Improved consistency between the modelling of ocean optics, biogeochemistry and physics, and its impact on the North-West European Shelf seas

Jozef Skakala^{1,1}, Jorn Bruggeman^{2,2}, David Andrew Ford^{3,3}, Sarah L Wakelin^{4,4}, Anil Akpınar^{4,4}, Tom Hull^{5,5}, Jan Kaiser^{6,6}, Benjamin Roger Loveday^{1,1}, Charlotte Anne June Williams^{4,4}, and Stefano Ciavatta^{7,7}

¹Plymouth Marine Laboratory
²PML
³Met Office
⁴National Oceanography Centre
⁵Centre for Environment, Fisheries and Aquaculture Science
⁶University of East Anglia
⁷Plymouth Marine Laboratory/National Centre for Earth Observation

November 30, 2022

Abstract

We use a recently developed spectrally resolved bio-optical module to better represent the interaction between the incoming irradiance and the heat fluxes in the upper ocean within the (pre-)operational physical-biogeochemical model on the North-West European (NWE) Shelf. The module attenuates light based on the simulated biogeochemical tracer concentrations, and thus introduces a two-way coupling between the biogeochemistry and physics. We demonstrate that in the late spring-summer the two-way coupled model heats up the upper oceanic layer, shallows the mixed layer depth and influences the mixing in the upper ocean. The increased heating in the upper oceanic layer reduces the convective mixing and improves by $\tilde{}5$ days the timing of the late phytoplankton bloom of the ecosystem model. This improvement is relatively small compared with the existing model bias in bloom timing, but sufficient to have a visible impact on model skill. We show that the changes to the model temperature and salinity introduced by the module have mixed impact on the physical model skill, but the skill can be improved by assimilating the observations of temperature, salinity and chlorophyll into the model. However, in the situations where we improved the simulated oxygen concentration as a result of the changes in the simulated air-sea gas flux. Overall, comparing different 1-year experiments showed that the best model skill is achieved with joint physical-biogeochemical assimilation into the two-way coupled model.

¹ Highlights

² Improved consistency between the modelling of ocean optics, bio-

³ geochemistry and physics, and its impact on the North-West Eu⁴ ropean Shelf seas

⁵ Jozef Skákala, Jorn Bruggeman, David Ford, Sarah Wakelin, Anıl Akpınar,

Tom Hull, Jan Kaiser, Benjamin R. Loveday, Charlotte A.J. Williams, Ste fano Ciavatta

- We established two-way coupling between biogeochemistry and physics.
- We assessed the impact of the coupling in free and assimilative exper iments.
- The two-way coupling improves the simulated biogeochemistry on the NWE Shelf.
- We recommend to implement this development into the operational system.

Improved consistency between the modelling of ocean optics, biogeochemistry and physics, and its impact on the North-West European Shelf seas

Jozef Skákala^{a,b}, Jorn Bruggeman^a, David Ford^c, Sarah Wakelin^d, Anıl

Akpınar^d, Tom Hull^{e,f}, Jan Kaiser^f, Benjamin R. Loveday^g, Charlotte A.J.
 Williams^d, Stefano Ciavatta^{a,b}

 ^aPlymouth Marine Laboratory, Prospect Place, The Hoe, Plymouth, PL1 3DH, United Kingdom
 ^bNational Centre for Earth Observation, Prospect Place, The Hoe, Plymouth, PL1 3DH, United Kingdom
 ^cMet Office, FitzRoy Road, Exeter, EX1 3PB, United Kingdom
 ^dNational Oceanography Centre, Joseph Proudman Building, 6 Brownlow Street, Liverpool, L3 5DA, United Kingdom
 ^eCentre for Environment, Fisheries and Aquaculture Science, Lowesoft, NR33 0HT, United Kingdom
 ^fCentre for Ocean and Atmospheric Science, University of East Anglia, Norwich, NR4 7TJ, United Kingdom
 ^gInnoflair UG, Richard-Wagner-Weg 35, Darmstadt, 64287, Germany

21 Abstract

We use a recently developed spectrally resolved bio-optical module to better represent the interaction between the incoming irradiance and the heat fluxes in the upper ocean within the (pre-)operational physical-biogeochemical model on the North-West European (NWE) Shelf. The module attenuates light based on the simulated biogeochemical tracer concentrations, and thus introduces a two-way coupling between the biogeochemistry and physics. We demonstrate that in the late spring-summer the two-way coupled model heats up the upper oceanic layer, shallows the mixed layer depth and influences the mixing in the upper ocean. The increased heating in the upper oceanic layer reduces the convective mixing and improves by ~5 days the timing of the late phytoplankton bloom of the ecosystem model. This improvement is relatively small compared with the existing model bias in bloom timing, but sufficient to have a visible impact on model skill. We show that the changes to the model temperature and salinity introduced by the module have mixed

Preprint submitted to Ocean Modelling

April 14, 2021

impact on the physical model skill, but the skill can be improved by assimilating the observations of temperature, salinity and chlorophyll concentrations into the model. However, in the situations where we improved the simulation of temperature, either via the bio-optical module, or via assimilation of temperature and salinity, we have shown that we also improved the simulated oxygen concentration as a result of the changes in the simulated air-sea gas flux. Overall, comparing different 1-year experiments showed that the best model skill is achieved with joint physical-biogeochemical assimilation into the two-way coupled model.

- 22 Keywords: two-way coupled physical-biogeochemical model, ocean
- ²³ chlorophyll concentration, sea surface temperature, phytoplankton spring
- ²⁴ bloom, North-West European Shelf (10E-10W, 40N-68N), data assimilation

25 1. Introduction

Physical-biogeochemical ocean models are an essential element in mon-26 itoring and forecasting of global and shelf-sea ecosystem indicators ([1, 2]). 27 However, coupled physical-biogeochemical marine modelling is a complex un-28 dertaking and a common way to simplify coupled models is to neglect the 20 impact of the biogeochemical model state on physics ([3, 2]). Although ma-30 rine ecosystem models often neglect the coupling from the biogeochemical 31 model state to physics, there are number of established mechanisms through 32 which biogeochemistry influences physics and climate ([4, 1, 2]): (i) marine 33 ecosystems play an essential part in the carbon cycle through biological and 34 microbial carbon pump, influencing atmospheric carbon concentrations and 35 the Earth surface temperature, (ii) phytoplankton influences oceanic albedo 36 (e.g. [5]) having an overall impact on the radiative terms and Earth energy 37 budget, (iii) some biogeochemical tracers influence light attenuation, mod-38 ifying the short-wave heat fluxes in the water column and therefore ocean 39 stratification ([6, 7, 8, 9, 10, 11, 12, 13, 14]), and (iv) marine ecosystems have 40 an impact on cloud condensation nuclei through the production of dimethyl 41 sulfide (DMS, [15, 16, 17, 18]), or more directly via bubble formation ([19]). 42 The size of life's impact on Earth's physics has been subject to much de-43 bate ([2]), often in connection with "the Gaia hypothesis" ([20, 21]), which 44 proposes that life plays a central role in regulating climate. 45

For coupled physical-biogeochemical marine models the main source of impact of ecosystems on physics is through the absorption and backscatter-

ing of short-wave radiation by some biogeochemical substances in the sea 48 water (e.g. [10]). The presence of optically active tracers, such as chloro-49 phyll, suspended particulate matter (SPM), or colored dissolved organic mat-50 ter (CDOM), in the oceanic upper layer increases light attenuation near the 51 oceanic surface, warms the sea temperature in the upper ocean, which typi-52 cally influences the mixing in the upper oceanic layer (e.g. [6]), e.g. shallow-53 ing the thermocline and the mixed layer depth (MLD). The changes to the 54 vertical mixing can in turn impact the biogeochemical model, by influencing 55 the nutrient concentrations and growth conditions in the upper ocean. 56

In this work we focus on the Copernicus Marine Environmental Moni-57 toring Service (CMEMS) operational system for the North-West European 58 (NWE) Shelf biogeochemistry, which is of a substantial societal benefit, as the 59 NWE Shelf is a key region for fisheries, and an important contributor to the 60 global carbon cycle ([22, 23, 24]). The presently used physical-biogeochemical 61 operational model for the NWE Shelf is the marine physical model Nucleus 62 for European Modelling of the Ocean (NEMO, [25]) coupled through the 63 Framework for Aquatic Biogeochemical Models (FABM, [26, 27]) to the Euro-64 pean Regional Seas Ecosystem Model (ERSEM, [28, 29, 30]). NEMO-FABM-65 ERSEM drives its physics and biogeochemistry by two separate irradiance 66 modules: (i) the physical model calculates heat fluxes from the incoming 67 net short-wave radiation (SWR) split into two wavebands, the 400-700nm 68 visible band reduced through attenuation obtained from a monthly climatol-69 ogy of a satellite surface K_d product at 490nm wavelength (European Space 70 Agency product version 2.0, https://www.esa-oceancolour- cci.org/), and the 71 UV/infrared band reduced with a preset attenuation with an e-folding scale 72 of 0.35m, (ii) the biogeochemical model reduces incoming photosynthetic ac-73 tive radiation (PAR) by taking into account both absorption and backscat-74 tering by the sea water and the simulated Phytoplankton Functional Types 75 (PFTs), and also by including absorption by Particulate Organic Matter 76 (POM), CDOM and sediment represented by an external satellite product 77 (for details see [29, 31]). The presently used scheme means that, although 78 some impact of biogeochemical tracers on the physical model is implicitly 79 included in the 490nm K_d satellite climatology, there is no feedback from the 80 biogeochemical model state to the simulated physics. 81

In [31] we implemented into ERSEM a stand-alone bio-optical module (based on OASIM, [32, 33, 34]), that resolves irradiance spectrally and splits the irradiance into diffuse and direct streams ([35]). The module then propagates irradiance through the water column, based on attenuation by sea water

and the biogeochemical substances in the water. The new module drove only 86 the biogeochemical part of the coupled NEMO-FABM-ERSEM model, sub-87 stantially improving the underwater irradiance, but without a major impact 88 on the ERSEM model skill on the NWE Shelf ([31]). This version of NEMO-89 FABM-ERSEM model will be used in the present study as a reference run 90 and will be called a "one-way coupled model". In this work we expand the 91 development implemented in [31] by using the bio-optical module to drive 92 both the biogeochemistry and the physics (i.e. heating by light absorption). 93 Since the physical heat fluxes will be driven by the underwater irradiance 94 that is attenuated by biogeochemical substances, the module establishes an 95 important feedback from the biogeochemical model to physics. We will fur-96 ther call this new implementation a "two-way coupled model", to distinguish 97 it from the "one-way coupled" reference run. 98

