# Extensive remineralization of peatland-derived dissolved organic carbon and acidification in the Sunda Shelf Sea, Southeast Asia

YONGLI ZHOU<sup>1</sup>, Christopher Evans<sup>2</sup>, Yuan Chen<sup>1</sup>, Kristy Chang<sup>3</sup>, and Patrick Martin<sup>1</sup>

November 26, 2022

#### Abstract

Southeast Asia is a hotspot of riverine export of terrigenous organic carbon to the ocean, accounting for  $^{\sim}10\%$  of the global land-to-ocean riverine flux of terrigenous dissolved organic carbon (tDOC). While anthropogenic disturbance is thought to have increased the tDOC loss from peatlands in Southeast Asia, the fate of this tDOC in the marine environment and the potential impacts of its remineralization on coastal ecosystems remain poorly understood. We collected a multi-year biogeochemical time series in the central Sunda Shelf (Singapore Strait), where the seasonal reversal of ocean currents delivers water masses from the South China Sea first before (during Northeast Monsoon) and then after (during Southwest Monsoon) they have mixed with run-off from peatlands on Sumatra. The concentration and stable isotope composition of dissolved organic carbon, and colored dissolved organic matter spectra, reveal a large input of tDOC to our site during Southwest Monsoon. Using isotope mass balance calculations, we show that 60-70% of the original tDOC input is remineralized in the coastal waters of the Sunda Shelf, causing seasonal acidification by up to 0.10 pH units. The persistent CO2 oversaturation drives a CO2 efflux of 4.1-8.2 mol C m-2 yr-1 from the Singapore Strait, suggesting that a large proportion of the remineralized peatland tDOC is ultimately emitted to the atmosphere. However, incubation experiments show that the remaining 30-40% tDOC exhibits surprisingly low lability to microbial and photochemical degradation, suggesting that up to 20-30% of peatland tDOC might be relatively refractory and exported to the open ocean.

<sup>&</sup>lt;sup>1</sup>Nanyang Technological University, Singapore

<sup>&</sup>lt;sup>2</sup>UK Centre for Ecology and Hydrology

<sup>&</sup>lt;sup>3</sup>Nanyang Technological University

Extensive remineralization of peatland-derived dissolved organic carbon and acidification in 1 2 the Sunda Shelf Sea, Southeast Asia 3 Yongli Zhou<sup>1,\*</sup>, Christopher D. Evans<sup>2</sup>, Yuan Chen<sup>1</sup>, Kristy Y.W. Chang<sup>1,†</sup>, Patrick Martin<sup>1,\*</sup> 4 5 <sup>1</sup> Asian School of the Environment, Nanyang Technological University, Singapore 6 <sup>2</sup> UK Centre for Ecology and Hydrology, Bangor, UK 7 8 † Current address: Singapore Centre for Environmental Life Sciences Engineering, Nanyang 9 Technological University, Singapore \* Correspondence to zhou0303@e.ntu.edu.sg or pmartin@ntu.edu.sg 10 11 12 **ORCID** list 13 Yongli Zhou: 0000-0001-8403-6315 14 15 Christopher D. Evans: 0000-0002-7052-354X Yuan Chen: 0000-0002-9625-0930 16 Kristy Y.W. Chang: 0000-0001-6387-2744 17 Patrick Martin: 0000-0001-8008-5558 18 19 Key words: terrigenous organic carbon, remineralization, acidification, Sunda Shelf Sea, 20 21 tropical peatlands

$\sim$	$\sim$
,	,
	_

## **Key Points:**

- 60–70% of the dissolved organic carbon exported from the tropical peatlands is remineralized in the coastal waters of the Sunda Shelf
- This drives a seasonal acidification by 0.10 pH units and a net CO<sub>2</sub> efflux of 4.1–8.2 mol C m<sup>-2</sup> yr<sup>-1</sup> in the central Sunda Shelf
- 20–30% of the peatland-derived dissolved organic carbon might be relatively
   refractory and exported to the open Indian Ocean

#### **Abstract**

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

Southeast Asia is a hotspot of riverine export of terrigenous organic carbon to the ocean, accounting for ~10% of the global land-to-ocean riverine flux of terrigenous dissolved organic carbon (tDOC). While anthropogenic disturbance is thought to have increased the tDOC loss from peatlands in Southeast Asia, the fate of this tDOC in the marine environment and the potential impacts of its remineralization on coastal ecosystems remain poorly understood. We collected a multi-year biogeochemical time series in the central Sunda Shelf (Singapore Strait), where the seasonal reversal of ocean currents delivers water masses from the South China Sea first before (during Northeast Monsoon) and then after (during Southwest Monsoon) they have mixed with run-off from peatlands on Sumatra. The concentration and stable isotope composition of dissolved organic carbon, and colored dissolved organic matter spectra, reveal a large input of tDOC to our site during Southwest Monsoon. Using isotope mass balance calculations, we show that 60–70% of the original tDOC input is remineralized in the coastal waters of the Sunda Shelf, causing seasonal acidification by up to 0.10 pH units. The persistent CO<sub>2</sub> oversaturation drives a CO<sub>2</sub> efflux of 4.1 – 8.2 mol C m<sup>-2</sup> yr<sup>-1</sup> from the Singapore Strait, suggesting that a large proportion of the remineralized peatland tDOC is ultimately emitted to the atmosphere. However, incubation experiments show that the remaining 30-40% tDOC exhibits surprisingly low lability to microbial and photochemical degradation, suggesting that up to 20–30% of peatland tDOC might be relatively refractory and exported to the open ocean.

#### Plain Language Summary

Peatlands are large stores of organic carbon and are extensively distributed across coastal Sumatra and Borneo in Southeast Asia. These peatlands give rise to a large flux of dissolved organic carbon via rivers to the Sunda Shelf Sea, the marginal sea in Southeast Asia, and land conversion of peatlands has likely increased this flux. In the marine environment, organic carbon can be remineralized to CO<sub>2</sub> by sunlight and by microbial respiration, but the fate of peatland tDOC in coastal waters of Southeast Asia remains unclear. In this study, we analyzed water samples collected from the Sunda Shelf and found that 60–70% of the dissolved organic carbon from peatlands is decomposed to CO<sub>2</sub> in the coastal waters of the shelf sea. We further found that this resulted in a decrease in seawater pH by 0.10 units, and that a large proportion of the CO<sub>2</sub> is probably emitted to the atmosphere. This implies that the reported increase in tDOC export due to peatland disturbance may have had a significant effect on coastal biogeochemistry and downstream greenhouse gas emissions.

#### 1 Introduction

Southeast Asia harbors the majority of the world's tropical peatlands, which are widely distributed in coastal areas of Sumatra and Borneo and store around 69 Pg C of terrestrial organic carbon (Dommain et al., 2014; Page et al., 2011). Consequently, Southeast Asia is also a hotspot of terrestrial organic carbon export to the ocean: the fluvial flux of terrigenous dissolved organic carbon (tDOC) to the Sunda Shelf Sea is ~21 Tg C yr<sup>-1</sup>, which accounts for ~10% of the annual global land-to-ocean tDOC flux by world's rivers (Baum et al., 2007; Moore et al., 2011). In addition, most of this peatland area has been anthropogenically disturbed by deforestation and land conversion (Miettinen et al., 2016), which is thought to have increased the peatland tDOC export to the ocean by 32% over the past three decades (Moore et al., 2013; Yupi et al., 2016). This ongoing anthropogenic pressure on peatlands could therefore have impacts extending into the marine environment of Southeast Asia, especially on the sensitive ecosystems such as coral reefs, which critically hinge on the biogeochemical processing of the peatland-derived tDOC in the shelf sea.

