

Development of the Miniature Robotic Electrodialysis (MR ED) System for Small-Scale Desalting of Liquid Samples with Recovery of Organics

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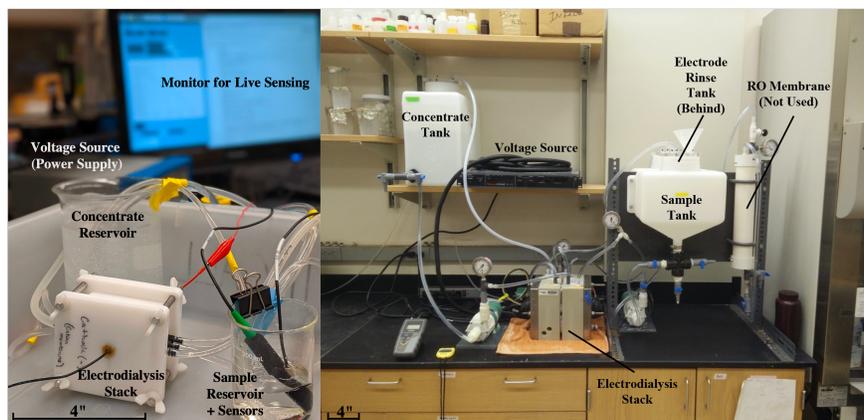
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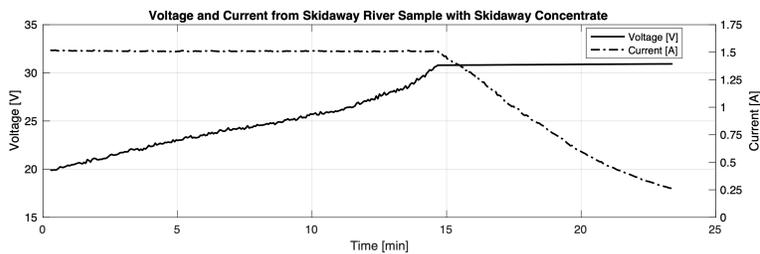
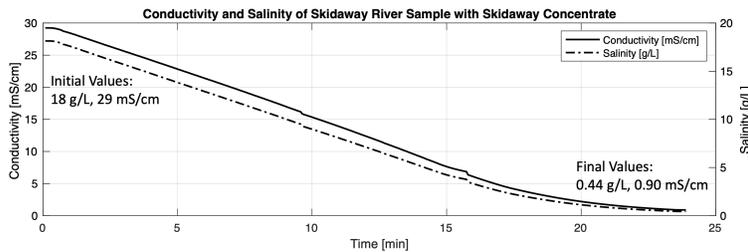
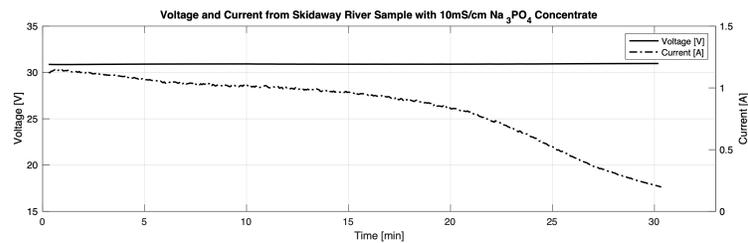
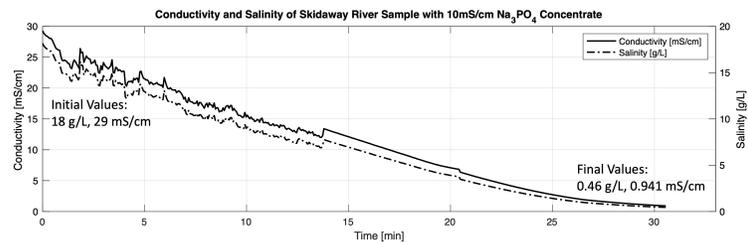
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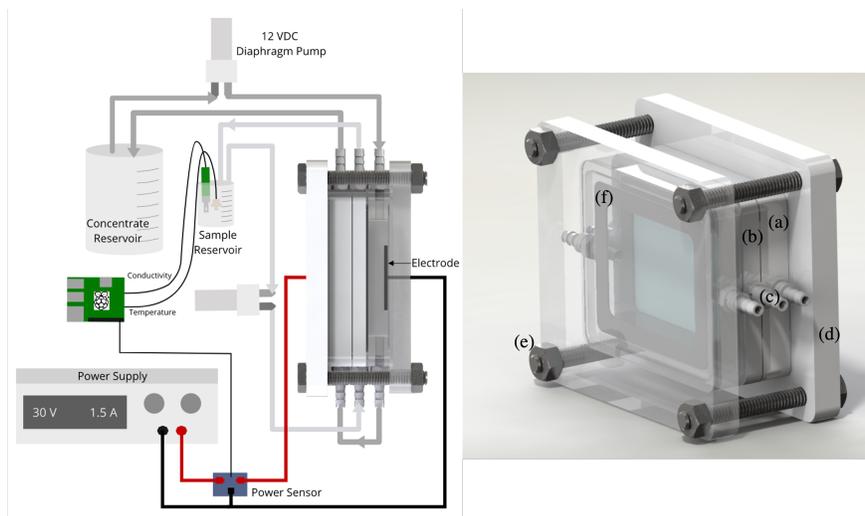
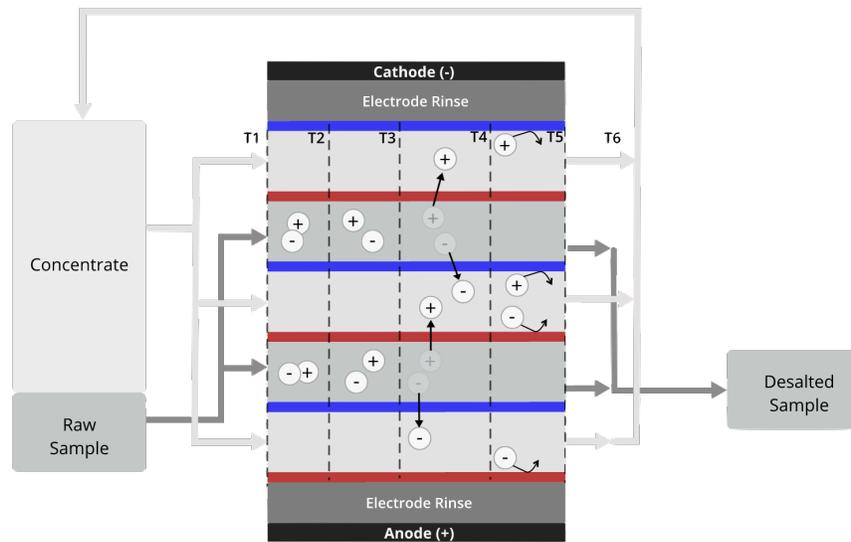
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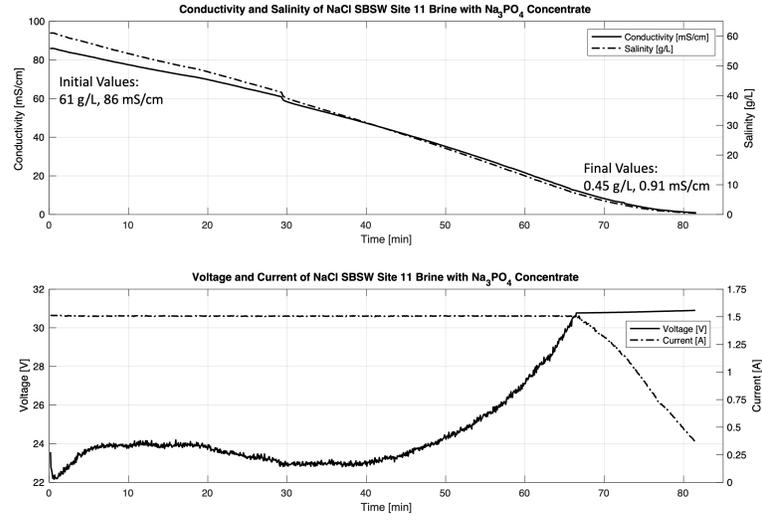
Abstract