This work aims at answering two main questions: (i) What is the size gc of the biogeochemical impact on the marine physics within the NWE Shelf? 100 (ii) Does the impact of the spectrally resolved bio-optical module on physics 101 lead to more internally consistent ecosystem dynamics on the NWE Shelf. 102 and hence, an improvement in the operational biogeochemical model skill? 103 Those two questions are answered both in the context of free simulations 104 and also in the context of (physical, biogeochemical, coupled) assimilative 105 runs. The second question is particularly relevant: It has been established 106 that NEMO-FABM-ERSEM displays on the NWE Shelf late and intense 107 spring blooms ([31, 36]). Since a spring bloom is a major ecosystem driver 108 ([37, 38]), the simulated late blooms severely limit the ecosystem model skill 109 ([31, 36]). Although many factors can influence the bloom timing (including 110 biological drivers, such as zooplankton grazing, e.g. [39]), one of the leading 111 hypotheses for how phytoplankton blooms are triggered in the North Atlantic 112 is based on the interplay between PAR and an effective mixing depth (the 113 critical turbulence hypothesis, [40, 41]), i.e. the bloom sets in when the 114 effective mixing depth becomes fully contained within the euphotic layer 115 ([42]). Within the scope of the critical turbulence hypothesis, the delay in 116 bloom timing could then be explained by multiple components within the 117 physical-biogeochemical coupled model: (a) atmospheric wind stress forcing, 118 (b) model upper-ocean mixing scheme, (c) vertical stratification (thermocline 119 and pycnocline), (d) incoming surface PAR, (e) underwater light attenuation, 120 (f) the phytoplankton growth response to light (e.g. model parameters, such 121 as P-I curves, maximum PFT chlorophyll-to-carbon ratios). In [31] we have 122 addressed to a varying degree the points (d) and (e) without a significant 123



Figure 1: A schematic representation of the hypothesis about the impact of the two-way coupled model on the timing of the simulated bloom.

impact on the bloom timing. However, [31] observed that attenuation of 124 light based on the satellite K_d product for the 490 nm wavelength is most 125 likely an underestimate of the total PAR absorbed in the upper oceanic layer. 126 Calculating heat fluxes using the bio-optical module is therefore expected to 127 produce extra heat in the upper oceanic layer (Fig.5 of [31]), which is thought 128 to shallow the MLD, but it can also reduce turbulent convective mixing near 129 the oceanic surface ([43, 44]). The hypothesis tested in this work (see Fig.1) is 130 that the reduced convective mixing can lead to a shallower turbulent mixing 131 depth and help trigger an earlier phytoplankton bloom, as suggested by the 132 critical turbulence hypothesis ([40, 44]). The biogeochemical feedback to the 133 simulated physics could therefore improve the ERSEM skill on the NWE 134 Shelf. 135

¹³⁶ 2. Methods

137 2.1. The physical model: NEMO

The NEMO ocean physics component (OPA) is a finite difference, hydro-138 static, primitive equation ocean general circulation model ([25]). The NEMO 139 configuration used in this study is similar to the one used by [45, 46, 31], and 140 identical to the configuration used in [36]: we use the CO6 NEMO version. 141 based on NEMOv3.6, a development of the CO5 configuration explained in 142 detail by [47]. The model has 7 km spatial resolution on the Atlantic Margin 143 Model (AMM7) domain using a terrain-following $z^* - \sigma$ coordinate system 144 with 51 vertical levels ([48]). The lateral boundary conditions for physical 145 variables at the Atlantic boundary were taken from the outputs of the Met 146 Office operational $1/12^{\circ}$ North Atlantic model (NATL12, [49]); the Baltic 147 boundary values were derived from a reanalysis produced by the Danish Me-148 teorological Institute for CMEMS. We use annually varying river discharge 149 based on data from [50]. 150

The model was forced at the surface by atmospheric fluxes provided by 151 an hourly and 31km resolution realisation (HRES) of the ERA5 data-set 152 (https://www.ecmwf.int/). In case of the one-way coupled model the ERA5 153 fluxes provide also the total incoming net shortwave radiation whose visible 154 fraction is attenuated inside the water column based on the Kd for 490nm 155 wavelength supplied by a monthly climatology from an Ocean Color - Climate 156 Change Initiative (OC-CCI) product of European Space Agency (ESA), ver-157 sion 4.1 (https://www.esa-oceancolour- cci.org/). For the two-way coupled 158 model the incoming net short-wave radiation is decomposed into direct and 159 diffuse streams and spectrally resolved, and is provided by the bio-optical 160 module ([31]) that will be described later in the ecosystem model section. 161 The direct and diffuse streams are attenuated throughout the water column 162 by the bio-optical module, and subsequently integrated by NEMO to calcu-163 late the heating within each vertical layer. 164

165 2.2. The ecosystem model: ERSEM

ERSEM ([28, 29]) is a lower trophic level ecosystem model for marine biogeochemistry, pelagic plankton, and benthic fauna ([51]). The model splits phytoplankton into four functional types largely based on their size ([28]): picophytoplankton, nanophytoplankton, diatoms and dinoflagellates. ERSEM uses variable stoichiometry for the simulated plankton groups ([52, 53]) and each Phytoplankton Functional Type (PFT) biomass is represented in terms of chlorophyll, carbon, nitrogen and phosphorus, with diatoms also represented by silicon. ERSEM predators are composed of three zooplankton types (mesozooplankton, microzooplankton and heterotrophic nanoflagellates), with organic material being decomposed by one functional type of heterotrophic bacteria ([29]). The ERSEM inorganic component consists of nutrients (nitrate, phosphate, silicate, ammonium and carbon) and dissolved oxygen. The carbonate system is also included in the model ([54]).

We applied in this study the ERSEM configuration from [36], based on a new ERSEM version 20.10, which has an updated benthic component with respect to [29]. The ERSEM parametrization is identical to the one described in [29]. The Atlantic boundary values for nitrate, phosphate, silicate and oxygen were taken from World Ocean Atlas ([55]) and dissolved inorganic carbon from the GLODAP gridded dataset ([56, 57]), while plankton and detritus variables were set to have zero fluxes at the Atlantic boundary.

The irradiance at the ocean surface was calculated using the bio-optical 186 module implemented into the NEMO-FABM-ERSEM AMM7 configuration 187 by [31]. The bio-optical module resolves irradiance spectrally and distin-188 guishes between downwelling direct and diffuse streams. The module is 189 forced by the ERA5 atmospheric inputs (https://www.ecmwf.int/) for to-190 tal vertically integrated ozone, water vapour, cloud cover, cloud liquid water 191 and sea-level air pressure, as well as by a satellite product for aerosol op-192 tical thickness (MODerate resolution Imaging Spectroradiometer, MODIS, 193 https://modis.gsfc.nasa.gov/data/dataprod), and also by data for surface 194 wind speed, air humidity, and air temperature, all provided by the NEMO at-195 mospheric (ERA5) forcing. The attenuation of the irradiance was described 196 in detail by [31], here it is briefly summarized: The module distinguishes 197 between the absorption and backscattering by the sea water and the 4 PFTs 198 based on the model of [58]. The scheme for the underwater irradiance was 199 based on [33], i.e. the irradiance was resolved at 33 wavelengths in the 200 250 - 3700nm range, and so were the wavelength-dependent absorption and 201 backscattering coefficients for clear water and PFTs. Although we included 202 the impact of backscattering on the light attenuation, similarly to [31], we 203 did not explicitly track the upwelling stream. Besides the clear sea water 204 and PFTs, we included into the light attenuation also the absorption by 205 POM, CDOM and sediment, which was (the same as in [31]) forced by an 206 external product extrapolated from the 443nm data of [59]. The bio-optical 207 module was extensively validated in [31], and was shown to be skilled in its 208 representation of SWR, PAR and the underwater irradiances. 209

210 2.3. Observations: assimilated and validation data

211 2.3.1. Assimilated data

In the physical data assimilation component we have included: a) sea 212 surface temperature data from the GCOM-W1/AMSR-2, NOAA/AVHRR, 213 MetOp/AVHRR, MSG/SEVIRI, Sentinel-3/SLSTR, Suomi-NPP/VIIRS satel-214 lite products and in situ SST observations from ships, surface drifters and 215 moorings, distributed over the Global Telecommunication System (GTS) in 216 near-real time, b) temperature and salinity from the EN4 dataset ([60]), 217 which includes in situ profiles from Argo floats, fixed moored arrays, XBTs, 218 CTDs, gliders, marine mammals, and c) temperature and salinity data from 219 a specific Slocum glider Cabot (Unit 345, see [36]) that has been deployed 220 in the central North Sea during 08/05/2018 - 15/08/2018 as a part of the 221 Alternative Framework to Assess Marine Ecosystem Functioning in Shelf 222 Seas (AlterECO) programme (https://altereco.ac.uk/). The satellite SST 223 was bias-corrected following the scheme from [61], using the VIIRS and in 224 situ SST data as the reference. 225

In the biogeochemical data assimilation we have included total log-chlorophyll derived from the ocean color based satellite product of ESA (version 2.0, [62]) and also log-chlorophyll derived from the fluorescence measurements by the same AlterEco glider Cabot, that was used in the physical data assimilation. The assimilation is performed for log-chlorophyll, rather than chlorophyll, as chlorophyll is widely known to be log-normally distributed ([63]).

The assimilated in situ (EN4, glider) observations were thinned to a resolution of 0.08° (EN4), or up-scaled to the AMM7 grid (glider), with additional temporal averaging applied to the same-day glider observations. The thinning/up-scaling is performed to avoid assimilating many observations at higher resolution than the model can represent. After the thinning/upscaling there were O(10⁵) EN4 and O(10⁴) Cabot glider data-points to assimilate throughout the year 2018.