Our understanding of the biogeochemical fate of tDOC in the global oceans is still limited, and in particular the degree to which ocean margins are efficient filters or conduits of tDOC between land and ocean remains debated. On the one hand, numerous studies have indicated that tDOC can be extensively and rapidly remineralized within ocean margins, as reported for the North Sea (Kitidis et al., 2019; Painter et al., 2018), the Louisiana Shelf (Fichot & Benner, 2014) and the Arctic Shelf (Alling et al., 2008; Humborg et al., 2017; Kaiser et al., 2017;

89

90

91

92

93

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

Letscher et al., 2011; Semiletov et al., 2016). Remineralization of tDOC fuels strong CO<sub>2</sub> emissions from coastal waters (Cai, 2011; Chen & Borges, 2009; Roobaert et al., 2019) and causes acidification in shelf seas (Semiletov et al., 2016). This is consistent with the canonical view that the DOC pool in the deep oceans only contains trace amounts of terrigenous DOC, as indicated by the low lignin concentrations (Meyers-Schulte & Hedges, 1986; Opsahl & Benner, 1997). On the other hand, some studies suggest that tDOC sometimes undergoes more limited remineralization within the ocean margins, and that a correspondingly larger proportion is delivered to the open oceans. For example, Medeiros et al. (2015) found that 50–76% of the tDOC exported by the Amazon River reaches the open Atlantic Ocean, likely due to the rapid transport of tDOC across the shelf. This might also be linked to the prior degradation history of tDOC before reaching the sea, as removal of the most labile fractions may result in the remaining tDOC having only limited degradability in some river systems (Chupakova et al., 2018; Shirokova et al., 2019; Stubbins et al., 2017). Recent molecular evidence shows that some fractions of tDOC are refractory and are exported to the open ocean (Cao et al., 2018; Medeiros et al., 2016), and carbon isotope data suggest that the terrestrial contribution to the oceanic DOC pool might be up to 30%, which is higher than previously believed (Follett et al., 2014; Zigah et al., 2017). It therefore remains unclear exactly how much tDOC is remineralized within ocean margins globally, with the extent of tDOC remineralization likely depending on factors such as the composition of the tDOC and the water circulation across the shelf, and varying substantially between different regions. However, most research has hitherto focused on mid- to high-latitude regions, and more research is therefore needed in tropical shelf seas that are the recipients of some of the highest global rates of tDOC input.

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

129

109

The peatland-draining rivers in Southeast Asia are characterized by very high DOC concentrations (1000–5000 µmol L<sup>-1</sup>) and high colored dissolved organic matter (CDOM, 50– 200 m<sup>-1</sup>) (Alkhatib et al., 2007; Martin et al., 2018; Moore et al., 2011; Rixen et al., 2008; Wit et al., 2015, 2018), which are among the highest values found in rivers globally (Meybeck, 1982). However, these peat-draining rivers only show moderate-to-low CO<sub>2</sub> efflux relative to their high DOC concentrations (Müller-Dum et al., 2019; Müller et al., 2015; Wit et al., 2015). In addition, field surveys have typically revealed conservative mixing of DOC across peatlanddraining river estuaries (Alkhatib et al., 2007; Martin et al., 2018; Rixen et al., 2008; Wit et al., 2015; Zhou et al., 2019). This suggests that remineralization of tDOC within the rivers is limited, possibly due to lack of oxygen, low pH and short water residence times (Wit et al., 2015), which means that a large proportion of the peatland-derived tDOC is presumably exported to the coastal sea. Consistent with this prediction, Zhou et al., (2019) estimated from fluorescent dissolved organic matter data that 20–40% of the bulk DOC pool in coastal waters of northwestern Borneo (with salinity >30) is of terrestrial origin, due to the large input of peatland tDOC. It is possible that a large proportion of this tDOC is remineralized after it is exported to the coastal waters, as suggested by the high photo-lability of Southeast Asian peatland tDOC (Martin et al., 2018; Rixen et al., 2008) and the large CO<sub>2</sub> emissions reported for the coastal waters of eastern Sumatra (Wit et al., 2018). Given the large scale of the potential anthropogenic perturbation to the peatland tDOC flux, it is particularly important to better

constrain the biogeochemical fate of this tDOC and the associated impacts on the shelf sea environment. Here, we analyzed a multi-year time series of dissolved organic carbon, carbonate system parameters, and carbon stable isotope composition using a mass balance approach to quantify the remineralization of peatland-derived tDOC in the Sunda Shelf. The seasonal acidification in the shelf sea and CO<sub>2</sub> emissions driven by the tDOC remineralization were further quantified. Laboratory decomposition experiments were performed to investigate the degradability of peatland-derived tDOC and estimate tDOC export to the open ocean. 

#### 2 Materials and methods

### 2.1 Study area

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

166

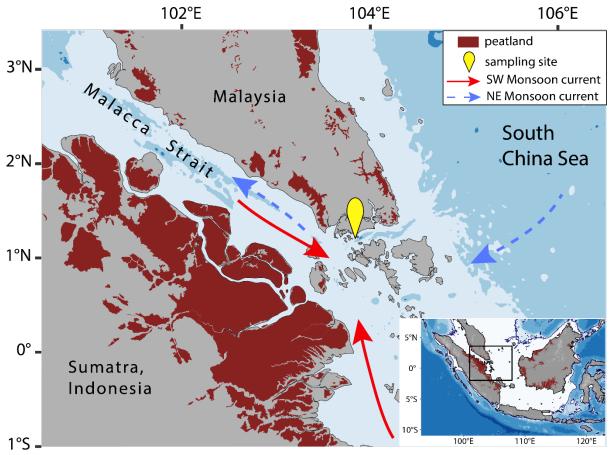
167

168

We collected biogeochemical time-series data at two sites in the Singapore Strait, Kusu Island (1.226°N, 103.860°E) and Hantu Island (1.227°N, 103.747°E). Both sites have coral reefs with >70 species of hard coral, and coral cover of around 40–50% (Bauman et al., 2017; Huang et al., 2009; Januchowski-Hartley et al., 2020). The Singapore Strait is located in the center of the Sunda Shelf Sea, and is close to the coastal peatlands on Sumatra (Fig. 1), with water depth mostly less than 40 m. The annual average circulation runs from the South China Sea into the Indian Ocean through the Java Sea and the Malacca Strait (Gordon et al., 2012). However, the monsoon system causes the main ocean currents to reverse direction seasonally (Fig. 1) (Mayer et al., 2018; Mayer & Pohlmann, 2014). During the Northeast Monsoon (November to February), water flows from the South China Sea westwards through the Singapore Strait and along the coast of Sumatra. During the Southwest Monsoon (May to September), the currents reverse and flow back eastwards through the Singapore Strait, carrying river discharge from the Sumatran peatlands (Mayer & Pohlmann, 2014; Siegel et al., 2019). Water in the Malacca and Singapore Straits is well mixed due to strong tidal currents (Mayer & Pohlmann, 2014). Because spatial variability between our two sampling sites was much smaller than the seasonal variability, we do not distinguish the data by site in this analysis.