While liquid environments with high salt content are of broad interest to the Earth and Planetary Science communities, instruments face challenges in detecting organics in hypersaline samples due to the effects of salts. Therefore, technology to desalt samples before analysis by these instruments would be enabling for liquid sampling on missions to Mars or ocean worlds. Electrodialysis (ED) removes salt from aqueous solutions by applying an electric potential across a series of ion-selective membranes, and is demonstrated to retain a significant percentage of dissolved organic molecules (DOM) in marine samples. However, current electrodialysis systems used for DOM recovery are too large for deployment on missions or for use in terrestrial fieldwork. Here we present the design and evaluation of the Miniature Robotic Electrodialysis (MR ED) system, which is approximately 1/20th the size of heritage instruments and processes as little as 50 mL of sample at a time. We present tests of the instrument efficiency and DOM recovery using lab-created solutions as well as natural samples taken from an estuary of the Skidaway River (Savannah, GA) and from South Bay Saltworks (San Diego, CA). Our results show that the MR ED system removed 97-99% of the salts in most samples, with an average DOC recovery range from 53 to 77%, achieving similar capability to tabletop instruments. This work both demonstrates MR ED as a possible field instrument and increases the technology readiness level of miniaturized electrodialysis systems for future missions.









Abstract

While liquid environments with high salt content are of broad interest to the Earth and Planetary Science communities, instruments face challenges in detecting organics in hypersaline samples due to the effects of salts. Therefore, technology to desalt samples before analysis by these instruments would be enabling for liquid sampling on missions to Mars or ocean worlds. Electrodialysis (ED) removes salt from aqueous solutions by applying an electric potential across a series of ion-selective membranes, and is demonstrated to retain a significant percentage of dissolved organic molecules (DOM) in marine samples. However, current electrodialysis systems used for DOM recovery are too large for deployment on missions or for use in terrestrial fieldwork. Here we present the design and evaluation of the Miniature Robotic Electrodialysis (MR ED) system, which is approximately 1/20th the size of heritage instruments and processes as little as 50 mL of sample at a time. We present tests of the instrument efficiency and DOM recovery using lab-created solutions as well as natural samples taken from an estuary of the Skidaway River (Savannah, GA) (Verity, 2002) and from South Bay Saltworks (San Diego, CA) (Survey, 2011; Roseman & Watry, 2008). Our results show that the MR ED system removed 97-99% of the salts in most samples, with an average DOC recovery range from 53 to 77%, achieving similar capability to tabletop instruments. This work both demonstrates MR ED as a possible field instrument and increases the technology readiness level of miniaturized electrodialysis systems for future missions.

Plain Language Summary

Liquids on other planetary bodies, such as Mars or the icy moons of the outer planets, are important sampling targets for the search for life. Salts help preserve these liquids but can clog small fluidic systems and alter and inhibit the capabilities of precision chemical measurement instruments. Therefore, a key technology development for liquid sampling on ocean worlds is a robust system to desalt samples before they are analyzed by these instruments. Electrodialysis (ED) is a process that removes salts from a sample using a voltage applied across charged membranes to separate the salts' ions from the solution. It has been used in laboratory systems to desalt aqueous solutions while recovering the dissolved organic carbon that would be desirable to measure after the process. However, current systems require further miniaturization and autonomy development to be suitable for deployment on spacecraft. We present the Miniature Robotic Electrodialysis (MR ED) system that has successfully removed 97-99% of the salts in samples and recovered between 53 to 77% of the dissolved organic matter, which is comparable to larger commercial systems at approximately 5% the size.

1 Introduction

The search for life in our solar system, both past and extant, is a primary goal of NASA missions (National Research Council, 2011). Targets of interest include subsurface habitable niches on Mars (Jakosky et al., 2003; Westall et al., 2013), and "ocean worlds" such as the moons Europa, Enceladus, and Ganymede (Hendrix et al., 2018). Potential biologically-relevant materials such as carbon-bearing compounds and even possible metabolic biproducts have been detected or implicated on Mars (Li et al., 2015; Niles et al., 2013; Wray et al., 2016), Europa (Carlson et al., 2009) and Enceladus (Glein et al., 2015; Glein & Waite, 2020; Waite et al., 2017). Particularly for the ocean worlds, future in situ missions would greatly enhance our understanding of the composition of their surfaces and oceans (Hendrix et al., 2018; Lunine, 2017). This increasingly drives interest in developing instruments and sample handling systems that are capable of interrogating these worlds for evidence of habitable environments and life (Committee on the Planetary Science and Astrobiology Decadal Survey et al., 2022).

64 Organic materials are the building blocks of life on Earth and are therefore their
65 detection is highly prioritized on both active and planned spacecraft missions. Detect-
66 ing organics in planetary environments has proved nonetheless challenging. Since plan-
67 etary environments such as Mars' surface and Europa's oceans may have low bioburden,
68 methods to enhance the signal from organics are needed. Amongst the most difficult chal-
69 lenges may be the confounding effects of salts. For example, modern evidence for brines
70 on Mars is accompanied by the detection of perchlorate salts (Hecht et al., 2009; Ojha
71 et al., 2015), and geologic evidence for ancient salty, acidic environments abounds (Rapin
72 et al., 2019; Wang et al., 2018). Analyses completed by the Sample Analysis at Mars (SAM)
73 have used a mass spectrometer and gas chromatograph to search for organic species; how-
74 ever the presence of perchlorates in Martian soil has been theorized to have affected their
75 detection (Franz et al., 2014; Mahaffy et al., 2012). Perchlorates can promote the com-
76 bustion of organic compounds under high temperatures, which occur during the gas pro-
77 cessing of solid samples (Li et al., 2015; Mahaffy et al., 2012; Navarro-González et al.,
78 2006; ten Kate, 2010) which may confound measurements of organic molecules and/or
79 destroy or alter such molecules in situ (S. A. Benner et al., 2000; Lewis et al., 2021).

80 Salts may also confound detection of organics on ocean worlds. Europa is the tar-
81 get of the Europa Clipper (Howell & Pappalardo, 2020) and Jupiter Icy Moons Explorer
82 (Witasse & JUICE Teams, 2020) as well as possible surface missions that would char-
83 acterize the composition of the surface materials and search for organics (Hand et al.,
84 2021, 2017) and subsurface mission concepts that seek to access and explore the global
85 liquid water ocean beneath Europa's thick ice shell (Kivelson et al., 2000; Bryson et al.,
86 2020; B. Schmidt et al., 2021; Stone et al., 2018; Zacny et al., 2018). Predictions for the
87 salt content and composition in Europa's ice shell and ocean are varied but include NaCl
88 and MgSO₄ salts (Carlson et al., 2009; McCord, 2000; Trumbo et al., 2019) at brack-
89 ish to saturated concentration (Buffo et al., 2020; Chivers et al., 2021; Hand & Chyba,
90 2007; Kivelson et al., 2000; B. E. Schmidt, 2020; Zolotov & Shock, 2004; Zolotov & Kargel,
91 2009). In addition to the ocean, there is potential for pockets of brine within the ice shell
92 that could vary strongly in salinity depending upon their detailed evolution and age (Chivers
93 et al., 2021; Collins & Nimmo, 2009; B. E. Schmidt et al., 2011). Potential preservation
94 of organic molecules within these brines makes them a target of high interest for life de-
95 tection (Bryson et al., 2020; Fisher et al., 2021; Lawrence et al., 2021; Merlini et al., 2018),
96 however their salt content presents challenges to instruments for detecting organics.