239 2.3.2. Validation data

The assimilated observations were used for the validation of those experiments in which they were excluded from the assimilation (e.g. chlorophyll data for the physical data assimilative run). However, we excluded the biascorrected satellite SST from the temperature validation, so that the only assimilated SST data used for validation were a) the high quality SST data from the VIIRS satellite product and from ships, drifters and moorings (we will call this "VIIRS/in situ SST data"), and the SST that was part of b) EN4

and c) Cabot glider data. Besides the assimilated observations, all the exper-247 iments were validated with other (non-assimilated) AlterEco glider data for 248 temperature, salinity, chlorophyll, oxygen and the sum of nitrate and nitrite 249 (all the gliders included in the validation are listed in Tab.1). The processing 250 of the physical, chlorophyll and oxygen data was described in [36]. The sum 251 of nitrate and nitrite concentrations (abbreviated as $NO_{x^-} = NO_{3^-} + NO_{2^-}$) 252 were determined using a Lab-on-Chip (LoC) analyser designed and fabricated 253 at the National Oceanography Centre ([64]), which were implemented by the 254 AlterEco team into Seagliders following a similar protocol as used by [65]. 255 The combined uncertainty (random and systematic errors) of measurements 256 made using these LoC analysers has been calculated as 5% (coverage interval 257 k = 1 ([66]). The nitrite concentrations were relatively negligible compared 258 to the nitrate concentrations, so the NO_x - data were used to validate model 259 nitrate outputs. All of the used AlterEco gliders operated during 2018 in 260 the central North Sea (for both the glider and the EN4 data locations see 261 Fig.S1 of the Supporting Information (SI), moving throughout the whole 262 water column. Similar to the assimilated Cabot glider, the remaining glider 263 data were up-scaled onto the model grid (on a daily basis) and after the up-264 scaling there remained $O(10^4)$ AlterEco glider observations for each variable 265 in 2018. 266

The EN4 data-set contained subsurface observations that were approxi-267 mately homogeneously distributed both with depth and in time, with slightly 268 lower number of observations towards the end of the year (November-December 269 2018). Beyond the assimilated data and the AlterEco data, we used for vali-270 dation a 1960-2014 monthly climatological dataset for total chlorophyll, oxy-271 gen, nitrate, phosphate and silicate concentrations, compiled in the North 272 Sea Biogeochemical Climatology (NSBC) project ([67]). The NSBC dataset 273 covers most of the NWE Shelf and the full range of depths. Finally, we also 274 included validation of surface CO_2 fugacity using 2018 SOCAT (v2019) data 275 (https://www.socat.info/index.php/about/). 276

277 2.4. The assimilative system: NEMOVAR

NEMOVAR is a variational (in this study a 3DVar) DA system ([68, 69, 70]) used at the Met Office for operational reanalyses and forecasting on
the NWE Shelf. The assimilation of ocean color-derived chlorophyll using
NEMOVAR is highly successful in improving the NWE Shelf phytoplankton
phenology, PFT community structure (using PFT chlorophyll assimilation),
underwater irradiance and to a more limited degree also carbon cycle ([46, 31,

Table 1: The AlterEco gliders and the variables measured by the gliders used for assimilation (6-th column), or validation (7-th column). The table uses the following abbreviations: deployment: "dpl", data assimilation: "DA", temperature: "T", salinity: "S", oxygen concentrations: "O₂", chlorophyll *a* concentrations: "Chl *a*" and sum of nitrate and nitrite concentrations: "NO_x-".

Campaign	platform	dpl	serial	mission period	DA	validation	
AlterEco 1	Stella	440	$unit_436$	02/02/2018 - 08/05/2018	none	$T,S,O_2,Chl a$	
AlterEco 1	Cook	441	$unit_194$	15/11/2017 - 07/02/2018	none	$T,S,O_2,Chl a,NO_x$ -	
AlterEco 2	Orca	493	SG510	07/03/2018 - 27/03/2018	none	Chl a ,NO _x -	
AlterEco 2	Melonhead	496	SG620	07/02/2018 - 02/04/2018	none	Chl a	
AlterEco 3	Cabot	454	$unit_{345}$	08/05/2018 - 15/08/2018	T,S,Chl a	$T,S,O_2,Chl a$	
AlterEco 3	Orca	455	SG510	16/03/2018 - 24/07/2018	none	Chl a ,NO _x -	
AlterEco 3	Humpback	497	SG579	09/05/2018 - 25/06/2018	none	Chl a	
AlterEco 4	Dolomite	477	$unit_{-}305$	13/08/2018 - 10/10/2018	none	T,S,Chl a ,NO _x -	
AlterEco 4	Eltanin	478	SG550	15/08/2018 - 28/09/2018	none	Chl a	
Altereco 5	Kelvin	481	$unit_444$	26/09/2018 - 02/12/2018	none	T,S,Chl a	
AlterEco 6	Dolomite	499	$unit_{-}305$	02/12/2018 - 12/03/2018	none	$T,S,O_2,Chl a$	
AlterEco 6	Coprolite	500	$unit_331$	02/12/2018 - 12/03/2018	none	$T,S,O_2,Chl a$	

²⁸⁴ 71]). NEMOVAR includes capability to assimilate multi-platform (satellite,
²⁸⁵ in situ) data, which has been established first for physics (e.g. [70, 72])
²⁸⁶ and subsequently for biogeochemistry ([73]), including validating the multi²⁸⁷ platform DA system for the NWE Shelf ([36]).

The NEMOVAR set-up used in this study for the multi-platform physical-288 biogeochemical assimilation is the same as the one described in detail by [36]. 289 Here we offer only a short summary: The 3DVar version of NEMOVAR uses 290 a First Guess at Appropriate Time (FGAT) to calculate a daily set of in-291 crements for the directly updated variables ([70, 72]). In the physical DA 292 application NEMOVAR applies balancing relationships within the assimi-293 lation step and delivers a set of increments for temperature, salinity, sea 294 surface height (SSH) and the horizontal velocity components. For the to-295 tal chlorophyll assimilation NEMOVAR calculates a set of log-chlorophyll 296 increments and then a balancing scheme is used to distribute those incre-297 ments into the PFT components (chlorophyll, carbon, nitrogen, phosphorus 298 and for diatoms also silicon), all of which are being updated based on the 290 background community structure and stoichiometric ratios (e.g. [46, 31, 36]). 300 After the assimilation step, the model is re-run with the increments applied 301 to the model variables gradually at each model time-step using incremental 302 analysis updates (IAU, [74]). 303

NEMOVAR uses externally supplied spatio-temporally varying observa-304 tion and background error variances, with the background error variances 305 typically 1-3 times larger than the observational error variances ([36]). The 306 system combines two horizontal correlation length-scales, one fixed 100km 307 length-scale with another length-scale based on the baroclinic Rossby radius 308 of deformation (72). The vertical length-scales follow the scheme from 72, 309 where NEMOVAR calculates directly the set of 3D increments using flow-310 dependent vertical length-scales (ℓ) , which are at the surface equal to half 311 of the MLD, decreasing in the mixed layer to become two-times the vertical 312 model grid spacing at, and beneath the MLD. 313

314 2.5. The experiments

In this study we compared the performance of both one-way and two-way 315 coupled versions of the NEMO-FABM-ERSEM model. We also tested the 316 impact of assimilating different types of data (physical-only, biogeochemical-317 only and physical and biogeochemical jointly) on the skill of both the one-318 way and two-way coupled models. The various experiments used exactly the 319 same model configuration, apart from the difference in the coupling between 320 physics and biogeochemistry. The experiments all started from the same 321 initial value conditions on the 01/09/2017 to allow a 4 month spin-up time for 322 the final 2018 simulation. The initial values were provided by the 2016-2018 323 free simulation (using bio-optical module) from the study of [31]. Finally, 324 Tab.2 provides a list of all experiments with their abbreviated names that 325 we will use in the paper. 326

327 2.6. Skill metrics

The performance of the different simulations will be evaluated using two skill metrics. The first metric is the model bias (ΔQ_{mo}) :

$$\Delta Q_{mo} = \langle Q_m - Q_o \rangle \tag{1}$$

where Q_o are the observations mapped into the model grid and the Q_m are the corresponding model outputs. The second metric is the bias-corrected root mean square difference (BC RMSD, $\Delta_{RD}Q_{mo}$):

$$\Delta_{RD}Q_{mo} = \sqrt{\langle (Q_m - Q_o - \Delta Q_{mo})^2 \rangle}.$$
 (2)

Table 2: The different experiments compared in this study. The first column shows the abbreviated experiment name, the second column indicates whether the two-way coupling is used and the following columns list the assimilated data. The table uses the following abbreviations: satellite: "sat", Cabot glider: "Cabot", EN4 dataset: "EN4", temperature: "T", sea surface temperature: "SST", salinity: "S", chlorophyll a: "Chl a".

abbreviation	two-way	SST (sat./in situ)	T & S (EN4)	T & S (Cabot)	Chl a (sat.)	Chl a (Cabot)
free 1-way	no	no	no	no	no	no
free 2-way	yes	no	no	no	no	no
phys DA 1-way	no	yes	yes	yes	no	no
phys DA 2-way	yes	yes	yes	yes	no	no
chl DA 1-way	no	no	no	no	yes	yes
chl DA 2-way	yes	no	no	no	yes	yes
phys+chl DA 1-way	no	yes	yes	yes	yes	yes
phys+chl DA 2-way	yes	yes	yes	yes	yes	yes

333 3. Results and Discussion

334 3.1. The impact of the two-way coupling and assimilation on the simulated 335 physics

The reference one-way coupled model simulates well the seasonal increase 336 of temperature in the surface ocean in late-spring / summer (Fig.2:A, Fig.3). 337 The novel two-way coupling further increased the temperature in the upper 338 20m by around 1°C (Fig.2:B, Fig.3). This is a relatively major change with 339 respect to the reference run, when compared to the changes introduced to 340 the simulated temperature by the physical data assimilation during the same 341 period of the year (Fig.2:D, for all the assimilative runs see Fig.S2-S3 in the 342 Supporting Information (SI). The increase in the upper ocean temperature 343 in the two-way coupled model cannot be explained by the enhanced shortwave 344 radiation flux in the water column, since the bio-optical module and the 345 ERA5 short-wave radiation product, which forces the one-way coupled run, 346 have a negligible mutual bias ([31]). Therefore, the temperature increase 347 is likely a consequence of an increased rate of absorption inside the upper 348 oceanic layer. The increased absorption in the two-way coupled run was 349 anticipated since: a) the bio-optical module appears to have higher level of 350 light attenuation near the water surface than the satellite observations used 351 to force the physics in the one-way coupled run (this was observed for 490nm 352 wavelength in Fig.5:A of [31]), b) the "broadband" visible light attenuation 353 in the one-way coupled run was represented by the satellite K_d for 490nm 354

wavelength, but K_d at 490nm wavelength is clearly an underestimate of the K_d for the 400-700nm waveband (see Fig.5:B of [31]).