169

170



**Figure 1.** Map of the study region showing the location of tropical peatlands, the time series sampling site in the Singapore Strait and the directions of the prevailing ocean currents across the Sunda Shelf Sea. During the Northeast (NE) Monsoon season, the currents transport water from the open South China Sea to our study site; during the Southwest (SW) Monsoon season, the currents reverse and deliver water with inputs from the tropical peatlands along the east coast of Sumatra to our site, resulting in the seasonal variations in biogeochemistry.

#### 2.2 Collection of water samples

Water samples were collected from both stations 1–2 times per month using a 1.5-L Niskin bottle at 5-m depth from October 2017 until August 2020. The water was immediately filtered on the boat through a pre-rinsed 47 mm diameter, 0.22 µm pore-size polyethersulfone membrane filter (Supor, Merck Millipore) in an in-line filter housing connected to a peristaltic pump and preserved for analyses of dissolved organic carbon (DOC), colored dissolved organic matter (CDOM) spectra, dissolved inorganic carbon (DIC), total alkalinity (TA), stable isotope

composition of DOC ( $\delta^{13}C_{DOC}$ ) and of DIC ( $\delta^{13}C_{DIC}$ ) (methods of collection are summarized in Table 1). The filter housing was bled of any air bubbles before collecting samples. Samples for chlorophyll-a were filtered onto 25 mm diameter Whatman GF/F filters instead, kept at  $-80^{\circ}C$  and analyzed within 3 months of collection. At each station, a depth profile of salinity and temperature was measured by a fastCTD Profiler (Valeport Ltd).

Table 1. Collection and preservation of each biogeochemical parameter after filtration.

Parameter	Collection method	Preservation	
		temperature	
$\delta^{13}\mathrm{C}_\mathrm{DIC}$	1 ml of sample injected into a 12-ml Exetainer vial	20°C	
	capped with butyl rubber septum containing 1 ml of		
	≥85% phosphoric acid and pre-flushed with helium.		
DIC	A 12-ml Exetainer vial was filled to overflowing,	4°C	
	left to overflow for 15s, and capped immediately		
	with a butyl rubber septum to prevent gas exchange.		
TA	Collected into a 140-ml HDPE bottle.	4°C	
$\delta^{13} C_{DOC}$	Filled 40 ml into a 50-ml polypropylene centrifuge tube and stored frozen. Samples were thawed and acidified with 37% hydrochloric acid immediately prior to being sent off for analysis.	-20°C	
CDOM	Filled 30 ml into a 40-ml pre-combusted EPA borosilicate vial.	4°C	
DOC	Filled 30 ml into a 40-ml EPA borosilicate vial, acidified with 100 µl of 50% sulfuric acid.	4°C	

### 2.3 Deployment of autonomous sensors

An SBE 19plus V2 SeaCAT CTD and a SeaFET pH sensor, equipped with copper biofouling

guards, (both Sea-Bird Electronics, Inc.) were deployed at Kusu Island in July 2015 to monitor seawater salinity, temperature, and pH every 10 minutes. A SAMI-CO<sub>2</sub> sensor (Sunburst Sensors), equipped with copper biofouling guards, was deployed at the same location from March 2018 until July 2019 to measure the partial pressure of CO<sub>2</sub> (pCO<sub>2</sub>) every two hours. Sensors were attached horizontally to iron stakes hammered into the reef sediment at roughly 5 m depth along the reef slope and were retrieved every 1–3 months for data download and maintenance.

The sensor data were validated with the CTD profile measurements and our measurements of dissolved inorganic carbon and total alkalinity of water samples collected during each field cruise, using the MATLAB CO2SYS Package Version 2.0 (referred to as CO2SYS below) (van Heuven et al., 2011). The sensor data are presented as daily mean values. Data gaps occurred whenever sensors were removed for maintenance or were sent for recalibration. The data quality of the SAMI-CO<sub>2</sub> sensor was rapidly affected by biofouling and sediment accumulation despite testing different sample pump and copper fouling guard configurations, and we therefore stopped deploying it after July 2019.

### 2.4 Sample analysis

Dissolved inorganic carbon (DIC) was measured with an Apollo SciTech AS-C5 DIC analyzer at room temperature (22±0.5°C) using an injection volume of 1 ml and a phosphoric acid solution of 3% v/v phosphoric acid with 7% w/v sodium chloride. The phosphoric acid was

220

221

222

223

224

225

226

227

228

229

230

231

232

233

234

235

236

237

238

239

sparged with N<sub>2</sub> gas for 5 min prior to each analysis run. Each sample was measured 3–5 times to achieve a relative standard deviation of  $\leq 0.1\%$  for three replicate injections. Calibration was performed using the certified reference material (CRM) from Andrew Dickson's laboratory, Scripps Institution of Oceanography (Batch 172) or an in-house secondary standard made from Singapore Strait seawater that we calibrated against the CRM. The analytical precision for DIC was  $\pm 0.15\%$  or lower based on replicate measurements of the CRM and the secondary standard. Total alkalinity (TA) was measured with an Apollo SciTech AS-ALK2 titrator, which automates the Gran titration (Gran, 1952), using a ROSS combination glass pH electrode (Orion 8302BNUMD). Samples and standards were warmed to room temperature (22±0.5°C) and samples were measured 2-4 times in 25-ml aliquots using 0.1 M hydrochloric acid. The acid concentration was calibrated against the CRM or our in-house secondary standard. The analytical precision for TA was ±0.13% or lower based on replicate measurements of the CRM and the secondary standard. Absorption spectra of CDOM were measured from 230 nm to 900 nm against a reference of ultrapure deionized water (18.2 MΩ cm<sup>-1</sup>, referred to as DI water below) using a Thermo Evolution 300 dual-beam spectrophotometer in a 10-cm quartz cuvette. The absorption coefficients, the spectral slope S<sub>275-295</sub> (Helms et al., 2008), and the specific UV absorbance at 254 nm, SUVA<sub>254</sub> (Weishaar et al., 2003), were calculated using the "hyperSpec" package in R (Beleites & Sergo, 2012). We express the CDOM concentration as the absorption coefficient at

240 350 nm, *a*<sub>350</sub>.

DOC samples (30 ml) were acidified with 100  $\mu$ l of 50% sulfuric acid and analyzed on a Shimadzu TOC-L system with a high-salt combustion kit, using an injection volume of 150  $\mu$ l and a 5-minute sparge time. Each sample was measured 5–7 times to achieve a coefficient of variation of  $\leq$ 2%. Potassium hydrogen phthalate was used for calibration, and the analytical accuracy was monitored using certified deep-sea water reference material (42–45  $\mu$ mol L<sup>-1</sup> DOC) from the University of Miami, USA. Our analyses consistently yielded slightly higher values for the CRM, with a long-term mean and standard deviation of 48.0±3.9  $\mu$ mol L<sup>-1</sup> DOC.