97 Increasingly, sampling and sample handling systems for planetary missions that seek
98 to detect evidence of life include multiple "phases" of measurements by suites of instru-
99 mentation with increasing sample processing complexity; some instruments require lit-
100 tle to no processing, whereas others require extensive processing (Hand et al., 2021, 2017;
101 Mahaffy et al., 2012). This latter category of instruments includes mass spectrometry
102 and nanopore sequencing, both of which require direct contact and interaction with a
103 sample within the spacecraft (Hand et al., 2021, 2017; Lawrence et al., 2021). In order
104 to measure organics and search for other biomarkers, future missions will require the abil-
105 ity to both concentrate material and remove confounding salts that can alter such chem-
106 ical measurements, in particular mass spectrometry that is a cornerstone of landed mis-
107 sions (Franz et al., 2014; Grubisic et al., 2021; Hand et al., 2017). Compositional mea-
108 surements with mass spectrometry involve ionizing the sample molecules, which is com-
109 plicated by high concentrations of background ions in the sample, such as salts. Mass
110 spectrometers have a limited volume of ions that can be analyzed, and unwanted salt
111 ions can effect interactions that can diminish the sensitivity and accuracy of the instru-
112 ment, and reduce the ability of the mass spectrometer to detect and quantify molecules
113 of interest (Zeichner et al., 2022). Salts can also impede the ionization of biological molecules
114 through suppression or breakdown of the molecule during ionization (Donnelly et al., 2019;
115 Duncan et al., 2019; Metwally et al., 2015). Moreover, as in situ sequencing of samples
116 becomes possible onboard spacecraft, extraction efficiency of DNA by nanopore sequencers
117 is greatly reduced in solutions with high salt concentration (Weng et al., 2019). Thus

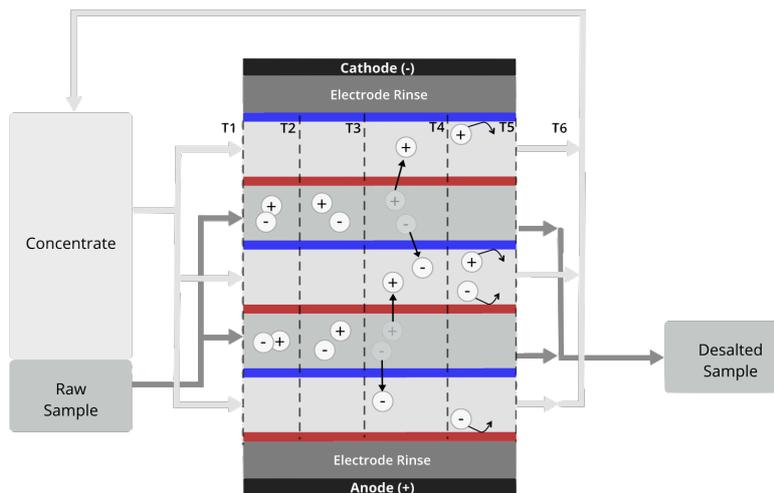


Figure 1. An illustration of the electrodesialysis process. Electrodes on either side of the stack provide the voltage potential and are in contact with a circulated electrode rinse. (T1) Sample and concentrate solutions are fed into the electrodesialysis stack. (T2) These solutions alternate between anion-selective membranes (blue) and cation selective membranes (red). (T3) Ionic salt compounds are split by the voltage potential applied across the stack. (T4) Ions are pulled across their respective membranes into the concentrate channels. For example, positively charged ions are drawn towards the negatively charged cathode through the cation-selective membranes. Similarly, negatively charged anions are pulled through the anion-selective membranes towards the anode. (T5) The positively charged ions are trapped in the concentrate channel as they cannot pass through the anion-selective membrane; negatively charged ions are similarly trapped as they cannot pass through the cation-selective membrane. (T6) The concentrate channels are circulated with an external reservoir, and the desalted sample is either circulated for further desalting or removed from the system. The arrangement of membranes, flow channels and electrodes deplete the sample of dissolved salts during sample processing.

118 it has been recommended that methods to remove salts in milliliter to microliter-scale
 119 samples while preserving organic compounds within the sample be developed, in order
 120 to create water samples suitable for analytical life detection techniques that are confounded
 121 by high salinities (Lawrence et al., 2021).

122 On Earth, characterizing the abundance of organics in the ocean is greatly improved
 123 by desalting (Gurtler et al., 2008; Mopper et al., 2007). Electrodesialysis (ED) is a tech-
 124 nique employed by oceanographers to remove salt while retaining and concentrating or-
 125 ganic molecules of interest for analysis. Electrodesialysis uses an electric potential applied
 126 across a stack of separated flow channels typically containing two different aqueous so-
 127 lutions — a ‘sample’ solution from which ions are removed and a ‘concentrate’ solution
 128 into which ions travel. These channels are separated by alternating anion and cation ex-
 129 change membranes, as shown in Figure 1, which allow negatively and positively charged
 130 ions to pass through, respectively, while preventing movement of oppositely charged ions.
 131 The electric potential across the alternating membranes pulls the ions from the sample
 132 flow channel into the adjacent flow channel where the arrangement of the membranes
 133 traps the ions in the receiving solution. Typically, ED systems also have an additional
 134 flow channel that circulates a solution in contact with the cathode and anode, called the

135 electrode rinse. This additional channel reduces unwanted interactions between the salts
136 pulled from the sample solution and the electrodes.

137 Electro dialysis is ideally suited for sample preparation as it has been shown to ef-
138 ficiently remove salts from seawater and brines while retaining a high percentage of dis-
139 solved organic molecules in laboratory-scale setups. Recovery of organic molecules is typ-
140 ically measured via the dissolved organic carbon (DOC) concentration of the sample be-
141 fore and after processing with electro dialysis (R. Benner & Strom, 1993; Grasshoff et al.,
142 2009). In coastal seawater samples and phytoplankton cultures 70% of the DOC is re-
143 tained in samples processed with laboratory-scale electro dialysis systems, while 99.7%
144 of the salts are removed (Chambers et al., 2016; Gurtler et al., 2008; Koprivnjak et al.,
145 2006; Vetter et al., 2007), although lab-created samples show that some organic molecules
146 (eg. glucose, vitamin B12) can near complete recovery after electro dialysis (>90%) (Chambers
147 et al., 2016). This recovery is typically much better than other techniques to recover or-
148 ganic molecules from seawater such as extraction resins ($\approx 50\%$) or ultrafiltration ($\approx 30\%$)
149 (Chambers et al., 2016). Combined reverse osmosis/electro dialysis processing has been
150 proven to concentrate and desalt samples on a much larger volume scale (200 L) while
151 retaining between 60-90% DOC in seawater samples (Vetter et al., 2007). The advan-
152 tage of using reverse osmosis (RO) is to decrease the sample volume to concentrate the
153 organic material; however, on smaller volumes electro dialysis alone can recover as much
154 DOC as the combined process. These successful electro dialysis systems are large and have
155 been used to process between 0.5 L and 200 L of sample; however, miniaturized systems
156 require further development to achieve similar recovery, and require design changes to
157 function as a part of a science package deployment on a spacecraft. Minimization of sam-
158 ple volumes has many advantages including reduction of instrument size and power re-
159 quirements if the science value can be preserved. Additionally, miniaturization is advan-
160 tageous for making the process field portable for investigations in remote locations on
161 Earth by reducing the logistics of collecting large volumes of sample. Field sampling and
162 analysis has relevance in many environments, such as deployment on oceanographic in-
163 vestigations, as well as investigating samples from hypersaline environments, which have
164 the potential to be analogs for ocean worlds (Buffo et al., 2021; Klempay et al., 2021);
165 however, their high concentrations of salts are detrimental to instrument processing.

166 The handling of liquid samples is the next frontier of planetary exploration. Given
167 the prevalence of compelling planetary environments to explore that contain moderate
168 to high concentrations of salts, the development of desalting as a part of sample prepa-
169 ration is critical. Relevant to planetary exploration, laboratory electro dialysis systems
170 developed for use on Earth are large and require manual operation, and thus, are not
171 well-suited to fly on a planetary mission. To realize the organic preservation of ED sys-
172 tems on a scale appropriate for planetary missions, we designed a miniaturized, robotic
173 electro dialysis system (MR ED), and tested the instrument to increase its technology
174 readiness level (TRL) for consideration on a future mission.