The impact of phytoplankton biomass on the simulated temperature can 357 be analysed by comparing the chlorophyll-assimilative run (chl DA 2-way) 358 with its corresponding two-way coupled free run (free 2-way): In the late 359 spring - summer, the assimilation of chlorophyll into the two-way coupled 360 model removes a large amount of phytoplankton biomass from the mixed 361 layer (see Fig.S4:B of SI), increases the light penetration into the water col-362 umn and heats up a deeper oceanic layer than the free run (Fig.2:B-C). The 363 temperature is then raised in the 20-60m depth range by 0.1-0.2°C in the 364 summer and by less than that in the late spring (see Fig.S5 of SI). The extra 365 heat captured by the two-way coupled model near the ocean surface shallows 366 the MLD (Fig.4:B, Fig.S6 of SI), which is indicative of important changes to 367 mixing of biogeochemical tracers in the upper ocean. 368

Outside of the late spring - summer, both two-way coupling (Fig.2:B) and 369 chlorophyll assimilation (Fig.2:C) have comparably smaller impact on the 370 simulated oceanic temperature than the physical data assimilation (Fig.2:D. 371 see also Fig.S2-S4 of SI). The impact of physical data assimilation is most 372 important around the winter, when it corrects a negative temperature bias 373 $(\sim -0.5^{\circ}\text{C})$ of the physical model (Fig.2-3, Fig.S3-S4 of SI). The physical 374 data assimilation influences the simulated temperature more evenly across 375 the water column than the bio-optical module (Fig.2), which is likely a com-376 bination of model dynamical response to the temperature increments in the 377 mixed layer and some assimilated sub-surface data (EN4 and Cabot glider). 378 If the reanalysis state is sufficiently stable with respect to the model dynam-379 ics, it is known ([46, 31, 36]) that, within NEMOVAR on the NWE Shelf, 380 the assimilated variables in the reanalysis tend to converge to the assimi-381 lated data. This is evident in the Fig.5:D, Fig.S3,S7 of SI, comparing the 382 SST of the physical data assimilation runs with the assimilated satellite SST 383 observations. 384

We evaluated (Fig.6 and Fig.7) the skill of both the two-way coupled 385 model and the different assimilative experiments to represent temperature 386 and salinity on the NWE Shelf. Fig.6 compares the two-way coupled free and 387 chlorophyll-assimilative runs with the temperature and salinity measured by 388 the Cabot glider mission in the central North Sea during late spring - summer 389 of 2018 (for more details about the mission see [36], Fig.S1 of SI and Tab.1). 390 Glider-observed temperature is warmer in the upper 30-40m of the water col-391 umn than the temperature simulated by the one-way coupled model, whereas 392



Figure 2: Panel A shows Hovmöller diagram (time on the x-axis vs depth on the yaxis) for the temperature (°C) of the one-way coupled free run, where the values for each day and depth represent the horizontal spatial averages throughout the NWE Shelf (bathymetry < 200m). Panels B-D show the same Hovmöller diagrams as panel A, but for the temperature differences between the two-way coupled, or assimilative runs and the reference, free one-way coupled model run from the panel A (for the abbreviations used in the titles see Tab.2). In particular, panels B-D compare the impact of two-way coupling on the simulated temperature (panel B), joint impact of chlorophyll-assimilation and two-way coupling on the simulated temperature (panel C) and the impact of physical data assimilation on the simulated temperature (panel D). The yellow line in the panel D shows the MLD of the physical data assimilative run to indicate the vertical scale of impact of the SST assimilation.



Figure 3: The 2018 time-series of SST averaged throughout the NWE Shelf compared between the one-way and two-way coupled free simulations, and the VIIRS satellite/in situ data. To consistently compare the model simulations with the observed SST, the model outputs were masked wherever there were missing observations. The missing satellite observations are due to the movements of clouds and atmospheric disturbances and the missing values are responsible for the small time-scale fluctuations in the different curves shown in the plot.



Figure 4: Panel A shows the mixed layer depth (MLD, in m) of the one-way coupled reference free run. The MLD values are averaged for the spring bloom period between March-May 2018. Panels B-C show the relative changes in MLD carried by the two-way coupled free run (panel B) and physical data assimilation into the one-way coupled model (panel C). Both panels B,C show the difference (in m) between the MLD of the two-way coupled, or physical data assimilative run and the one-way coupled model free run (panel A). The blue color in panels B-C (negative values) indicates shallowing of MLD, whilst the red color (positive values) indicates deepening of MLD. The black line shows the boundary of the continental shelf (bathymetry < 200m).



Figure 5: The assimilated 2018 median satellite data for SST (panel A, in $^{\circ}$ C) and the corresponding model to VIIRS/in situ SST differences (panels B-D, in $^{\circ}$ C) for one-way coupled model free run (panel B), two-way coupled model free run (panel C) and physical data assimilation into the one-way coupled free run (panel D). The masked values indicate the regions where there was no assimilation of VIIRS/in situ data into the model.

the opposite is true beneath 40m depth (Fig.6:A). This means the observed 393 thermocline represents a larger gradient in temperature than the simulated 394 thermocline. The bio-optical module substantially (by > 1°C) heats up the 395 upper 20-30m layer, increasing the vertical temperature gradient (Fig.6:C), 396 however the near-surface temperature of the two-way coupled run rises well 397 above the levels observed by the glider (Fig.6:D). The thermocline of the 398 two-way coupled model free run appears to be located above the glider ther-399 mocline (e.g. Fig.6:D) and the impact of the two-way coupling on the model 400 skill in representing glider temperature is somewhat mixed (it improves bias, 401 but degrades BC RMSD, Fig.7:A). The skill validation presented in Fig.7 402 shows similarly mixed results: the summer temperature bias is improved 403 across the EN4 and AlterEco glider data, but degraded relative to the VI-404 IRS/in situ data (see also Fig.3), with the BC RMSD consistently degraded 405 across the different validation data. The Fig.7:A indicates that the two-406 way coupling produces better results for sub-surface summer temperature. 407 than for SST (VIIRS/in situ data). The two-way coupling has a similarly 408 mixed impact on the free run skill to represent summer salinity (Fig.7:C), 409 and both small ($< 0.05^{\circ}$ C) and mixed impact on winter temperature and 410 salinity (Fig.7:B,D, for temperature see also Fig.2-3). However, it should be 411 noted that chlorophyll assimilation into the two-way coupled model slightly 412 improves the skill of the free run in representing temperature and salinity 413 across most of the data and throughout the whole year 2018 (Fig.7). Finally, 414 the comparison with the non-assimilated temperature validation data clearly 415 demonstrates that the physical data assimilation improves the model skill in 416 temperature both in summer and winter half-year (Fig.7:A-B) and also the 417 model skill in salinity in the winter half-year (Fig.7:D). 418

419 3.2. The impact of the two-way coupling and assimilation on biogeochemistry

As the days in spring become longer, the layer that is effectively lit by 420 the sunlight expands into the water column, whilst the effective mixing depth 421 shrinks. It is often assumed, that the effective mixing depth reaching a critical 422 threshold marks the onset of the spring bloom (Fig.1). This process might be 423 misrepresented by the one-way coupled reference free simulation, which could 424 be why the model shows on the NWE Shelf late (by $\sim 1 \text{ month}$) and intense 425 blooms (Fig.8, see also [31, 36]). The effective mixing depth has often been 426 interpreted as the seasonal MLD (this is the frequent understanding of the 427 critical depth hypothesis of [75]), but it is assumed that on the NWE Shelf 428 the onset of the bloom might be better described by the critical turbulence 429



Figure 6: Hovmöller diagram for temperature (°C) along the trajectory covered by the Cabot glider in the central North Sea during an early May to mid-August 2018 mission. The right-hand panels (B,D,F) show the temperature differences between the free one-way coupled run (panel B), free two-way coupled run (panel D), the chlorophyll assimilation into the two-way coupled model (panel F) and the Cabot glider observations (model minus glider). The left hand panels (A,C,E) show the differences between the observations, or model simulations and the reference, free one-way coupled model run. The purpose of the left-hand panels is to show the desired changes to the one-way coupled model (panel A) and how these changes are realized by the biogeochemical feedback in the free run (panel C) and in the chlorophyll-assimilative run (panel E). The main advantage of those left-hand (A,C,E) panels is that they allow relatively easy interpretation of the dynamical changes introduced to the reference run by the biogeochemical feedback to physics and/or data assimilation.



Figure 7: Skill of the different model simulations to represent temperature (°C, panels A-B) and practical salinity (panels C-D). The skill is measured by bias (x-axis, Eq.1) and BC RMSD (y-axis, Eq.2). The skill is evaluated for two half-year periods of 2018, the "summer" (panels A,C) defined as May-October and the "winter" (panels B,D) defined as November-April (data averaged through January-April 2018 and November-December 2018). The different simulations are represented by different colors: free run of the oneway coupled model (red), free run of the two-way coupled model (blue), assimilation of chlorophyll into the two-way coupled model (cyan), physical data assimilation into the one-way coupled model (lime), physical data assimilation into the two-way coupled model (grey) and joint physical data-chlorophyll assimilation into the two-way coupled model (orange). The different markers show comparison with different data-sets: the star stands for the VIIRS/in situ SST, the circle for the Cabot glider observations, the diamond for the remaining available glider observations (the 2018 AlterEco mission without Cabot) and the cross for the EN4 data-set. The data (SST, Cabot, EN4) which were assimilated in some of the simulations were used to validate only the simulations that avoided their assimilation.



Figure 8: The 2018 time-series of surface chlorophyll a concentrations (mg/m³) averaged throughout the NWE Shelf compared between the one-way and two-way coupled free simulations, the satellite data, as well as with the NSBC climatological data-set. The satellite data were considered only in the March-September period as the data outside this period are scarce and limited only to the southern part of the NWE domain. The small time-scale fluctuations in the satellite data are due to the missing values caused by the movement of clouds and atmospheric disturbances.

hypothesis ([40]). In the critical turbulence hypothesis the bloom starts when
the turbulent mixing in the upper ocean drops beneath a critical level, whilst
the effective rate of turbulent mixing is largely decoupled from the seasonal
MLD ([40, 41, 42]).