The  $\delta^{13}C_{DOC}$  samples were thawed, acidified to pH of 2–3 with 37% w/v hydrochloric acid, and shipped to the Ján Veizer Stable Isotope Laboratory, University of Ottawa, Canada for analysis using an OI Analytical Aurora Model 1030W TOC Analyzer interfaced to a Finnigan Mat DeltaPlusXP isotope ratio mass spectrometer. The 2-sigma analytical precision is  $\pm 0.4\%$  (https://isotope.uottawa.ca/en/services-waters).

 $\delta^{13}$ C of DIC measurements were made using a Gas Bench connected to an isotope ratio mass spectrometer at the Stable Isotope Facility, University of California, Davis (UC Davis) and by ourselves at the Marine Geochemistry Laboratory, Nanyang Technological University, Singapore (NTU). At NTU, final  $\delta^{13}$ C values were obtained after instrumental drift correction, blank correction for standards (Assayag et al., 2006; Humphreys et al., 2016), and calibration

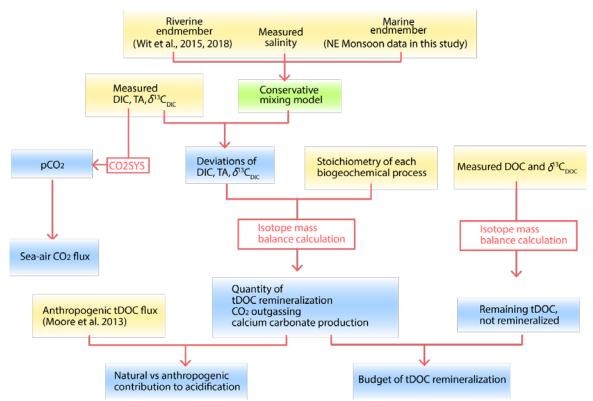
using NBS-18 ( $\delta^{13}$ C = -5.01%) and Estremoz ( $\delta^{13}$ C = 1.63‰) calcium carbonate standards. All  $\delta^{13}$ C values of the samples were further corrected by accounting for the difference between the international consensus value of  $\delta^{13}$ C<sub>DIC</sub> of the Dickson CRM (0.78‰) (Cheng et al., 2019) and our laboratory measurements of  $\delta^{13}$ C<sub>DIC</sub> of the Dickson CRM at NTU (the magnitude of this correction was mostly below 0.4‰). The long-term standard deviation for  $\delta^{13}$ C of DIC measurements at UC Davis is reported as  $\pm 0.1\%$  (https://stableisotopefacility.ucdavis.edu/) and at NTU was  $\pm 0.2\%$  (Carrara marble, n=24, 4 runs).

Chlorophyll-*a* was extracted in 90% acetone at 4°C in the dark for 24 hours and measured on a HORIBA Floromax-4 fluorometer with excitation at 436 nm and emission at 680 nm, with bandpasses of 5 nm (Welschmeyer, 1994). Data were normalized to the reference signal and calibrated using a spinach chlorophyll-*a* standard (Sigma-Aldrich C5753).

#### 2.5 Analyses of the time-series data

Due to the monsoonal current reversal, the Singapore Strait seasonally receives water directly from the South China Sea with little terrestrial influence. This water subsequently mixes with peat-draining river input off the coast of Sumatra, and then seasonally flows back into the Singapore Strait when the currents reverse direction (Fig. 1). The seasonal variation in the Singapore Strait is therefore equivalent to the spatial variability from the South China Sea into the coastal waters of Sumatra in the Malacca and Karimata Straits, and therefore reflects the mixing of water from the open sea with peatland-draining river input.

Our data analysis consists of two components: first, we built a conservative mixing model to predict the values of DIC, TA, pH and  $\delta^{13}C_{DIC}$  if this mixing was strictly conservative. We therefore interpret any deviations in our measurements relative to the predicted values as indicative of biogeochemical processing. Second, we performed isotope mass balance calculations for DOC and the carbonate system to estimate how much of the DOC pool was tDOC, how much of the excess DIC (relative to conservative mixing) was contributed from tDOC remineralization, and, when the measured deviations in  $\delta^{13}C_{DIC}$  were greater than could be explained from the measured deviation in DIC concentration, how much additional tDOC must have been remineralized and already outgassed upstream of our site to explain the measured  $\delta^{13}C_{DIC}$ . These calculations are summarized in Fig. 2 and below, and full details are provided in the Supplementary Information.



**Figure 2.** A flowchart summarizing the data analyses for this study. Yellow boxes indicate the inputs and blue boxes indicate outputs of calculations. We built a conservative mixing model to predict the biogeochemical parameters if there was only mixing between riverine water and marine water. Isotope mass balance calculations were performed with the deviations of the measured data from the mixing model to quantify tDOC remineralization and the potential anthropogenic contribution to the seasonal acidification.

#### 2.5.1 Conservative mixing model

We built a two-endmember mixing model to calculate the DIC, TA, pH and  $\delta^{13}C_{DIC}$  expected from conservative mixing between the riverine endmember and the marine endmember. This allowed us to calculate the deviations of the measured data from the values expected from conservative mixing. The relative contribution of riverine input and marine input was calculated from the measured salinity. Seawater pH expected from conservative mixing was calculated from the DIC and TA of conservative mixing by CO2SYS.

We calculated the discharge-weighted average of the published data of DIC, TA (calculated

from pH and pCO<sub>2</sub>) and  $\delta^{13}C_{DIC}$  from the main peat-draining rivers on Sumatra (the Batanghari, Indragiri, Kampar and Siak Rivers) (Wit et al., 2015, 2018), which we adopted as the riverine endmember values. The marine endmember values were estimated from our measurements in the Singapore Strait from the end of the NE Monsoon and the subsequent inter-monsoon season (late February to March), when the water in the Singapore Strait consists predominantly of marine water from the open South China Sea and our data indicated minimal presence of tDOC and freshwater input (Section 3.1). All the endmember values and the uncertainties are provided in Table 2.

#### 

#### 2.5.2 Isotope mass balance calculations

Based on the known changes in DIC, TA and  $\delta^{13}C_{DIC}$  relative to the values expected from conservative mixing caused by primary production, remineralization, calcium carbonate production and dissolution and air-sea CO<sub>2</sub> exchange (Zeebe & Wolf-Gladrow, 2001) (Table S1), we back-calculated the amount of carbon that had been processed by these biogeochemical processes from the measured deviations in DIC, TA and  $\delta^{13}C_{DIC}$  using an isotope mass balance approach following Samanta et al. (2015). The proportion of the bulk DOC pool that was tDOC was calculated from the measured  $\delta^{13}C_{DOC}$  and DOC concentration.

