNA H19 in plasma as a novel biomarker for breast cancer. *Cancer Biomark*, 17(2):187-194. <https://doi.org/10.3233/CBM-160630>
- 141) Zhang KM, Liu P, Tang HL, et al., 2018. AFAP1-AS1 promotes epithelial-mesenchymal transition and tumorigenesis through Wnt/ β -catenin signaling pathway in triple-negative breast cancer. *Front Pharmacol*, 9:1248. <https://doi.org/10.3389/fphar.2018.01248>
- 142) Zhang R, Xia LQ, Lu WW, et al., 2016. LncRNAs and cancer. *Oncol Lett*, 12(2):1233-1239. <https://doi.org/10.3892/ol.2016.4770>
- 143) Zhang YY, He Q, Hu ZY, et al., 2016. Long noncoding RNA LINP1 regulates repair of DNA double-strand breaks in triple-negative breast cancer. *Nat Struct Mol Biol*, 23(6): 522-530. <https://doi.org/10.1038/nsmb.3211>
- 144) Zhao D, Besser AH, Wander SA, et al., 2015. Cytoplasmic p27 promotes epithelial-mesenchymal transition and tumor metastasis via STAT3-mediated TWIST1 upregulation. *Oncogene*, 34(43):5447-5459. <https://doi.org/10.1038/onc.2014.473>
- 145) Zhao M, Ding XF, Shen JY, et al., 2017. Use of liposomal doxorubicin for adjuvant chemotherapy of breast cancer in clinical practice. *J Zhejiang Univ-Sci B (Biomed & Biotechnol)*, 18(1):15-26. <https://doi.org/10.1631/jzus.B1600303>
- 146) Zhao ZT, Li L, Du PN, et al., 2019. Transcriptional downregulation of miR-4306 serves as a new therapeutic target for triple negative breast cancer. *Theranostics*, 9(5):1401-1416. <https://doi.org/10.7150/thno.30701>
- 147) Zheng LH, Zhang YH, Fu YJ, et al., 2019. Long non-coding RNA MALAT1 regulates BLCAP mRNA expression through binding to miR-339-5p and promotes poor prognosis in breast cancer. *Biosci Rep*, 39(2):BSR20181284. <https://doi.org/10.1042/BSR20181284>
- 148) Zuo YG, Li Y, Zhou ZY, et al., 2017. Long non-coding RNA MALAT1 promotes proliferation and invasion via targeting miR-129-5p in triple-negative breast cancer. *Biomed Pharmacother*, 95:922-928. <https://doi.org/10.1016/j.biopha.2017.09.005>