The implementation of the bio-optical module was shown to shallow the 434 MLD (Fig.4), but it can also reduce convection within the mixed layer and 435 the turbulent mixing. The starting hypothesis of this work was that the ex-436 tra heat captured in the upper oceanic layer could trigger an earlier bloom 437 and improve the ERSEM skill. Fig.8, Fig.9:B and Fig.10:C-D show that the 438 changes to the simulated physics introduced through the two-way coupled 439 model indeed trigger an earlier phytoplankton bloom, but the difference in 440 the bloom timing is only on the scale of several days, rather than weeks. 441 However, the shift to the bloom timing has an impact on many subsequent 442 features, such as the deep chlorophyll maxima (e.g. [36]), so the changes 443 to the bloom onset can gradually propagate to the subsurface chlorophyll 444 (Fig.9:C). The model skill to simulate chlorophyll is improved by the two-445 way coupling quite notably in the central North Sea and the period covered 446 by the Cabot glider (Fig.11:A), however comparisons with other data spread 447 throughout the year 2018 (satellite ocean color, remaining AlterEco gliders 448 and the NSBC climatology) show only small improvement (Fig.11:A). The 449

⁴⁵⁰ modest improvement to the timing of the (delayed) spring bloom through ⁴⁵¹ the changed mixing is certainly a disappointment, and we suspect that to ⁴⁵² introduce a larger correction to the timing of the bloom it would be nec-⁴⁵³ essary to either improve the physical model mixing scheme, or to improve ⁴⁵⁴ some key ERSEM parameters and processes, such as P-I curves, the max-⁴⁵⁵ imum chlorophyll-to-carbon ratios, zooplankton grazing and representation ⁴⁵⁶ of plankton mixotrophy ([29]).

Although the (modest) improvements to the simulated chlorophyll by the 457 two-way coupled model originate from its changes to the simulated physics 458 (i.e. vertical mixing), the physical data assimilation, which substantially im-459 proves the simulated physics (Fig.7) does not improve (even slightly degrades) 460 the model skill in chlorophyll (Fig.11:A). This is likely because the physical 461 data assimilation is for large part the assimilation of SST. The improvement 462 in the ecosystem model skill depends mostly on the vertical mixing and lim-463 ited changes to vertical mixing are expected by assimilating SST. Assimilated 464 subsurface temperature and salinity data are quite sparse, and have only a 465 limited impact on the modelled biogeochemistry. In the case of the Cabot 466 glider "case-study" presented in Fig.10 (for a more complete view see Fig.S8 467 of SI), the glider temperature and salinity assimilation did not improve the 468 simulated chlorophyll at the glider locations (Fig.11:A) mostly because the 469 impact of physics on biogeochemistry needs some spin-up time. In fact in 470 the last part of the glider mission period (late July-August in Fig. 10:E) the 471 physical assimilation has some potential to improve the chlorophyll concen-472 trations, as was demonstrated by the assimilation of the same Cabot glider 473 data in Fig.6E of [36]. Finally, the chlorophyll assimilation dominates over 474 both physical assimilation and two-way coupling in its impact on the simu-475 lated chlorophyll concentrations across the whole water column and the whole 476 simulation year (Fig.9:D and Fig.S9 of SI). Since the chlorophyll assimila-477 tion is almost entirely based on the satellite ocean color, chlorophyll beneath 478 the mixed layer is updated through the model dynamical response to the 479 assimilation (e.g. vertical mixing). Similarly to temperature, the chlorophyll 480 reanalyses look very similar to the assimilated data (Fig.12:B-C, Fig.S5 and 481 Fig.S10 of SI) and also validate much better than the free runs relative to 482 the non-assimilated AlterEco glider data (Fig.11:A). 483

We validated the model simulation of additional biogeochemical variables with available observational data: oxygen, nitrate, phosphate, silicate and CO₂ fugacity. The oxygen concentrations are mostly driven by the primary productivity, respiration and outgassing, which largely depends on the sea

temperature. The two-way coupled model improves the model skill in repre-488 senting Cabot oxygen (Fig.11:B), which is likely triggered by the fact that the 489 same simulation improves both Cabot chlorophyll (Fig.11:A) and the temper-490 ature bias (Fig.7:A). Equivalently, model skill in representing Cabot glider 491 oxygen can be improved by assimilating physical data into the model (phys 492 DA 1-way), and it is to some degree also improved by assimilating chloro-493 phyll (chl DA 1-way, chl DA 2-way), with the best performance achieved 494 when both the physical data and chlorophyll are assimilated into the model 495 (Fig.11:B). However, the Cabot glider study is specific, since the glider mis-496 sion took place in the period of the largest discrepancy in the simulated 497 and observed productivity (Fig.8) and the oxygen concentrations were mea-498 sured by the same glider that provided temperature, salinity and chlorophyll 499 data for assimilation. For the remaining non-assimilated AlterEco gliders 500 the impact of two-way coupling and assimilation on simulated oxygen is less 501 clear (Fig.11:B), i.e. even though AlterEco chlorophyll is improved by the 502 chlorophyll-only assimilative runs (Fig.11:A) they mostly degrade simulated 503 oxygen (Fig.11:B). This is likely due to the complex relationship between phy-504 toplankton chlorophyll and oxygen (see [36]), which includes respiration of 505 oxygen by the higher trophic-level species (in ERSEM it is zooplankton and 506 heterotrophic bacteria). However, improved representation of temperature 507 consistently improves model oxygen bias across all the used data (Fig.11:A), 508 which indicates that an important part of oxygen bias is due to model biases 509 in temperature and not due to errors in the simulated biogeochemistry. 510

Besides oxygen, we looked at the model skill in how it represents the 511 surface CO_2 fugacity, which is influenced by the model skill in simulating 512 primary productivity and sea temperature (gas solubility). Fig.11:C shows 513 that CO_2 fugacity is substantially improved by all the runs that included 514 chlorophyll assimilation, which indicates that the assimilation of chlorophyll 515 improved the phytoplankton carbon biomass and therefore the simulated 516 carbon cycle (see also [46]). The physical data-only assimilative runs, and 517 the two-way coupled free run, had more limited impact on the model skill 518 to represent surface CO_2 fugacity, but they sometimes reduced the model 519 bias in CO_2 fugacity. Both the two-way coupling and the physical assimi-520 lation, have a relatively small impact on the nitrate and phosphate concen-521 trations (Fig.11:D-E), however the changed phytoplankton biomass through 522 the chlorophyll assimilation lowers the nitrate and phosphate concentrations 523 at the NSBC data-set locations. This has a positive impact on the nitrate 524 bias and a negative impact on the phosphate bias (Fig.11:D-E). Silicate is 525

⁵²⁶ impacted more by the physical data assimilation than nitrate and phosphate, ⁵²⁷ but it is mostly degraded by all the assimilative runs (Fig.11:F).

528 4. Summary

In this work we used a recently developed bio-optical module to improve 529 the representation of oceanic heat fluxes and to introduce a biogeochemical 530 feedback to the physical marine model (we call the model with such feedback 531 "a two-way coupled model"). We have estimated the scale of the biogeochem-532 ical impact on the simulated physics and we have shown that in the upper 533 oceanic layer, in the late spring - summer period, the feedback is comparable 534 to the physical data assimilation in its impact on the simulated tempera-535 ture. The bio-optical module increases the heat captured in the upper part 536 of the water column, steepens the vertical temperature gradient and shal-537 lows the mixed layer depth. We have shown that the changes introduced by 538 the bio-optical module into the physical marine model have a mixed impact 530 on the physical model skill. The skill is however (slightly) improved by the 540 chlorophyll assimilation into the two-way coupled model and substantially 541 improved by the physical data assimilation. 542

The increased stratification of the water column and the shallowed mixed 543 layer depth have a modest positive impact on the timing of the late bloom 544 displayed by the biogeochemical model. The shift in the timing of the bloom 545 in the two-way coupled model improves the model skill in representing chloro-546 phyll. We conclude that, for a more substantial improvement of the timing 547 of the bloom, it will be necessary to either improve the physical model mix-548 ing scheme, or to improve the process description, or parametrization of the 549 biogeochemical model. We have expanded our analysis to include other bio-550 geochemical tracers, and we have found that the two-way coupled model and 551 the physical data assimilation may sometimes help improve the agreement of 552 simulated oxygen concentrations and CO_2 fugacity with observations, both 553 due to improved simulation of the sea water temperature (saturation levels) 554 and productivity. 555

This study provides important evidence to support the inclusion of twoway coupling into future operational models of the NWE Shelf. Furthermore, the physical-biogeochemical assimilative runs on the NWE Shelf, including this work, are typically only weakly coupled (for one recent exception see [76]), in the sense that the physical and the biogeochemical variables are updated independently and interact only through the model dynamics. The



Figure 9: Impact of two-way coupling and assimilation on the simulated chlorophyll concentrations (mg/m³). Panel A shows Hovmöller diagram (time on the x-axis vs depth on the y-axis) for the one-way coupled model free run, where the values for each day and depth represent the horizontal spatial averages throughout the NWE Shelf (bathymetry < 200m). Panels B-D show the same Hovmöller diagrams, but for the difference between the specific simulation and the reference, free one-way coupled run. The purpose of the panels B-D is to provide an understanding of how the two-way coupling (panel B), the biogeochemical feedback (panel C) and the chlorophyll-assimilation (panel D) influence the chlorophyll concentrations of the reference free one-way coupled run. The yellow line in the panel D shows the mixed layer depth, providing the boundary of the region in which the ocean color assimilation directly updates the simulated chlorophyll.



Figure 10: Hovmöller diagram for chlorophyll concentrations (mg/m^3) along the Cabot glider trajectory in the central North Sea during an early May to mid-August 2018 mission. The right-hand panels (B,D,F) show the chlorophyll differences between the free one-way coupled model run (panel B), free two-way coupled model run (panel D), the physical data assimilation into the one-way coupled model (panel F), and the Cabot glider observations (model minus glider). The left hand panels (A,C,E) show the differences between the observations, or model simulations and the reference, free one-way coupled model run. The purpose of the left-hand panels is to show the desired changes to the one-way coupled model (panel A) and how these changes are realized by the biogeochemical feedback in the free run (panel C) and in the physical data-assimilative run (panel E). The main advantage of those left-hand panels is that they allow relatively easy interpretation of the dynamical changes introduced to the reference run by the biogeochemical feedback to physics and/or data assimilation.