The  $\delta^{13}C_{DOC}$  data were not available for rivers on Sumatra, but the  $\delta^{13}C_{DOC}$  values of the peatland-derived tDOC across peatlands on Sumatra and rivers on Borneo and Peninsular

Malaysia mostly range between -30% and -28% (Baum, 2008; Evans et al., 2014; Gandois et al., 2014; Zhu et al., 2020). We therefore adopted -29% as the riverine endmember value of  $\delta^{13}C_{DOC}$  for the peatland-derived tDOC. For the  $\delta^{13}C$  of the DIC produced from remineralization, microbial respiration causes little carbon isotopic fractionation, while results from our experiments (Section 3.6) and previous studies (Opsahl & Zepp, 2001; Osburn et al., 2001; Spencer et al., 2009) indicated that photodegradation (and combined photo-biodegradation) of tDOC can cause fractionation in  $\delta^{13}$ C of -1.4% to -5.8% between the produced DIC and the initial DOC (Table S2). Because of the low bio-degradability (Nichols & Martin, submitted) but high photo-degradability of Southeast Asian peatland tDOC (Section 3.6), we infer that a major part of the remineralization of tDOC in this region might be via photodegradation, and therefore entails a carbon isotopic fractionation. We adopted a fractionation of -3%, and thus used -32% as the  $\delta^{13}$ C of the DIC produced from tDOC remineralization. Note that by assuming this additional fractionation, our calculated amount of remineralized tDOC is smaller, since a smaller amount of carbon is needed to yield the observed  $\delta^{13}C_{DIC}$  values.

349

350

351

352

353

354

334

335

336

337

338

339

340

341

342

343

344

345

346

347

348

These mass balance calculations provide the quantity of (i) the amount of tDOC at our site, representing the remaining fraction of the tDOC input from peatlands that has not been remineralized, (ii) the remineralized tDOC as DIC, (iii) the remineralized tDOC that had been removed by outgassing from the DIC pool upstream of our sampling site, and (iv) the amount of calcium carbonate production and dissolution. The sum of the fractions (i) – (ii) represents

the initial concentration of tDOC for a given salinity after estuarine mixing and before biogeochemical processing has occurred.

The uncertainty of the mass balance calculations was estimated by a Monte Carlo algorithm, in which we perturbed each input parameter with a  $1\sigma$  normally distributed error (Table 2) by random sampling, repeated the calculations 10,000 times, and then calculated the resulting standard deviation of each output parameter. For the riverine endmember values as input parameters, the  $1\sigma$  error of the DIC and TA were adopted from Wit et al. (2018); the  $1\sigma$  error of the  $\delta^{13}C_{DIC}$  and  $\delta^{13}C_{DOC}$  were both assumed to be  $\pm 1\%$ , because the uncertainties are not available and the reported  $\delta^{13}C_{DOC}$  values of the peatland-derived tDOC in Southeast Asia mostly range between -30% and -28%. The  $1\sigma$  error of the marine endmember variables are the standard deviations of the measurements from late February to March. The  $1\sigma$  error of the measured DOC, DIC, TA,  $\delta^{13}C_{DIC}$  and  $\delta^{13}C_{DOC}$  are the long-term  $1\sigma$  precision of measurements of the standard materials. The  $1\sigma$  error of the measured salinity ( $\pm 0.01$ ) was estimated based on the accuracy of the conductivity measurement of the Valeport fastCTD ( $\pm 0.01$  mS cm<sup>-1</sup>).

#### 2.5.3 Acidification and putative anthropogenic contribution

Seasonal acidification was indicated by the time series measurements of DIC and TA and the SeaFET pH sensor data (Section 3.5). Saturation states of calcite and aragonite were calculated from the DIC and TA data by CO2SYS.

From the results of the mass balance calculations, we further reconstructed the individual contribution to the deviations of DIC and TA from (i) calcium carbonate dissolution, (ii) remineralization of natural tDOC, and (iii) remineralization of anthropogenic tDOC. The reconstructed DIC and TA deviations were translated to the pH deviations relative to conservative mixing by CO2SYS for assessment of the putative anthropogenic contribution to the observed acidification. We distinguished the natural tDOC remineralization from the anthropogenic component based on the report that 35% of the tDOC flux is caused by peatland perturbations (Moore et al., 2013). While this estimate is based on a limited and small-scale comparison of disturbed and intact peatland catchments that has yet to be demonstrated at large scales, it has already been applied by several other studies to gain insight into the possible downstream impacts of peatland disturbance (Wit et al., 2018).

### 2.5.4 Sea-air CO<sub>2</sub> flux

We calculated the annual areal sea-air CO<sub>2</sub> flux following Lønborg et al. (2019) using the pCO<sub>2</sub> calculated from the measured DIC and TA (the *in-situ* sensor record was too limited to be used). We obtained average wind speeds for the area of 0.5°N–1.5°N and 102.5°E – 105.5°E from the CYGNSS Level 2 Science Data Record Version 2.1 (CYGNSS, 2017). The gas transfer velocity was calculated following Wanninkhof et al. (2009). The CO<sub>2</sub> solubility was calculated following Weiss (1974) using the measured water temperature. The uncertainty of the CO<sub>2</sub> flux was estimated by a Monte Carlo algorithm, based on the uncertainty of the salinity, DIC and

TA measurements (Table 2) and the reported uncertainty of the wind speed measured by CYGNSS ( $\pm 1.4$  m/s) (Ruf et al., 2019).

### 2.6 Degradation experiments

Incubation experiments were performed to determine the degradability of tDOC by photochemical remineralization and by microbial remineralization. For photo-chemical remineralization, we additionally measured the carbon isotopic fractionation.

For photodegradation experiments, surface water samples (<1m depth) were collected from the Maludam River in Malaysia (1.636°N 111.049°E), which drains a large area of intact peatland on northwestern Borneo, in December 2018 and June 2019, and from Kusu Island in the Singapore Strait in January 2020 (NE Monsoon) and July 2020 (SW Monsoon). Samples were filtered through pre-rinsed Whatman Polycap filters or 0.22 µm pore-size polyethersulfone membrane filters on the day of collection, stored at 4°C and re-filtered prior to the experiments. For the filtered water collected from the Maludam River in December 2018 and from the Singapore Strait, we filled 40 ml aliquots into replicate cylindrical quartz cells (50 mm pathlength, 50 mm diameter, with Teflon screw caps) and irradiated the samples in an Atlas Suntest CPS+ solar simulator fitted with a xenon lamp and daylight optical filter. The temperature control was set to 40 °C (the lowest possible setting), and light output between 300–400 nm was set to 40 W m<sup>-2</sup>. The quartz cells were opened and ventilated regularly to avoid depletion of oxygen and then recapped. A dark control was prepared by filling 200 ml

filtered water into a 250 ml Duran glass bottle, which was wrapped in aluminum foil and placed in the solar simulator. One or two replicate quartz cells were removed at regular time-points to measure DOC and CDOM (using 2-mm, 1-cm, or 10-cm pathlength quartz cuvettes) with parallel measurements of the dark control.

To measure carbon isotopic fractionation associated with photodegradation, the filtered water collected from the Maludam River in June 2019 was filled into replicate 150-ml quartz bottles with ground stoppers, leaving 30 ml headspace, and placed in an unshaded outdoor location at Nanyang Technological University, Singapore. Weather conditions during the experiment were partly sunny on most days, sometimes with heavy rain showers lasting up to a few hours. The ambient temperature varied between 26–32°C within a diurnal cycle. The quartz bottles were opened and ventilated once a week to avoid oxygen depletion and then recapped. A dark control was prepared as above, and placed in the same location. One replicate bottle was removed to measure DOC, CDOM (using 2-mm or 1-cm pathlength quartz cuvettes) and  $\delta^{13}C_{DOC}$  on Days 0, 60 and 98, with parallel measurements of the dark control.