175 2 Materials and Methods

176 We sought to develop a miniaturized electro dialysis system that removes salts from
177 a sample solution while minimally disrupting organics and maintains a compact design
178 to progress towards desalting in challenging planetary environments. To meet these goals,
179 we implemented a design shown in Figure 2. The Miniature Robotic Electro dialysis sys-
180 tem uses a single cell membrane pair; that is, there is one pair of ion-exchange membranes
181 and only one sample flow chamber in the system. The exchange membranes separate a
182 sample channel from two combined concentrate/electrode rinse channel, as shown in Fig-
183 ure 2. The channels are made of machined Delrin and 3D-printed UV-sensitive resin, which
184 is inert after curing and allows for printing unique shapes such as interior fluid routing
185 and hose barb connectors. The sample chamber holds approximately 25 mL, and the com-
186 bined electrode rinse/concentrate chambers hold approximately 40 mL of fluid. Rather

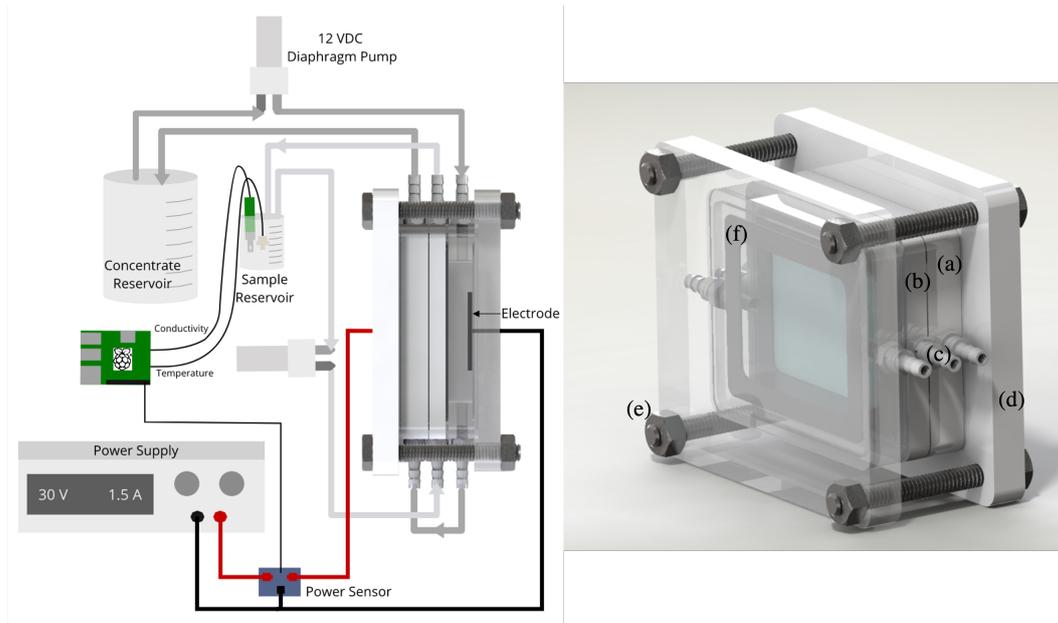


Figure 2. The Miniaturized Robotic ElectroDialysis system. To the left, a schematic of the MR ED setup. Excess concentrate and sample are contained in separate reservoirs, with conductivity and temperature sensors in the sample reservoir. Pumps continuously circulate the fluids, and a power supply provides the voltage potential across the electro dialysis stack. To the right, a model of the MR ED system, which holds approximately 40 mL in the combined electrode rinse/ concentrate chambers (a) and 25 mL in its sample chamber (b). Hose barb connectors (c) thread into the separate channels to allow fluid routing. The assembly is held together using the endcaps (d) through which electrodes access using thru holes, and threaded screws and nuts (e) to maintain an adjustable tight seal. Between each piece in the assembly there is rubber gasket material (f) to prevent leaks and ion-exchange membranes between each concentrate chamber and the sample chamber.

187 than use separate electrode rinse channels, MR ED's electrodes are in direct contact with
188 the concentrate. This follows from MR ED's intended applicability for use on a plan-
189 etary mission for which mass must be conserved, and thus a separate volume of electrode
190 rinse is not required. To avoid corrosion of the electrodes, the electrodes are a platinized
191 titanium mesh, which is resistant to corrosion and oxidation (Hayfield, 1983).

192 Ion-exchange membranes are used between each chamber to selectively allow only
193 cations or anions to pass. The ion-exchange membranes used here are anion Fumasep
194 FAS-PET-130 and cation Fumasep FKS-PET-130 (*Fumasep - ion exchange membranes
195 for water treatment*, 2020), and are laser cut to fit MR ED's form factor. The selection
196 of ion exchange membrane is driven by parameters including the membrane's thickness,
197 permselectivity, and area resistance. The thickness of the membrane affects its conduct-
198 ivity as well as its susceptibility to adsorption of organics, or fouling, which can increase
199 the membrane's resistance and thus reduce its transfer efficiency (Lindstrand et al., 2000).
200 The membrane's permselectivity is a measure of its ability to differentiate between an-
201 ions and cations (Luo et al., 2018), and the area resistance is a measure of how much cur-
202 rent can flow across the membrane in the presence of a voltage potential (Galama et al.,
203 2016). The membranes we used with MR ED have heritage use in laboratory-scale elec-
204 trodialysis systems, and have high specific area resistance and selectivity. They are re-
205 inforced with polyester, which increases the thickness compared to similar membranes,
206 but provides resistance to degradation by acids, bases, and oxidation.

207 To assemble the system as shown in Figure 2, the chambers are stacked with mem-
208 branes between them and rubber gaskets on either side of each membrane. Rubber gas-
209 kets are additionally used against the endplates to seal the concentrate channels. Elec-
210 trodes, which are connected to the endplates with an epoxy-sealed hole to allow a wire
211 through the endplate, are connected to the power supply. Four threaded rods connect
212 the endplates with thumb screws on either end, so that the assembly is tightly but ad-
213 justably sealed. Hose barb connectors screw into each chamber to connect to the tub-
214 ing that routes the fluids. The sample fluid is routed using silicon tubing from its mon-
215 itoring reservoir to a 12 VDC diaphragm pump, which circulates the sample to the sam-
216 ple chamber and back to the reservoir. The concentrate fluid is first routed from its reser-
217 voir beaker through a second pump, then to the concentrate chambers in series before
218 it returns to the reservoir. Circulating the fluid in this way greatly increases the efficiency
219 of desalting by electrodialysis by keeping the fluid well-mixed. A well-mixed fluid keeps
220 the ionic composition near the membrane surfaces replete with ions. In contrast, in static
221 fluid ions are rapidly depleted at membrane surface which decreases desalting efficiency
222 and results in the hydrolysis of water molecules that impacts sample pH.

223 We selected a Raspberry Pi computer to record and display information gathered
224 from the sensors using a custom graphic user interface. Using a Raspberry Pi allows for
225 further development to increase the system's autonomy or sensing capabilities while main-
226 taining a compact footprint. It additionally provides convenient interface to the sensors.
227 MR ED uses a temperature sensor (AtlasScientific Micro PT-1000 Temperature Probe)
228 and conductivity sensor (AtlasScientific Mini Conductivity Probe K 1.0) to monitor the
229 salinity of the sample as well as an Adafruit INA260 sensor to measure the voltage and
230 current applied across the cells. Monitoring the voltage and current throughout process-
231 ing allows the operator to see issues in the processing that is hidden inside the assem-
232 bly; for example, precipitating salts result in noticeable difference in the power response
233 compared to that of a smoothly running process. The sensing locations can be seen in
234 the schematic in Figure 2. We used a BK Precision DC Regulated 1670A benchtop power
235 supply capable of a 30 V output. We imposed a limit of 1.5 A using the power supply
236 to avoid exceeding the limiting current density of the membranes, above which increased
237 potential does not increase the transport of ions across the membranes, and operation
238 can damage the membranes.