Figure 11: Skill of the different model simulations to represent chlorophyll a (mg/m³, panel A), oxygen (mmol/m³, panel B), CO₂ fugacity (μ bar, panel C), nitrate (mmol/m³, panel D), phosphate (mmol/m³, panel E) and silicate (mmol/m³, panel F) concentrations. The skill is measured by bias (x-axis, Eq.1) and BC RMSD (y-axis, Eq.2). The skill is evaluated for the full year 2018. The different simulations are represented by different colors: free run of the one-way coupled model (red), free run of the two-way coupled model (blue), assimilation of chlorophyll into the one-way coupled model (cyan), physical data assimilation into the one-way coupled model (grey), joint physical data-chlorophyll assimilation into the one-way coupled model (green) and joint physical data-chlorophyll assimilation into the two-way coupled model (orange). The different markers show comparison with different data-sets: the star stands for the satellite ocean color data, the circle for the Cabot glider observations, the diamond for the remaining available glider observations (the 2018 AlterEco mission without Cabot), the cross for the SOCAT data and the square for the NSBC climatological data-set.



Figure 12: The 2018 mean surface chlorophyll concentrations (in mg/m³). The different panels compare: the one-way coupled model free run (panel A), the chlorophyll assimilation into the one-way coupled model free run (panel B), and the assimilated satellite ocean color observations (panel C). In the annual averaging we masked the model outputs wherever the satellite data were missing. The black line shows the continental shelf boundary (bathymetry < 200m).

interaction between physics and biogeochemistry via the coupled model dy-562 namics has been strengthened through the two-way coupling, but it would 563 be much more efficient if the assimilative updates to the physics and bio-564 geochemistry interacted directly through their cross-covariances, or a bal-565 ancing component within a data assimilation system. Such scheme is called 566 "strongly coupled", and would provide the physical assimilation with both 567 faster and greater impact on the biogeochemical model skill, and vice versa. 568 Future work will use the two-way coupled model and expand the data assim-569 ilation scheme to include such strong coupling into our operational system. 570

571 Acknowledgments

This work was supported by a Natural Environment Research Council 572 (NERC) funded project of the Marine Integrated Autonomous Observing 573 Systems (MIAOS) programme: Combining Autonomous observations and 574 Models for Predicting and Understanding Shelf seas (CAMPUS). It also ben-575 efitted from another NERC funded project Alternative Framework to Assess 576 Marine Ecosystem Functioning in Shelf Seas (AlterECO, http://projects.noc-577 .ac.uk/altereco/), grant no. NE/P013899/1. The work also benefited from 578 the Copernicus Marine Environment Monitoring Service (CMEMS) funded 579 projects OPTIcal data Modelling and Assimilation (OPTIMA) and NOWMAPS. 580

Furthermore, this work was also partially funded by the SEAMLESS project, 581 which received funding from the European Union's Horizon 2020 research 582 and innovation programme under grant agreement No 101004032. We would 583 like to thank Dawn Ashby for drawing the schematic Fig.1. The ocean color 584 data were provided by the European Space Agency Climate Initiative "Ocean 585 Color" (https://esa-oceancolour-cci.org/). The glider data used in the study 586 (doi:10.5285/b57d215e-065f-7f81-e053-6c86abc01a82 and doi:10.5285/b58e83f0-587 d8f3-4a83-e053-6c86abc0bbb5) are publicly available on https://www.bodc.ac.uk/-588 data/published_data_library/catalogue/. The model was forced by the atmo-580 spheric ERA5 product of The European Centre for Medium-Range Weather 590 Forecasts (ECMWF, https://www.ecmwf.int/). The river forcing data used 591 by the model were prepared by Sonja van Leeuwen and Helen Powley as part 592 of UK Shelf Seas Biogeochemistry programme (contract no. NE/K001876/1) 593 of the NERC and the Department for Environment Food and Rural Affairs 594 (DEFRA). We acknowledge use of the MONSooN system, a collaborative 595 facility supplied under the Joint Weather and Climate Research Programme, 596 a strategic partnership between the Met Office and the NERC. The differ-597 ent outputs for the free run simulations and reanalyses are stored on the 598 MONSooN storage facility MASS and can be obtained upon request. 599

600 References

- [1] M. Gehlen, R. Barciela, L. Bertino, P. Brasseur, M. Butenschön, F. Chai,
 A. Crise, Y. Drillet, D. Ford, D. Lavoie, et al., Building the capacity for
 forecasting marine biogeochemistry and ecosystems: recent advances
 and future developments, Journal of Operational Oceanography 8 (sup1)
 (2015) s168-s187.
- [2] D. Ford, S. Kay, R. McEwan, I. Totterdell, M. Gehlen, Marine biogeo chemical modelling and data assimilation for operational forecasting,
 reanalysis, and climate research, New Frontiers in Operational Oceanog raphy (2018) 625–652.
- [3] C. Heinze, M. Gehlen, Modeling ocean biogeochemical processes and
 the resulting tracer distributions, in: International Geophysics, Vol. 103,
 Elsevier, 2013, pp. 667–694.
- [4] U. Riebesell, A. Körtzinger, A. Oschlies, Sensitivities of marine carbon

- fluxes to ocean change, Proceedings of the National Academy of Sciences 106 (49) (2009) 20602–20609.
- ⁶¹⁶ [5] Z. Jin, T. P. Charlock, W. L. Smith Jr, K. Rutledge, A parameterization ⁶¹⁷ of ocean surface albedo, Geophysical research letters 31 (22) (2004).
- [6] A. Morel, Optical modeling of the upper ocean in relation to its
 biogenous matter content (case i waters), Journal of geophysical research: oceans 93 (C9) (1988) 10749–10768.
- [7] J.-y. Simonot, E. Dollinger, H. Le Treut, Thermodynamic-biologicaloptical coupling in the oceanic mixed layer, Journal of Geophysical Research: Oceans 93 (C7) (1988) 8193–8202.
- [8] S. Sathyendranath, A. D. Gouveia, S. R. Shetye, P. Ravindran, T. Platt,
 Biological control of surface temperature in the arabian sea, Nature
 349 (6304) (1991) 54.
- [9] A. M. Edwards, D. G. Wright, T. Platt, Biological heating effect of
 a band of phytoplankton, Journal of Marine Systems 49 (1-4) (2004)
 89–103.
- [10] M. Manizza, C. Le Quéré, A. J. Watson, E. T. Buitenhuis, Bio-optical feedbacks among phytoplankton, upper ocean physics and sea-ice in a global model, Geophysical Research Letters 32 (5) (2005).
- [11] C. Sweeney, A. Gnanadesikan, S. M. Griffies, M. J. Harrison, A. J.
 Rosati, B. L. Samuels, Impacts of shortwave penetration depth on largescale ocean circulation and heat transport, Journal of Physical Oceanography 35 (6) (2005) 1103–1119.
- [12] M. Lengaigne, C. Menkes, O. Aumont, T. Gorgues, L. Bopp, J.-M.
 André, G. Madec, Influence of the oceanic biology on the tropical pacific
 climate in a coupled general circulation model, Climate Dynamics 28 (5)
 (2007) 503-516.
- [13] L. Zhai, C. Tang, T. Platt, S. Sathyendranath, Ocean response to attenuation of visible light by phytoplankton in the gulf of st. lawrence,
 Journal of Marine Systems 88 (2) (2011) 285–297.

- [14] A. Turner, M. Joshi, E. Robertson, S. Woolnough, The effect of arabian
 sea optical properties on sst biases and the south asian summer monsoon
 in a coupled gcm, Climate dynamics 39 (3-4) (2012) 811-826.
- ⁶⁴⁷ [15] J. E. Lovelock, R. Maggs, R. Rasmussen, Atmospheric dimethyl sulphide ⁶⁴⁸ and the natural sulphur cycle, Nature 237 (5356) (1972) 452–453.
- [16] R. J. Charlson, J. E. Lovelock, M. O. Andreae, S. G. Warren, Oceanic
 phytoplankton, atmospheric sulphur, cloud albedo and climate, Nature
 326 (6114) (1987) 655–661.
- [17] K. D. Six, S. Kloster, T. Ilyina, S. D. Archer, K. Zhang, E. MaierReimer, Global warming amplified by reduced sulphur fluxes as a result
 of ocean acidification, Nature Climate Change 3 (11) (2013) 975–978.
- [18] J. Schwinger, J. Tjiputra, N. Goris, K. D. Six, A. Kirkevåg, Ø. Seland,
 C. Heinze, T. Ilyina, Amplification of global warming through ph dependence of dms production simulated with a fully coupled earth system
 model, Biogeosciences 14 (15) (2017) 3633.
- [19] T. W. Wilson, L. A. Ladino, P. A. Alpert, M. N. Breckels, I. M. Brooks,
 J. Browse, S. M. Burrows, K. S. Carslaw, J. A. Huffman, C. Judd, et al.,
 A marine biogenic source of atmospheric ice-nucleating particles, Nature
 525 (7568) (2015) 234–238.
- [20] J. Lovelock, Gaia: A new look at life on earth, Oxford Paperbacks, 1979.
- [21] J. Lovelock, The ages of Gaia: A biography of our living earth, Oxford
 University Press, USA, 2000.
- [22] A. Borges, L.-S. Schiettecatte, G. Abril, B. Delille, F. Gazeau, Carbon
 dioxide in european coastal waters, Estuarine, Coastal and Shelf Science
 70 (3) (2006) 375–387.
- [23] R. A. Jahnke, Global synthesis, in: Carbon and nutrient fluxes in con tinental margins, Springer, 2010, pp. 597–615.
- [24] O. Legge, M. Johnson, N. Hicks, T. Jickells, M. Diesing, J. Aldridge,
 J. Andrews, Y. Artioli, D. C. Bakker, M. T. Burrows, et al., Carbon on
 the northwest european shelf: Contemporary budget and future influences, Frontiers in Marine Science 7 (2020) 143.