For the biodegradation experiment,  $10 \, \text{L}$  of surface water (<1m depth) was collected from Kusu Island in the Singapore Strait in July 2020 (SW Monsoon), part of which was kept unfiltered as an inoculum while the rest was filtered through 0.22  $\mu$ m pore-size polyethersulfone membrane filters one day after collection. Three treatments were created: filtered water only (as sterile control), filtered water + inoculum (v/v = 5%), and filtered water + inoculum + labile

DOC (leucine and glucose, each contributed approximately 10 μmol L <sup>-1</sup> carbon, to test for a
possible priming effect). For each treatment, replicate 150 ml aliquots of the appropriate water
sample were filled into 250-mL Duran glass bottles, capped with airtight Teflon screw caps,
and kept in the dark in a shaded outdoor location at Nanyang Technological University,
Singapore. The ambient temperature varied between 26–32°C within a diurnal cycle. Bottles
were gently swirled every other day. For each treatment, three replicate samples were removed
and filtered through 0.22 $\mu m$ pore-size polyethersulfone membrane filters for DOC and CDOM
measurements on Days 0, 3, 7, 14, 27, 48 and 109. Bottles were cleaned with 10% hydrochloric
acid and DI water and combusted at 450 °C for 4 hours before use.

### 3 Results

460

461

462

463

464

465

466

467

468

469

470

471

472

473

474

475

476

477

478

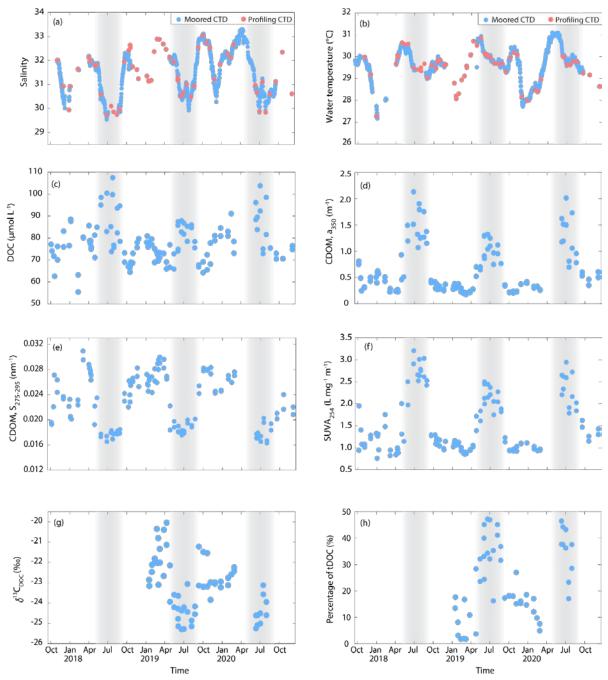
479

480

3.1 Monsoon-driven seasonal variation of tDOC input to the Singapore Strait During the SW Monsoon, the currents transport water from the east coast of Sumatra and the Malacca Strait to our sampling site in the Singapore Strait. These waters receive large riverine inputs from the extensive peatlands on Sumatra, and correspondingly we observed a decrease in salinity from ~33 to ≤30 during this period (Fig. 3a, Fig. S1a). The DOC concentration increased to 75–100 µmol L<sup>-1</sup>, which was 10–35 µmol L<sup>-1</sup> higher than in the inter-monsoon seasons (Fig. 3c). The CDOM absorption ( $a_{350}$ ) increased to 0.5–2 m<sup>-1</sup>, which was 2–10 times higher compared to inter-monsoon seasons (Fig. 3d). In addition, the SW Monsoon exhibited the lowest spectral slope of CDOM (S<sub>275-295</sub>) (0.016–0.020 nm<sup>-1</sup>), the highest SUVA<sub>254</sub> (2–3 L mg<sup>-1</sup> m<sup>-1</sup>) and the lightest  $\delta^{13}$ C<sub>DOC</sub> (-25.5 to -24‰) of the year, all of which indicate a large input of tDOC (Fig. 3e-g). The mass balance calculations showed that 20-50% of the DOC pool in the Singapore Strait was terrigenous during this period (Fig. 3h). During the early NE Monsoon (Dec-Jan), when the water flows from the South China Sea, we also observed a brief decrease in salinity by ~2 units. Because this is the time of year with the greatest precipitation in Singapore and southern Malaysia, some freshwater input from run-off would be expected at this time. However, the DOC and CDOM concentrations, the SUVA<sub>254</sub>, the  $S_{275-295}$ , and the  $\delta^{13}C_{DOC}$  only showed small changes relative to the inter-monsoon periods (Fig. 3c-g), which suggest that the tDOC input was limited. During the late NE Monsoon and

the following inter-monsoon season (late February to March; the season with lowest local

481 precipitation), continued water flow from the open South China Sea results in the highest salinity of the year ( $\sim$ 32–33), low DOC concentration ( $\sim$ 65 µmol L<sup>-1</sup>), low  $a_{350}$  (<0.25 m<sup>-1</sup>), 482 high  $S_{275-295}$  (>0.026 nm<sup>-1</sup>) and more enriched values of  $\delta^{13}C_{DOC}$  (-23 to -21‰) (Fig. 3c-g). 483 We therefore took the averages of our data during this period as the marine endmember values 484 485 for the conservative mixing model (Table 2). 486 The seawater temperature showed only small seasonal variation, mostly between 28–31°C (Fig. 487 3b). The chlorophyll-a concentration mostly ranged between 0.5–2 μg L<sup>-1</sup> and showed no clear 488 489 seasonal variation (full data set in Supplementary Data Table). 490



**Figure 3.** Seasonal variations in biogeochemistry in the Singapore Strait. Time series of (a) salinity, (c) DOC, (d-f) CDOM and (g) stable isotopic composition ( $\delta^{13}$ C) of DOC indicates a large input of freshwater and terrigenous dissolved organic matter to our site during the Southwest Monsoon season (May to September, highlighted by grey shading). The water temperature (b) shows small seasonal variation by ~3°C. The salinity and water temperature were both measured by a profiling CTD during each sampling cruise and by a moored CTD. The percentage of tDOC in the bulk DOC pool (h) was estimated from the  $\delta^{13}$ CDOC data.

Table 2. Endmember values  $(\pm 1SD)$  and the uncertainties (shown as  $\pm 1SD$ ) of the parameters measured at our site as input to the conservative mixing model and Monte Carlo uncertainty calculation.