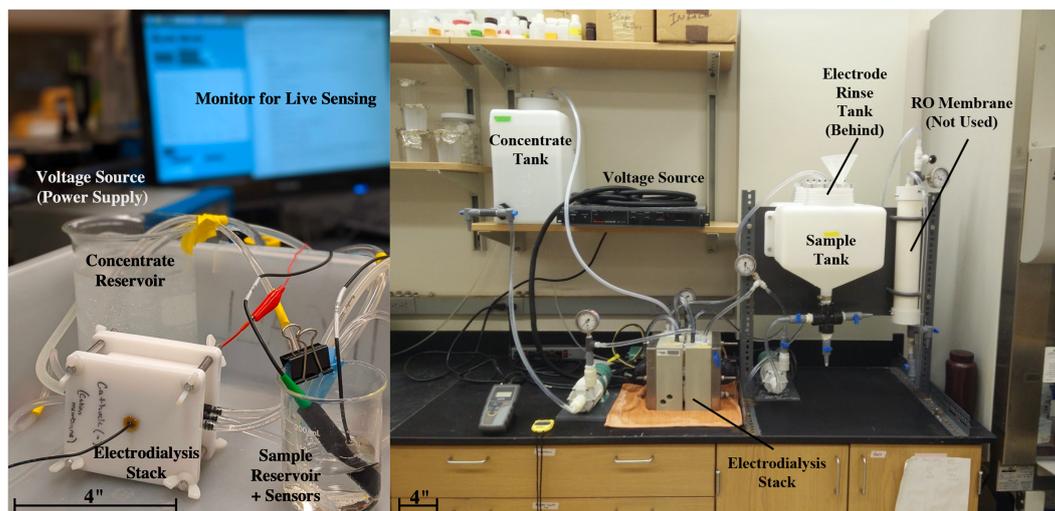


Figure 3. A comparison of the MR ED system with its contemporary, from Chambers et al. (2016). MR ED processes between 50 and 100 mL of sample, whereas the larger system has been used to process between 0.5 and 200 L of sample. The similar components of the two systems are labeled for comparison. MR ED is approximately 1/20 the size of its laboratory contemporary, based on the sum of the volumes of the systems' components.

239 The testing process for each type of sample and concentrate, which are detailed in
 240 Table 1, begins with an initial circulation of the sample and concentrate prior to apply-
 241 ing power to the electrodes. The sample is then emptied from the reservoir and circu-
 242 lated twice before the final circulation. This is done to rinse the membranes and tub-
 243 ing with the new solutions – organic content may be absorbed onto the membrane dur-
 244 ing this circulation or may be dislodged from absorption during a previous test, so cir-
 245 culation is necessary before the initial DOC analysis. After circulation, four milliliters
 246 are taken from the sample and concentrate for the initial DOC analysis before power is
 247 applied. We employ a Shimadzu TOC-VCSN total organic carbon analyzer to analyze
 248 the DOM recovery of each sample processing run, which measures the organic carbon
 249 content in a sample taken from the solutions (R. Benner & Strom, 1993; Grasshoff et al.,
 250 2009). This provides a starting dissolved organic carbon (DOC) concentration to be used
 251 for comparison for recovery. Samples with low expected DOC content are analyzed undi-
 252 luted, while samples with higher expected DOC content, such as natural samples, are
 253 diluted up to 40-fold before analysis to be better conditioned for the instrument. The
 254 power is applied to the system until the sample's conductivity falls below 1 mS/cm, which
 255 corresponds to ~ 8 mM for NaCl solutions, after which four milliliters of sample and
 256 concentrate are once again taken for the final DOC analysis.

257 MR ED's design focus is the miniaturization of an electro dialysis system for con-
 258 sideration for use on a future instrument payload. The benchtop system we have created
 259 is a fraction of the size of established systems, as exhibited in Figure 3, and has an added
 260 benefit of low complexity and using cells that are either 3D printed or are easy-to-machine
 261 pieces makes the system easily replicable, and thus makes it accessible for a wide range of
 262 uses. The design allows flexibility in the system for adding additional cell pairs to in-
 263 crease sample volume or desalting efficiency, or to include an additional electrode rinse
 264 channel. Further miniaturization of the system can be achieved by reducing the size of
 265 the chambers, which will reduce fluid volumes in each channel. To improve the desalt-
 266 ing efficiency, we have included constant circulation of all fluid to promote mixing of the
 267 ion-depleted and ion-enriched sample nearest to the membranes, and the thick channels

Table 1. Types of solution used for the sample and concentrate during testing

Solution Description	Solution Name	Notes
0.5 N NaCl, 200 ppm glucose	Sample 1	Lab-created
Skidaway River (GA, USA)	Sample 2	Estuarine seawater
South Bay Salt Works Site 11 Brine	Sample 3	Primary salt is NaCl
South Bay Salt Works Site 5 Brine	Sample 4	8x Dilution with MilliQ water, primary salt is MgCl ₂
10 mS/cm Na ₃ PO ₄	Concentrate 1	Lab-created
20 mS/cm Na ₃ PO ₄	Concentrate 2	Lab-created

268 of fluid further promote this mixing. We eliminated the use of the electrode rinse to re-
 269 duce the complexity specifically with the motivation for planetary mission use, which ad-
 270 ditionally improves the terrestrial field-portability of the device. With the ambition of
 271 the device being used autonomously, we included a suite of sensors connected to a Rasp-
 272 berry Pi, which can be used in the future to control an integrated power system and pumps.
 273 We have tested the device with as little as 50 mL of sample, which is more than 10 times
 274 smaller than that of commercially available systems (Chambers et al., 2016; Koprivn-
 275 jak et al., 2006).

276 3 Results and Discussion

277 We designed the Miniature Robotic Electrodialysis system to desalt samples rep-
 278 resentative of those that would be expected on Europa. In the absence of specific salin-
 279 ity measurements on Europa, we tested the system with samples with composition sim-
 280 ilar to Earth seawater to samples many times more saline than seawater. Key results from
 281 benchtop testing of the MR ED system are shown in Table 2, and descriptions of the sam-
 282 ple and concentrate solutions that were used are in Table 1. Initial tests to verify MR
 283 ED’s functionality included circulating sample and concentrate without power applied
 284 to verify that the system had no leaks, as well as test of the diffusion of salts through
 285 the membranes without power applied. This latter test used a sample of 1 M NaCl and
 286 a lightly salty (2 mS/cm) NaCl solution as the concentrate; when left in the system, ions
 287 will naturally diffuse from the higher concentration sample channel to the lower concen-
 288 tration concentrate channels. Without circulation, the sample only was desalted to 58%
 289 its original salinity after 24 hours; circulation can increase the efficiency by replenish-
 290 ing the ions in the fluid nearest to the membranes. Applying a voltage potential across
 291 the membranes additionally increases the speed of the process and promotes further de-
 292 salting of the sample.

293 We conducted Tests 1 and 2 to initially assess the desalting capabilities and DOC
 294 recovery of the MR ED system. In Test 1, we prepared 300 mL of Sample 1, created from
 295 a 0.5 M NaCl solution spiked with 186 mg of glucose to achieve a concentration of 200
 296 ppm glucose. Both concentrate and sample solutions began at a temperature of 21 °C;
 297 however, the temperature rose quickly as the power was applied. The temperature of the
 298 sample after desalting the sample from 42.1 mS/cm to 0.978 mS/cm was 37.5 °C, and
 299 the temperature of the concentrate was 34.2 °C. The concentrate’s temperature did not
 300 increase as much as that of the sample, as we used a larger volume of concentrate than
 301 sample; 150.5 mL of the prepared sample solution was used in comparison to 1000 mL
 302 of concentrate. Throughout the test, the Raspberry Pi adjusted the conductivities for

Table 2. Results from benchtop system tests with initial and final conductivity, % DOC recovered, and calculated salinity (based on temperature and conductivity data) for NaCl-based solutions. Data available from Bryson et al. (2022)