- $_{675}$ [25] G. Madec, et al., Nemo ocean engine (2015).
- ⁶⁷⁶ [26] J. Bruggeman, K. Bolding, A general framework for aquatic biogeochem-⁶⁷⁷ ical models, Environmental modelling & software 61 (2014) 249–265.
- ⁶⁷⁸ [27] J. Bruggeman, K. Bolding, Framework for aquatic biogeochemical models (2020). doi:http://doi.org/10.5281/zenodo.3817997.
- [28] J. Baretta, W. Ebenhöh, P. Ruardij, The european regional seas ecosystem model, a complex marine ecosystem model, Netherlands Journal of Sea Research 33 (3-4) (1995) 233-246.
- [29] M. Butenschön, J. Clark, J. N. Aldridge, J. I. Allen, Y. Artioli, J. Blackford, J. Bruggeman, P. Cazenave, S. Ciavatta, S. Kay, et al., Ersem
 15.06: a generic model for marine biogeochemistry and the ecosystem
 dynamics of the lower trophic levels, Geoscientific Model Development
 9 (4) (2016) 1293–1339.
- [30] P. M. L. Marine Systems Modelling Group, European regional seas
 ecosystem model (2020). doi:http://doi.org/10.5281/zenodo.3817997.
- [31] J. Skákala, J. Bruggeman, R. J. Brewin, D. A. Ford, S. Ciavatta, Im proved representation of underwater light field and its impact on ecosys tem dynamics: a study in the north sea, Journal of Geophysical Re search: Oceans (2020) e2020JC016122.
- [32] W. W. Gregg, N. W. Casey, Skill assessment of a spectral ocean–
 atmosphere radiative model, Journal of Marine Systems 76 (1-2) (2009)
 49–63.
- [33] W. W. Gregg, C. S. Rousseaux, Directional and spectral irradiance in
 ocean models: effects on simulated global phytoplankton, nutrients, and
 primary production, Frontiers in Marine Science 3 (2016) 240.
- [34] W. W. Gregg, C. S. Rousseaux, Simulating pace global ocean radiances,
 Frontiers in Marine Science 4 (2017) 60.
- [35] J. Bruggeman, J. Skákala, J. Lawrence, D. Ford, R. Brewin, S. Ciavatta,
 Fabm-spectral (2021). doi:http://doi.org/10.5281/zenodo.4594277.

- [36] J. Skákala, D. A. Ford, J. Bruggeman, T. Hull, J. Kaiser, R. R. King,
 B. R. Loveday, M. R. Palmer, T. J. Smyth, C. A. J. Williams, S. Ciavatta, Towards a multi-platform assimilative system for ocean biogeochemistry, Earth and Space Science Open Archive ESSOAr, submitted
 to JGR-Oceans (2021).
- [37] M. J. Lutz, K. Caldeira, R. B. Dunbar, M. J. Behrenfeld, Seasonal rhythms of net primary production and particulate organic carbon flux to depth describe the efficiency of biological pump in the global ocean, Journal of Geophysical Research: Oceans 112 (C10) (2007).
- [38] S. A. Henson, J. P. Dunne, J. L. Sarmiento, Decadal variability in north atlantic phytoplankton blooms, Journal of Geophysical Research: Oceans 114 (C4) (2009).
- [39] M. J. Behrenfeld, E. S. Boss, Student's tutorial on bloom hypotheses
 in the context of phytoplankton annual cycles, Global change biology
 24 (1) (2018) 55–77.
- [40] J. Huisman, P. van Oostveen, F. J. Weissing, Critical depth and critical turbulence: two different mechanisms for the development of phytoplankton blooms, Limnology and oceanography 44 (7) (1999) 1781–1787.
- [41] J. J. Waniek, The role of physical forcing in initiation of spring blooms
 in the northeast atlantic, Journal of Marine Systems 39 (1-2) (2003)
 57–82.
- [42] A. Ferreira, H. Hátún, F. Counillon, M. Payne, A. Visser, Synoptic-scale analysis of mechanisms driving surface chlorophyll dynamics in the north atlantic, Biogeosciences 12 (11) (2015) 3641–3653.
- [43] J. R. Taylor, R. Ferrari, Shutdown of turbulent convection as a new criterion for the onset of spring phytoplankton blooms, Limnology and Oceanography 56 (6) (2011) 2293–2307.
- [44] T. J. Smyth, I. Allen, A. Atkinson, J. T. Bruun, R. A. Harmer, R. D.
 Pingree, C. E. Widdicombe, P. J. Somerfield, Ocean net heat flux influences seasonal to interannual patterns of plankton abundance, PloS one
 9 (6) (2014).

- [45] D. A. Ford, J. van der Molen, K. Hyder, J. Bacon, R. Barciela,
 V. Creach, R. McEwan, P. Ruardij, R. Forster, Observing and modelling
 phytoplankton community structure in the north sea, Biogeosciences
 14 (6) (2017) 1419–1444.
- [46] J. Skákala, D. Ford, R. J. Brewin, R. McEwan, S. Kay, B. Taylor,
 L. de Mora, S. Ciavatta, The assimilation of phytoplankton functional
 types for operational forecasting in the northwest european shelf, Journal of Geophysical Research: Oceans 123 (8) (2018) 5230-5247.
- [47] E. O'Dea, R. Furner, S. Wakelin, J. Siddorn, J. While, P. Sykes, R. King,
 J. Holt, H. Hewitt, The co5 configuration of the 7 km atlantic margin
 model: large-scale biases and sensitivity to forcing, physics options and
 vertical resolution, Geoscientific Model Development 10 (8) (2017) 2947.
- [48] J. Siddorn, R. Furner, An analytical stretching function that combines
 the best attributes of geopotential and terrain-following vertical coordinates, Ocean Modelling 66 (2013) 1–13.
- [49] D. Storkey, E. Blockley, R. Furner, C. Guiavarc'h, D. Lea, M. Martin,
 R. Barciela, A. Hines, P. Hyder, J. Siddorn, Forecasting the ocean state
 using nemo: The new foam system, Journal of operational oceanography
 3 (1) (2010) 3–15.
- ⁷⁵⁴ [50] H.-J. Lenhart, D. K. Mills, H. Baretta-Bekker, S. M. Van Leeuwen,
 ⁷⁵⁵ J. Van Der Molen, J. W. Baretta, M. Blaas, X. Desmit, W. Kühn,
 ⁷⁵⁶ G. Lacroix, et al., Predicting the consequences of nutrient reduction on
 ⁷⁵⁷ the eutrophication status of the north sea, Journal of Marine Systems
 ⁷⁵⁸ 81 (1-2) (2010) 148–170.
- [51] J. Blackford, An analysis of benthic biological dynamics in a north sea
 ecosystem model, Journal of Sea Research 38 (3-4) (1997) 213–230.
- [52] R. Geider, H. MacIntyre, T. Kana, Dynamic model of phytoplankton
 growth and acclimation: responses of the balanced growth rate and the
 chlorophyll a: carbon ratio to light, nutrient-limitation and temperature,
 Marine Ecology Progress Series 148 (1997) 187–200.
- J. Baretta-Bekker, J. Baretta, W. Ebenhöh, Microbial dynamics in the
 marine ecosystem model ersem ii with decoupled carbon assimilation
 and nutrient uptake, Journal of Sea Research 38 (3-4) (1997) 195–211.

- [54] Y. Artioli, J. C. Blackford, M. Butenschön, J. T. Holt, S. L. Wakelin,
 H. Thomas, A. V. Borges, J. I. Allen, The carbonate system in the north
 sea: Sensitivity and model validation, Journal of Marine Systems 102
 (2012) 1–13.
- [55] H. E. Garcia, R. A. Locarnini, T. P. Boyer, J. I. Antonov, O. K. Baranova, M. M. Zweng, J. R. Reagan, D. R. Johnson, A. V. Mishonov,
 S. Levitus, World ocean atlas 2013. volume 4, dissolved inorganic nutrients (phosphate, nitrate, silicate) (2013).
- [56] R. M. Key, A. Olsen, S. van Heuven, S. K. Lauvset, A. Velo, X. Lin,
 C. Schirnick, A. Kozyr, T. Tanhua, M. Hoppema, et al., Global ocean data analysis project, version 2 (glodapv2) (2015).
- [57] S. K. Lauvset, R. M. Key, A. Olsen, S. van Heuven, A. Velo, X. Lin,
 C. Schirnick, A. Kozyr, T. Tanhua, M. Hoppema, et al., A new global
 interior ocean mapped climatology: The 1× 1 glodap version 2, Earth
 System Science Data 8 (2016) 325–340.
- [58] Z.-P. Lee, K.-P. Du, R. Arnone, A model for the diffuse attenuation
 coefficient of downwelling irradiance, Journal of Geophysical Research:
 Oceans 110 (C2) (2005).
- [59] T. J. Smyth, Y. Artioli, Global inherent optical properties from SeaWiFS data (2010). doi:10.1594/PANGAEA.741913.
 URL https://doi.org/10.1594/PANGAEA.741913
- [60] S. A. Good, M. J. Martin, N. A. Rayner, En4: Quality controlled ocean temperature and salinity profiles and monthly objective analyses with uncertainty estimates, Journal of Geophysical Research: Oceans 118 (12) (2013) 6704–6716.
- [61] J. While, M. J. Martin, Variational bias correction of satellite sea-surface
 temperature data incorporating observations of the bias, Quarterly Journal of the Royal Meteorological Society 145 (723) (2019) 2733–2754.
- [62] S. Sathyendranath, R. J. Brewin, C. Brockmann, V. Brotas, B. Calton,
 A. Chuprin, P. Cipollini, A. B. Couto, J. Dingle, R. Doerffer, et al., An
 ocean-colour time series for use in climate studies: The experience of the
 ocean-colour climate change initiative (oc-cci), Sensors 19 (19) (2019)
 4285.

- [63] J. W. Campbell, The lognormal distribution as a model for bio-optical variability in the sea, Journal of Geophysical Research: Oceans 100 (C7) (1995) 13237–13254.
- [64] A. D. Beaton, C. L. Cardwell, R. S. Thomas, V. J. Sieben, F.-E. Legiret,
 E. M. Waugh, P. J. Statham, M. C. Mowlem, H. Morgan, Lab-on-chip
 measurement of nitrate and nitrite for in situ analysis of natural waters,
 Environmental science & technology 46 (17) (2012) 9548–9556.
- [65] A. G. Vincent, R. W. Pascal, A. D. Beaton, J. Walk, J. E. Hopkins,
 E. M. S. Woodward, M. Mowlem, M. C. Lohan, Nitrate drawdown during a shelf sea spring bloom revealed using a novel microfluidic in situ
 chemical sensor deployed within an autonomous underwater glider, Marine Chemistry 205 (2018) 29–36.
- [66] A. Birchill, G. Clinton-Bailey, R. Hanz, E. Mawji, T. Cariou, C. White,
 S. Ussher, P. Worsfold, E. P. Achterberg, M. Mowlem, Realistic measurement uncertainties for marine macronutrient measurements conducted
 using gas segmented flow and lab-on-chip techniques, Talanta 200 (2019)
 228–235.
- [67] I. Hinrichs, V. Gouretski, J. Pätch, K. Emeis, D. Stammer, North sea
 biogeochemical climatology (2017).
- [68] K. Mogensen, M. Balmaseda, A. Weaver, M. Martin, A. Vidard,
 Nemovar: A variational data assimilation system for the nemo ocean
 model, ECMWF newsletter 120 (2009) 17–22.
- [69] K. Mogensen, M. A. Balmaseda, A. Weaver, et al., The nemovar ocean
 data assimilation system as implemented in the ecmwf ocean analysis
 for system 4 (2012).
- [70] J. Waters, D. J. Lea, M. J. Martin, I. Mirouze, A. Weaver, J. While,
 Implementing a variational data assimilation system in an operational
 1/4 degree global ocean model, Quarterly Journal of the Royal Meteorological Society 141 (687) (2015) 333–349.
- [71] S. Kay, R. McEwan, D. Ford, North west european shelf production
 centre northwestshelf_analysis_forecast_bio_004_011, quality information
 document, Copernicus Marine Environment Monitoring Service (2019).