Parameter	Riverine	Marine	Uncertainty of
	endmember	endmember	measured data
Salinity	0	$32.69 \pm 0.23$	± 0.01
DIC (μmol kg <sup>-1</sup> )	$453 \pm 34$	$1903 \pm 13$	$\pm 0.15\%$
TA (μmol kg <sup>-1</sup> )	$310 \pm 34$	$2164 \pm 15$	$\pm 0.13\%$
$\delta^{13}\mathrm{C}_{\mathrm{DIC}}$ (‰)	$-15.32 \pm 1$	$-0.25 \pm 0.09$	± 0.2
$\delta^{13}C_{DOC}$ (‰)	$-29 \pm 1$	$-21.88 \pm 0.79$	± 0.2
$\delta^{13}\mathrm{C}_{\mathrm{tDIC}}$ (‰)*	$-32 \pm 1$	-	-
DOC (μmol L <sup>-1</sup> )**	-	-	± 3.9

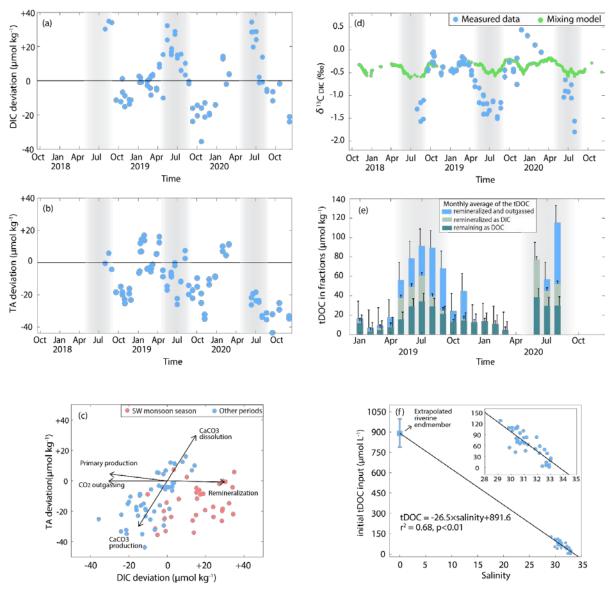
<sup>\*</sup> The  $\delta^{13}$ C of DIC produced from remineralization of peatland-derived tDOC.

### 3.2 Seasonal variation of DIC, TA and $\delta^{13}$ C<sub>DIC</sub> driven by tDOC remineralization

During the SW Monsoon, the DIC was mostly 15–40 μmol kg<sup>-1</sup> higher than the values expected from the conservative mixing model, while the TA typically showed a depletion of 10–40 μmol kg<sup>-1</sup> relative to conservative mixing (Fig. 4a, b). By comparing the measured deviations with the known changes caused by different biogeochemical processes, we found that the variation in DIC and TA during the SW Monsoon was chiefly the result of remineralization and calcium carbonate production (Fig. 4c). Because the Singapore Strait was always oversaturated with respect to pCO<sub>2</sub> (Section 3.4), net CO<sub>2</sub> outgassing, instead of CO<sub>2</sub> invasion, is expected throughout the year. Because the net CO<sub>2</sub> outgassing and calcium carbonate production both remove DIC from seawater, it is likely that the total quantity of remineralized tDOC is in fact greater than the measured positive deviation of DIC concentration during the SW Monsoon. We therefore turn to δ<sup>13</sup>C<sub>DIC</sub> to estimate the total amount of tDOC remineralization.

<sup>\*\*</sup> DOC endmember values were not used in the calculations.

The  $\delta^{13}C_{DIC}$  in the inter-monsoon seasons varied between -0.4 and 0‰. It decreased to between -1.0 and -1.8‰ during the SW Monsoon (Fig. 4d). However, our conservative mixing model predicts that  $\delta^{13}C_{DIC}$  would vary by only 0.3‰ as a result of riverine DIC input (Fig. 4d). This small predicted range is a result of the low DIC concentration in peat-draining, low-pH rivers. The remaining 0.7–1.5‰ decrease in the  $\delta^{13}C_{DIC}$  therefore indicates production of extra DIC from an isotopically depleted carbon source, consistent with the remineralization of tDOC within the shelf waters.



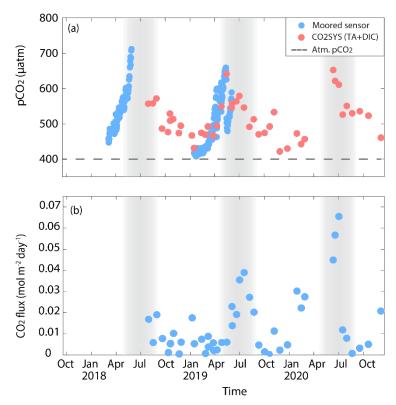
**Figure 4.** (a – b) Time series of deviations of DIC and TA relative to values expected from the conservative mixing model. DIC shows positive deviation during the SW Monsoon Season (grey shading), coincident with the large tDOC input. (c) Scatterplot of deviation of DIC versus deviation of TA, which indicates a strong signal of remineralization during the SW Monsoon season. (d) The  $\delta^{13}$ C of DIC showed strong depletion relative to the mixing model during the SW Monsoon season, indicating remineralization of tDOC. (e) Monthly average quantity of (1) the remaining tDOC, (2) remineralized tDOC that was still present as DIC, (3) remineralized tDOC that had already outgassed to the atmosphere; as estimated by the isotope mass balance calculations. The total height of each stacked bar represents the total initial tDOC concentration in that water before remineralization. The error bars show one standard deviation of each tDOC fraction, as estimated by a Monte Carlo simulation (shown in one direction for clarity). (f) The relationship between salinity and the total initial tDOC concentration yields a predicted peatland riverine endmember DOC concentration of 892±104 µmol L<sup>-1</sup> (mean ± standard error).

## 544 3.3 Quantification of tDOC remineralization by isotope mass balance calculations The isotope mass balance calculations provide a breakdown of tDOC remineralization into 545 three fractions (see Section 2.4.2). We assumed that all remineralized terrigenous carbon is 546 547 derived from tDOC, i.e. that terrigenous POC does not contribute to the signals we observe, and that production and remineralization of autochthonous (i.e., marine) DOC are 548 549 approximately in steady state and do not vary seasonally. The validity of these assumptions is discussed in Section 4.1. 550 551 552 Our SW Monsoon data (Fig. 4e) show that the initial total tDOC concentration (represented by the total height of each stacked bar) amounted to between 60–140 µmol kg<sup>-1</sup>. Of this initial 553 tDOC, 20-40 µmol kg<sup>-1</sup>, or 30-40%, was still present as unremineralized tDOC at our site, 554 555 while the remaining 60-70% had been remineralized in the shelf sea before reaching our site. Of this remineralized tDOC, 10-40 µmol kg<sup>-1</sup> was still present in the DIC pool, while 15-80 556 umol kg<sup>-1</sup> had been lost to the atmosphere. The proportion of remineralized tDOC that had 557 been removed by outgassing increased over the course of the SW Monsoon. The uncertainty of 558 each carbon fraction ranged from 5-15 µmol kg<sup>-1</sup> as estimated by a Monte Carlo simulation, 559 indicating an overall quite well-constrained tDOC budget. 560 561 562 The initial total tDOC concentration showed a strong linear relationship with salinity ([DOC] = $-26.5 \times salinity + 891.6$ , $r^2=0.68$ , p<0.01, Fig. 4f), the intercept of which implies an average 563 riverine endmember DOC concentration of $892 \pm 104 \mu mol L^{-1}$ (mean $\pm$ standard error). This 564

value is in remarkably close agreement with the reported discharge-weighted mean DOC concentration of Sumatra's main peat-draining rivers, which is  $890 \pm 159 \,\mu\text{mol}\,L^{-1}$  (Wit et al., 2015), and is too high to be explained by non-peatland tDOC sources.