Test	Sample	Initial conductivity [mS/cm] (Salinity [g NaCl/L])	Final conductivity [mS/cm] (Salinity [g NaCl/L])	DOC % recovery	Concentrate	Notes
Test 1	Sample 1	42.1 (27.1)	0.978 (0.487)	53%	Concentrate 1	No temperature control
Test 2	Sample 1	40.2 (25.8)	0.996 (0.493)	69%	Concentrate 1	
Test 3	Sample 2	21.4 (12.9)	1.04 (0.517)	71%	Concentrate 1	
Test 4	Sample 2	26.3 (16.1)	0.928 (0.458)	72%	Concentrate 1	
Test 5	Sample 2	29.3 (18.2)	0.945 (0.466)	-	Sample 2	Testing with a higher salinity concentrate, but carbon analysis results were inconsistent
Test 6	Sample 2	29.3 (18.1)	0.899 (0.443)	67%	Sample 2	Testing with a higher salinity concentrate
Test 7	Sample 3	85.1 (61.1)	0.913 (0.451)	77%	Concentrate 2	Higher salinity concentrate used to speed process
Test 8	Sample 3	90.1 (64.5)	49.1 (32.2)	-	Sample 3	Testing with higher salinity concentrate. Significant precipitation necessitated premature termination
Test 9	Sample 4	72.76 (TDS 400 g/L) ¹	27.0	-	Concentrate 1	Decrease in current indicated low ion flow across membranes
Test 10	Sample 4	71.48 (TDS 400 g/L) ¹	27.1	53%	Sample 4	Testing with higher salinity concentrate. Significant precipitation necessitated premature termination

¹Total dissolved solids measurement courtesy of (Klempay et al., 2021)

303 the temperature so that they could be read as conductivity at 25 °C, as temperature af-
304 fects the solution’s conductivity and would lead to inflated conductivity readings as the
305 temperature increased. In this test, the current was initially limited at 1.5 A, and de-
306 creased as the sample lost its ions and became less conductive. The total DOC recov-
307 ery for Test 1 was 53%, which is calculated from the final and initial DOC analyses, as
308 well as the final and initial volumes of sample.

309 In Test 2, we created the same sample and concentrate solutions as in the first test.
310 The initial conductivities of the sample and concentrate were 40.4 mS/cm and 10.4 mS/cm
311 respectively. However, both solutions were stored in beakers within an ice bath before
312 and during the test to address the temperature increase seen in Test 1. The initial tem-
313 perature of the sample was 10.1 °C, and the initial temperature of the concentrate was
314 12.7 °C. In this test, the sample heated 3.4 °C from the applied current, but the tem-
315 perature of the concentrate decreased to 12.1 °C – there was a large enough volume of
316 concentrate that the current did not increase the temperature faster than the cooling the
317 ice bath. In comparison to Test 1, the current never reached the imposed limit of 1.5 A
318 – the initial current was 1.35 A, and decreased throughout the test to 0.25 A. Because
319 both Tests 1 and 2 used the same sample solution, this difference in the current could
320 be a result of the difference in the temperature of the solutions; a colder solution is less
321 conductive, and thus supports less ion flow across it. The DOC recovery for Test 2 was
322 69% after the sample was desalted from 40.2 mS/cm to 0.996 mS/cm conductivity.

323 In addition to laboratory produced samples, we tested the system with natural sam-
324 ples that were sourced from the Skidaway River in Savannah, Georgia (USA). A total
325 of two gallons of sample were gathered in June 2021 during high tide at (31.997222, -
326 81.030500), an outlet of the Skidaway river across from Skidaway Institute of Oceanog-
327 raphy. The Skidaway River is an estuarine river that derives its higher salinity from the
328 ocean’s tides, thus its salinity reaches its maximum during high tide (Verity, 2002). Two
329 liters of sample were gathered and filtered to remove particles larger than 0.7 μm before
330 they were tested with MR ED to prevent clogging the tubing, interior channels, and mem-

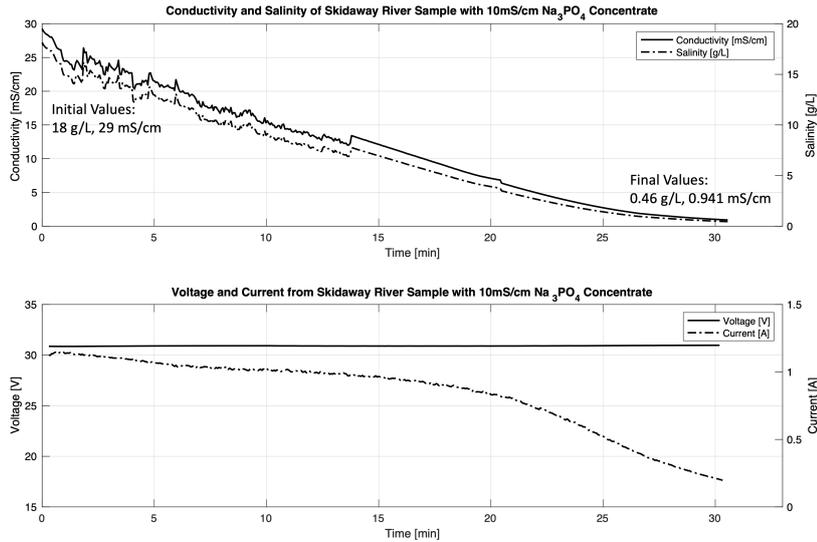


Figure 4. The processing of Sample 2 from the Skidaway River in Savannah, GA shows progressive desalting as power is applied across the membrane stack. In this experiment, the concentrate is a low salinity (initial conductivity of 10 mS/cm) Na_3PO_4 solution. The solution was desalted to 1 mS/cm conductivity within 33 minutes, removing 97.4% of the salts. Using the Na_3PO_4 solution provides a low salinity solution to transfer ions into (aided for a large portion of the processing by diffusion), and the current draw remains below the limit as the low salinity channels of the ED stack act as insulators.

branes. In planetary missions, pre-filtration would be a likely step prior to other processing (Lawrence et al., 2021), as is standard in ocean sampling. We processed these samples with Concentrate 1 and achieved an average of 72% DOC recovery. The conductivity, salinity – which was calculated using the Gibbs Seawater Toolbox from the measured conductivity and temperature (McDougall & Barker, 2011) – and power data from the process is displayed in Figure 4. The power data shows that the current began below the imposed limit at 1.5 A and decreased as the loss of ions made the sample less conductive, corresponding to the sample’s salinity decreasing. Current across the system can be related to the ion movement, so the higher current is an indicator of the faster desalting. As the sample’s conductivity approaches 0 mS/cm, the current decreases more quickly; the end of the desalting process takes much longer than its start as the lack of ions prevents current flow. From this we can conclude that although it is possible to desalt the sample below our chosen conductivity limit of 1 mS/cm, there is a trade space among final salinity, processing time, and DOC recovery.

On an in situ science package, a large quantity of concentrate would be needed for the duration of the mission; however, the need to carry this concentrate can be minimized by using the ambient water in its place. This is a difficult operation as the concentrate quickly becomes saltier than the sample that it is desalting, and it becomes more difficult for a power system to pull the ions into the saltier solution. In Tests 5 and 6, we used a 1 L volume of Sample 2 as the concentrate (see Table 1) to test this operation. We separated this volume from a 50 mL volume of Sample 2 that would be used as the sample. Both containers were placed in an ice bath, and the system was rinsed and circulated before power was applied. Immediately we noticed a difference in the power data from Tests 3 and 4, which can be seen in Figure 5 compared to Figure 4. When using

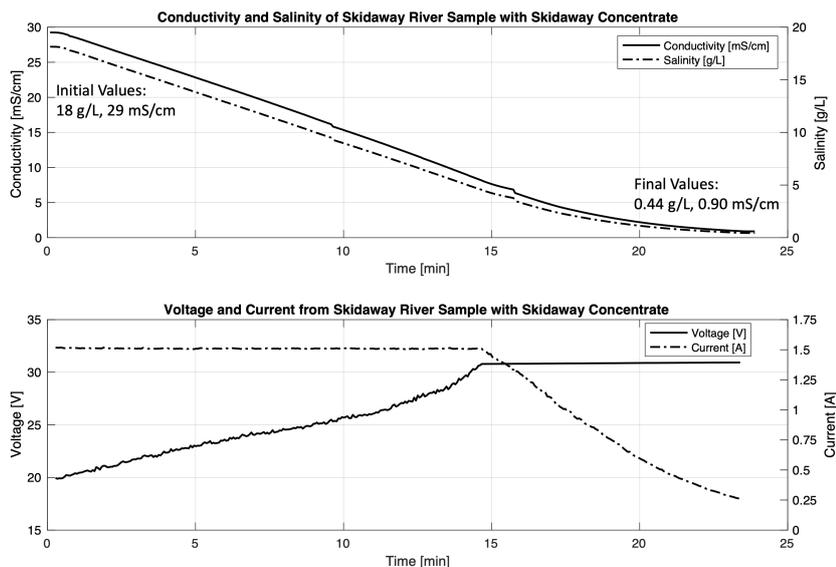


Figure 5. Processing of Sample 2 from the Skidaway River in Savannah, GA shows the desalting process in which 97.5% of the salts were removed. In this experiment, a separate container of this sample has been prepared to use as the concentrate to mimic the operation of using ambient water as the concentrate. The effect of using an ambient concentrate increases the initial current draw to its maximum. This accelerates the process, but this high current throughout the process may be responsible for the decreased DOC recovery from tests with the same sample and Concentrate 1.