- [72] R. R. King, J. While, M. J. Martin, D. J. Lea, B. Lemieux-Dudon, J. Waters, E. O'Dea, Improving the initialisation of the met office operational shelf-seas model, Ocean Modelling 130 (2018) 1–14.
- [73] D. Ford, Assimilating synthetic biogeochemical-argo and ocean colour
 observations into a global ocean model to inform observing system design, Biogeosciences 18 (2) (2021) 509–534.
- [74] S. Bloom, L. Takacs, A. Da Silva, D. Ledvina, Data assimilation using
 incremental analysis updates, Monthly Weather Review 124 (6) (1996)
 1256–1271.
- [75] H. Sverdrup, On conditions for the vernal blooming of phytoplankton,
 J. Cons. Int. Explor. Mer 18 (3) (1953) 287–295.
- ⁸⁴⁴ [76] M. Goodliff, T. Bruening, F. Schwichtenberg, X. Li, A. Lindenthal,
 ⁸⁴⁵ I. Lorkowski, L. Nerger, Temperature assimilation into a coastal ocean⁸⁴⁶ biogeochemical model: assessment of weakly and strongly coupled data
 ⁸⁴⁷ assimilation, Ocean Dynamics 69 (10) (2019) 1217–1237.

Supporting Information for "Improved consistency between the modelling of ocean optics, biogeochemistry and physics, and its impact on the North-West European Shelf seas"

Jozef Skákala^{1,2}, Jorn Bruggeman¹, David Ford³, Sarah Wakelin⁴, Anıl Akpınar⁴, Tom Hull^{5,6}, Jan Kaiser⁶, Benjamin R. Loveday⁷, Charlotte A.J. Williams⁴and Stefano Ciavatta^{1,2}

¹Plymouth Marine Laboratory, The Hoe, Plymouth, PL1 3DH United Kingdom.

 $^2\mathrm{National}$ Centre for Earth Observation, Plymouth, PL1 3DH, UK.

³Met Office, FitzRoy Road, Exeter, EX1 3PB UK.

⁴National Oceanography Centre, Joseph Proudman Building, 6 Brownlow Street, Liverpool, L3 5DA UK.

⁵Centre for Environment, Fisheries and Aquaculture Science, Lowestoft, NR33 0HT UK.

⁶Centre for Ocean and Atmospheric Science, University of East Anglia, Norwich, NR4 7TJ, UK.

⁷Innoflair UG, Richard-Wagner-Weg 35, 64287, Darmstadt, Germany.

Contents of this file

1. Figures S1 to S10

Figures

The panels in the Fig.S1-S10 use the following abbreviations: "free 1-way": free run of the one-way coupled model, "free 2-way": free run of the two-way coupled model, "phys DA 1-way": physical data assimilation into the one-way coupled model, "phys DA 2-way": physical data assimilation into the two-way coupled model, "chl DA 1-way": chlorophyll assimilation into the one-way coupled model, "chl DA 2-way": chlorophyll assimilation

X - 2

into the two-way coupled model, "phys+chl DA 1-way": joint physical data - chlorophyll assimilation into the one-way coupled model, "phys+chl DA 2-way": joint physical data - chlorophyll assimilation into the two-way coupled model.



Figure S1. The locations of the 2018 in situ data used both for the assimilation and the validation. The panel A shows the locations of the AlterEco glider measurements and the bottom panel B shows the locations of the EN4 data for temperature and salinity. The EN4 data located outside of the NWE Shelf (bounded by the black line) were used only for assimilation, not for validation.

X - 4



Figure S2. The panel A shows Hovmöller diagram (time on the x-axis vs depth on the y-axis) for the temperature (C) of the one-way coupled free run ("free 1-way"), where the values for each day and depth represent the horizontal spatial averages throughout the NWE Shelf (bathymetry < 200m). Panels B-F show the same Hovmöller diagrams, but for the differences between the two-way coupled, or assimilative runs and the reference, free one-way coupled model run. The purpose of the panels B-F is to provide an understanding of how the bio-optical module and the assimilative model components influence the temperature of the reference free one-way coupled run. The yellow lines in the panels D-F show the MLD of the physical data assimilative runs to indicate the vertical scale of impact of the SST assimilation.



Figure S3. The 2018 time-series of SST averaged throughout the NWE Shelf compared between the different one-way, two-way coupled, free, or assimilative simulations and the VIIRS/in situ data. Panel A compares the different one-way coupled runs, i.e. the one-way coupled free run with the physical data assimilative run, panel B compares the different two-way coupled runs, i.e. the two-way coupled free run with the physical data assimilative run, the chlorophyll assimilative run and the run assimilating both physical data and chlorophyll. To consistently compare the model simulations with the VIIRS/in situ SST, the model outputs were masked wherever there were missing satellite data. The missing satellite data are due to the movements of clouds and atmospheric disturbances and the missing values are responsible for the small time-scale fluctuations in the different curves shown in the three panels. We do not show the one-way coupled runs assimilating chlorophyll, as those have by definition no impact on the simulated temperature.



Figure S4. The seasonal differences in temperature (x-axis, $^{\circ}$ C) between the two-way coupled, or assimilative runs and the reference, one-way coupled free run. The differences are shown as a function of depth (y-axis, m), and averaged throughout the seasonal period and the NWE Shelf.



Figure S5. The 2018 time-series of surface chlorophyll *a* concentrations (mg/m^3) averaged throughout the NWE Shelf compared between the different one-way, two-way coupled, free, or assimilative simulations and the satellite data, as well as with the NSBC climatological dataset. Panel A compares the different one-way coupled runs, i.e. the one-way coupled free run with the physical data assimilative run, the chlorophyll assimilative run and the joint physical data-chlorophyll assimilative run, panel B compares the different two-way coupled runs, i.e. the two-way coupled free run with the physical data assimilative run, the chlorophyll assimilative run and the joint physical data-chlorophyll assimilative run. The chlorophyll assimilative run from both panels A and B is hard to see, as the line is nearly identical with the joint physicalchlorophyll assimilative run. The satellite data were considered only in the March-September period as the data outside this period are scarce and limited only to the southern part of the NWE domain. The small time-scale fluctuations in the satellite data are due to the missing values caused by the movement of clouds and atmospheric disturbances.



Figure S6. Panel A shows the mixed layer depth (MLD, in m) of the one-way coupled free run (the reference run). The MLD values are averaged for the spring bloom period between March-May 2018. The panels B-F show the relative changes (relative to the one-way coupled free reference run, in m) in MLD carried by the two-way coupled free run (panel B), chlorophyll assimilation into the two-way coupled model (panel C), physical data assimilation into the oneway coupled (panel D) and into the two-way coupled model (panel E) and the joint physical data-chlorophyll assimilation into the two-way coupled model (panel F). All panels B-F show the difference between the MLD of the specific two-way coupled, or assimilative simulation and the one-way coupled free run (panel A). The black line shows the boundary of the continental shelf (bathymetry < 200m).



Figure S7. The model to VIIRS/in situ SST differences in °C. The differences are shown for the: free two-way coupled model (panel A), physical data assimilation into the two-way coupled model (panel B), chlorophyll assimilation into the two-way coupled model (panel C), and joint physical data-chlorophyll assimilation into the two-way coupled model (panel D).

April 14, 2021, 9:15am



Figure S8. Hovmöller diagram for chlorophyll concentrations (mg/m^3) measured by the Cabot glider in the central North Sea during an early May to mid-August 2018 mission. The right-hand panels (B,D,F,H) show the chlorophyll differences between the free one-way coupled model run (panel B), free two-way coupled model run (panel D), the physical data assimilation into the one-way coupled model (panel F), the physical data assimilation into the two-way coupled model (panel H), and the Cabot glider observations (model minus glider). The left hand panels show the differences between the observations, or model simulations and the reference, free one-way coupled model run. The purpose of the left-hand panels is to show the desired changes to the one-way coupled model (panel A) and how these changes are realized by the biogeochemical feedback in the free run (panel C) and in the physical data-assimilative runs (panels E and G).

April 14, 2021, 9:15am



Figure S9. Impact of two-way coupling and assimilation on the simulated chlorophyll concentrations (mg/m^3) . The panel A shows Hovmöller diagram (time on the x-axis vs depth on the y-axis) for the one-way coupled model free run, where the values for each day and depth represent the horizontal spatial averages throughout the NWE Shelf (bathymetry < 200m). Panels B-H show the same Hovmöller diagrams, but for the differences between the two-way coupled, or assimilative runs and the reference, free one-way coupled run. The yellow lines in the panels E-H show the mixed layer depth, providing the boundary of the region in which the ocean color assimilation directly updates the simulated chlorophyll.



Figure S10. The 2018 mean surface chlorophyll concentrations (in mg/m^3). The different panels compare: the one-way coupled model free run (panel A), the two-way coupled model free run (panel B), the physical data assimilation into the one-way coupled model (panel C), the physical data assimilation into the two-way coupled model (panel D), the chlorophyll assimilation into the one-way coupled model (panel E), the chlorophyll assimilation into the two-way coupled model (panel F), the joint physical data-chlorophyll assimilation into the one-way coupled model (panel G), the joint physical data-chlorophyll assimilation into the two-way coupled model H), and the assimilated satellite ocean color observations (panel I). The black line shows the continental shelf boundary (bathymetry < 200m).

April 14, 2021, 9:15am