355 a higher salinity solution as the concentrate, the electro dialysis stack is initially much
 356 more conductive than when using a lower salinity concentrate. Thus, the current across
 357 the system was higher than in previous tests, and immediately reached the 1.5 A limit,
 358 and, because of the current limit placed on the power supply, required less voltage po-
 359 tential across the system than the 30 V limit. This is not apparent in Figure 4, as the
 360 low salinity concentrate resists ion movement, so the current draw is below the imposed
 361 limit and the full 30 V is applied. This higher current overall also caused the temper-
 362 ature of both solutions to increase (sample + 0.93 °C, concentrate + 1.7 °C) despite the
 363 ice bath. However, Tests 5 and 6 were notably faster than the previous tests; the total
 364 processing time was 23 minutes, compared to Tests 3 and 4, which lasted 32 minutes us-
 365 ing the same sample and a lower salinity concentrate. Test 6 achieved 67% DOC recov-
 366 ery; the difference between it and the recoveries for Tests 3 and 4 may fall within the er-
 367 ror bounds of calculating the DOC recovery or be a slightly lower recovery compared to
 368 those using a lower salinity concentrate because the higher current draw in Test 6 com-
 369 pared to Tests 3 and 4.

370 Complexity increases in high salinity solutions, in comparison to seawater. The first
 371 brine we tested was a NaCl-dominated brine taken from South Bay Salt Works (SBSW).
 372 SBSW is a salt-harvesting facility in Chula Vista, CA, in which water from the San
 373 Diego Bay has evaporated to create shallow ponds replete with NaCl and MgCl₂ salts
 374 (Roseman & Watry, 2008). These briny salt ponds have been proposed as analogs for
 375 future life detection missions (Klempay et al., 2021). Samples from Site 5 (MgCl₂-saturated)
 376 and Site 11 (NaCl-saturated) of SBSW had been collected previously (Survey, 2011; Klem-
 377 pay et al., 2021), and were filtered to remove particles larger than 0.7 μm before test-

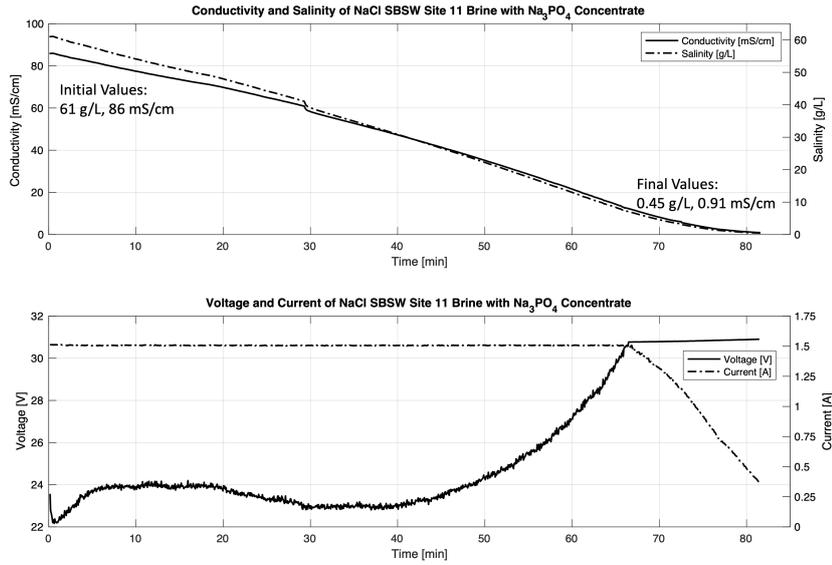


Figure 6. MR ED processing of 50 mL of a natural hypersaline solution, shows a less linear power increase throughout desalting as compared to the less saline solutions of ocean water from Skidaway River. Here, we show results for SBSW Site 11 sample. A low salinity (mixed to be 20 mS/cm conductivity) Na_3PO_4 solution is used as the concentrate. In this experiment, the brine was successfully desalted to 0.91 mS/cm conductivity without precipitation – which is 0.8% its original salinity.

378 ing with the MR ED system. The SBSW Site 11 brine selected for use as Sample 3 is
 379 1.86 times more concentrated in salt than compared to Earth’s seawater and was used
 380 in Tests 7 and 8. This brine was used to characterize the MR ED system’s ability to de-
 381 salt a high salinity sample to 1 mS/cm conductivity, to establish such baselines as the
 382 time to desalt and power required, as well as to investigate the DOC recovery with more
 383 complex samples. Test 7 successfully desalted Sample 3 to the 1 mS/cm conductivity limit
 384 with a 77% DOC recovery, and the sensing data from this process is shown in Figure 6.
 385 Both salinity and power data are similar to those in Figure 5; the current is initially lim-
 386 ited due to the high conductivity across the system, which is a result of using Concen-
 387 trate 2, and the voltage increases in response to conductivity across the sample cham-
 388 ber decreasing. Concentrate 2 was a Na_3PO_4 solution mixed to 20 mS/cm; the higher
 389 conductivity of the concentrate allowed for faster initial desalting. Test 7 was the longest
 390 test with a total processing time of 82 minutes, due to the high initial salinity of Sam-
 391 ple 3. Additionally, we observed a decrease of 12 mL from the initial 50 mL sample vol-
 392 ume, which stems from the osmosis of water molecules through the membranes (Jiang
 393 et al., 2015), and resulted in a concentration of the DOC in the sample and supported
 394 a higher DOC recovery.

395 In Test 8 we used Sample 3 as both the sample and concentrate in a manner simi-
 396 lar to Test 5. Because both sample and concentrate were highly conductive (initial con-
 397 ductivity of 89.7 mS/cm), the current limit caused a much lower initial voltage (10.5 V)
 398 than in previous tests. After 50 minutes of desalting the conductivity of the sample stopped
 399 decreasing, and the experiment was terminated. The lowest conductivity that the sam-
 400 ple reached was 48.2 mS/cm at which point 50% of the salts had been removed from the
 401 sample. When we disassembled the system for inspection, we observed precipitated salts

402 around the electrodes. Precipitate can cause clogging in fluidic channels and membranes,
403 which can decrease the desalting efficiency as well as instigate reactions at the electrodes.
404 Monitoring the sensing data during an experiment is important in order to observe po-
405 tential issues such as precipitation.

406 In Tests 9 and 10 we tested Sample 4 to evaluate the MR ED system's performance
407 with compositionally diverse brines. Mg brines in particular, such as Sample 4, may be
408 of relevance to Europa (Zolotov & Shock, 2004). Sample 4 was prepared as an 8x dilu-
409 tion of SBSW Site 5 brine sample, and in Test 9 we prepared a concentrate of 1 L of Con-
410 centrate 1 (Table 1). While the conductivity of the sample decreased linearly for the 100-
411 minute experiment, we noticed discrepancies in the current data. In Tests 2-4 that used
412 Concentrate 1, the current was initially below the limit and steadily increased to the 1.5
413 A limit as the concentrate became more conductive. However, in the data from Test 9
414 the current started to decrease at 0.8 A, well below the limit. The low and decreasing
415 current indicated low ion flow across the membranes, and the experiment was terminated.
416 Although we did not observe precipitate around the electrodes that would explain the
417 decreased ion flow due to clogging the membranes, the decreasing ion flow indicated that
418 the process was unsuccessful. In Test 10 we used 500 mL of Sample 4 as the concentrate
419 to improve the ion movement across membranes with a high salinity concentrate. This
420 improved the ion flow, as the current draw was 1.5 A across the entirety of the test; how-
421 ever, we quickly noticed that the concentrate solution turned a pale-yellow color, and
422 there was precipitation in the sample reservoir. Additionally, although the current data
423 appeared as we would expect, the voltage initially decreased as the conductivity of the
424 sample decreased, rather than increasing as in previous tests. The experiment ultimately
425 stopped as a pump began to leak, and when the system was opened for inspection, salts
426 had precipitated out of solution in the concentrate channels, particularly around the elec-
427 trodes. The increased acidity of the sample and concentrate had corroded holes in the
428 membranes, which were disposed of after the experiment. In Test 10 we recovered the
429 sample solution to analyze the DOC recovery; the DOC recovery was 53% after the con-
430 ductivity had decreased to 38% its initial value. These results indicate that a different
431 procedure is required for highly saturated or acidic brines, such as lowering the current
432 limit or applying a pulsed current to reduce the rate of ion transport and has been used
433 to increase DOC recovery during late-stage ED when the desalting rate decreases (Gurtler
434 et al., 2008).

435 4 Conclusions

436 The Miniature Robotic Electrodialysis system was designed for desalting small vol-
437 umes of sample (less than 100 mL) as technology development for a liquid sample han-
438 dling system for future ocean world missions for which proposed instruments would re-
439 quire desalting prior to analysis (Lawrence et al., 2021). However, such a small system
440 has applications both on Mars and for field use Earth, allowing desalting of samples for
441 many uses. We designed the MR ED system in order to increase the technology readi-
442 ness level (TRL) of small-scale electrodialysis systems for desalting samples for the in-
443 vestigation of organics. Technology readiness levels provide an assessment of a particu-
444 lar technology's maturity with respect to spaceflight, from the observation of the basic
445 principle (TRL1) to having been used successfully on a flown mission (TRL9) (Mai,
446 2015). After miniaturizing an ED system and successfully desalting natural samples con-
447 taining primarily NaCl salts in a laboratory setting, we expect the technology readiness
448 level of a milliliter-scale electrodialysis system to be elevated to TRL4, in which the tech-
449 nology has been validated in a laboratory environment. The extent of this work has val-
450 idated the miniaturized, single cell-pair system for use on small volumes of Earth sea-
451 water and brines with NaCl salts. Further TRL elevation requires the system to be tested
452 in a relevant environment to its planned use, as well as further developing the system
453 to be used in-line with the organic-detecting instruments. For instance, to increase TRL

454 for use on Martian environments, the system should be tested with solutions contain-
455 ing perchlorates at the expected concentration.

456 The design of MR ED has a low complexity and ease of configuration. The system
457 can be reliably opened for inspection and reassembled, and the separate chambers that
458 comprise the assembly can be either machined or 3D printed from resin, making it eas-
459 ily replicable. These channels of fluid, which are thicker than cells in commercial ED stacks,
460 allow greater mixing of the fluid that enables a higher DOC recovery as well as enhances
461 the efficiency. Experimentation and sensing illuminated us to certain components of the
462 design, such as temperature management. The inclusion of the ice bath for sample and
463 concentrate containers was introduced after tests showed large temperature increases.
464 As the temperature of both the sample and concentrate affect their conductivity, a colder
465 sample and concentrate will be slightly more insulative than a warmer sample and con-
466 centrate. This prevents excess current from conducting through the system, and could
467 lead to a larger recovery of DOC. Thus, temperature management for this system will
468 be important in the design in an instrument package; this would additionally benefit down-
469 stream instruments by keeping the sample at close to its natural state. A similar aspect
470 of the design was the in-line power sensing, which allowed us a greater understanding
471 of the process that was sealed inside the system. Monitoring the power, and understand-
472 ing the nominal operation of the power system, allowed us to see that the desalting ef-
473 ficiency was reduced due to precipitated salts in several tests. Future development plans
474 include developing the autonomy by using the Raspberry Pi to control the pumps and
475 the voltage and current limits of a power system according to conductivity and temper-
476 ature readings of the sample. This all should be contained to a waterproof housing to
477 allow in situ testing and field deployments.

478 Processing with electrodialysis is a destructive technique; however, we have shown
479 that a significant amount of dissolved organic carbon can be recovered. The issues salts
480 pose to instruments that detect organic signatures are great, thus we conclude that a de-
481 structive technique is useful to remove destructive elements of the sample while retain-
482 ing measurable components. In our tests, the MR ED system, processing less than 10%
483 of the volume of sample processed by established instruments, achieved successful de-
484 salting of natural samples and NaCl brines to a conductivity of 1 mS/cm with DOC re-
485 covery spanning a range from 53% to 77%, which is comparable to desalting with estab-
486 lished laboratory electrodialysis systems that recover on average 70% DOC (Chambers
487 et al., 2016; Gurtler et al., 2008; Koprivnjak et al., 2006; Vetter et al., 2007). Best re-
488 covery was achieved after optimizing the experimental set up, where MR ED success-
489 fully desalted NaCl brines using a low salinity Na₃PO₄ concentrate and retained 77%
490 DOC after removing 99.2% the salts. However, tests with brines containing MgCl₂ salts
491 resulted in salt precipitated out of solution and further investigation is needed to opti-
492 mize desalting brines. Potential measures that could be used to improve operation with
493 brines include processing with a different composition concentrate, processing at a lower
494 power draw or with a pulsed current, or circulation at a different flow rate. Finally, we
495 showed that MR ED achieved 67% DOC recovery when using a separate stock of the ini-
496 tial natural sample as the concentrate, thus operating as an in situ system would by us-
497 ing surrounding water as the concentrate. This is a unique operation that has not yet
498 been tested on established systems. These experiments establish the utility and base-
499 line capabilities of an autonomous miniature electrodialysis system to be used with an
500 instrument package in high salinity environments.

501 5 Open Research

502 The sensing data, measurement data, and observations used for analysis of the elec-
503 trodialysis efficiency and effectiveness in this study are available at Zenodo, Github via
504 <https://zenodo.org/record/7076436> under the Creative Commons Attribution 4.0 Inter-
505 national license (Bryson et al., 2022). A persistent link to the Jupyter notebook used

506 for data collection and observations is available at [https://mybinder.org/v2/gh/fbryson820/](https://mybinder.org/v2/gh/fbryson820/Development-of-MRED-Data.git/main?labpath=Development%20of%20the%20Miniature%20Robotic%20Electrodialysis%20(MR%20ED)%20System%20for%20Small-Scale%20Desalting%20of%20Liquid%20Samples%20with%20Recovery%20of%20Organics.ipynb)
 507 [Development-of-MRED-Data.git/main?labpath=Development%20of%20the%20Miniature%](https://mybinder.org/v2/gh/fbryson820/Development-of-MRED-Data.git/main?labpath=Development%20of%20the%20Miniature%20Robotic%20Electrodialysis%20(MR%20ED)%20System%20for%20Small-Scale%20Desalting%20of%20Liquid%20Samples%20with%20Recovery%20of%20Organics.ipynb)
 508 [20Robotic%20Electrodialysis%20\(MR%20ED\)%20System%20for%20Small-Scale%20Desalting%](https://mybinder.org/v2/gh/fbryson820/Development-of-MRED-Data.git/main?labpath=Development%20of%20the%20Miniature%20Robotic%20Electrodialysis%20(MR%20ED)%20System%20for%20Small-Scale%20Desalting%20of%20Liquid%20Samples%20with%20Recovery%20of%20Organics.ipynb)
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