### 3.4 Seasonal variation of pCO<sub>2</sub> and the sea-air CO<sub>2</sub> yield

The pCO<sub>2</sub> in the Singapore Strait showed oversaturation throughout the year, as well as pronounced seasonal variation. During the inter-monsoon season, pCO<sub>2</sub> was below 500  $\mu$ atm, but it increased to 550–700  $\mu$ atm during the SW Monsoon (Fig. 5a), consistent with remineralization of tDOC having occurred prior to reaching the Singapore Strait. The high pCO<sub>2</sub> values during the SW Monsoon translated to a daily areal sea-to-air CO<sub>2</sub> flux of 15–68 mmol C day<sup>-1</sup> m<sup>-2</sup> (Fig. 5b). The annual areal sea-to-air CO<sub>2</sub> flux in the Singapore Strait, amounted to  $4.10\pm0.47$  mol C m<sup>-2</sup> yr<sup>-1</sup> in 2019 and  $8.20\pm1.17$  mol C m<sup>-2</sup> yr<sup>-1</sup> in 2020.



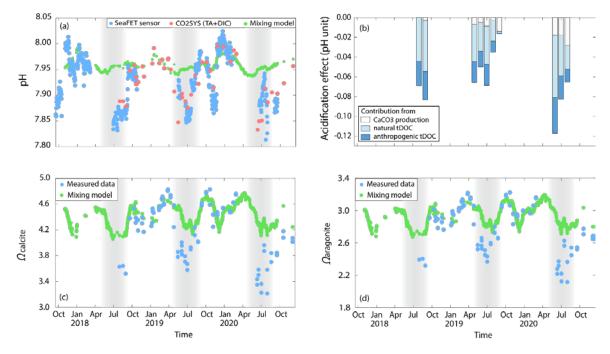
**Figure 5.** Time series of (a) pCO<sub>2</sub> and the (b) daily areal CO<sub>2</sub> flux. The pCO<sub>2</sub> was measured by a moored SAMI pCO<sub>2</sub> sensor and was validated by the laboratory analysis of TA and DIC using CO2SYS. The data from the latter approach were used for calculation of the CO<sub>2</sub> yield. The Southwest Monsoon season is highlighted by the grey shading.

### 3.5 Seasonal acidification and putative anthropogenic contribution

Seawater pH in the Singapore Strait was 7.95–8.03 during the NE Monsoon and the intermonsoon seasons, when terrestrial input was minor. The pH decreased by up to 0.14 units, to 7.90–7.85, during the SW Monsoon (Fig. 6a, Fig. S1b). This pH decrease could be driven by freshwater input, calcium carbonate production, and/or remineralization of organic carbon. Our conservative mixing model only predicts decreases in seawater pH at our site of ~0.02 units, indicating that input of river water can only explain a small proportion of this variation (Fig. 6a). The contribution from calcium carbonate production was constrained from the isotope

mass balance calculations, and was only responsible for a small additional pH reduction, mostly <0.02 units (Fig. 6b). The majority of the pH decrease, 0.06–0.10 units, was therefore attributable to the remineralization of tDOC. Assuming that 35% of the total tDOC input during the SW Monsoon, and hence also 35% of the tDOC remineralization, might be an anthropogenic contribution caused by peatland disturbance (Moore et al., 2013; Yupi et al., 2016), we further estimate that 0.01–0.04 pH units of this seasonal acidification could be anthropogenic (Fig. 6b).

This seasonal variation in the carbonate system also entails changes in the saturation state of calcite ( $\Omega_{CA}$ ) and aragonite ( $\Omega_{AR}$ ). During the late NE Monsoon and inter-monsoon,  $\Omega_{CA}$  and  $\Omega_{AR}$  were 4.62±0.09 and 3.07±0.06, respectively (Fig. 6c). During the SW Monsoon, the  $\Omega_{CA}$  decreased to 3.2–4.0 and the  $\Omega_{AR}$  decreased to 2.1–2.7. These decreases exceeded the decreases predicted by the conservative mixing model by a factor of  $\geq$ 2, demonstrating the potential for tDOC remineralization to affect calcifying communities in this region.



**Figure 6.** Acidification during the SW Monsoon season (grey shading) due to the extensive remineralization of tDOC. (a) Time series of pH from the SeaFET sensor and laboratory measurements of TA and DIC shows reduced pH relative to values expected from conservative mixing during the SW Monsoon. (b) Individual contribution to the seasonal acidification from  $CaCO_3$  production, remineralization of natural, pre-anthropogenic tDOC, and remineralization of the putative anthropogenic tDOC fraction, based on the results from the mass balance calculations and the reported anthropogenic increase in tDOC export from peatlands (Moore et al. 2013; Yupi et al. 2016). The total acidification effect (height of the stacked bars) is the difference between the measured pH and the predicted value from the conservative mixing model. The results are presented as monthly averages for clarity.  $(\mathbf{c} - \mathbf{d})$  The saturation states of calcite and of aragonite show large decreases compared to the conservative mixing model during the SW Monsoon season.

### 3.6 Degradability of the peatland-derived tDOC

The tDOC from the peatland-draining Maludam River showed high photo-lability (Fig. 7, Table 3): the DOC concentration decreased first from 3250 µmol L<sup>-1</sup> to 850 µmol L<sup>-1</sup> over the course of 525 hours in the solar simulator (Fig. 7a), after which we observed pronounced photo-induced flocculation in the remaining replicates. Although POC was not measured, the quantity

628

629

630

631

632

633

634

635

636

637

638

639

640

641

642

643

644

645

646

647

648

of flocs did not visibly decrease during the remaining hours of the experiment, suggesting that photo-flocculated DOC probably remained stable. CDOM  $a_{350}$  decreased from 167.4 m<sup>-1</sup> to 5.9 m<sup>-1</sup>, and the decrease in SUVA<sub>254</sub> and increase in S<sub>275-295</sub> indicated the preferential loss of aromatic and high-molecular-weight compounds (Helms et al., 2008; Weishaar et al., 2003) (Fig. 7b-d). The decrease in DOC observed up to 525 hours implies that around 74% of tDOC was labile to photochemical remineralization. The experiment in which Maludam River water was exposed to natural sunlight, revealed that photodegradation also resulted in a small but sizable isotopic fractionation (Fig. 7e-h, Table 3). The DOC concentration decreased by 45% from 3069  $\mu$ mol L<sup>-1</sup> to 1674  $\mu$ mol L<sup>-1</sup>, and  $\delta$ <sup>13</sup>C<sub>DOC</sub> increased from -31.1% to -29.9%. This implies that the resulting DIC had a  $\delta^{13}$ C value of -32.5%, and that photodegradation of this tDOC therefore entailed a fractionation of -1.4%. As in the previous experiment, CDOM and SUVA<sub>254</sub> decreased strongly, while S<sub>275-295</sub> increased. Samples collected from the Singapore Strait during the NE (Fig. 7i–l) and SW Monsoon (Fig. 7m-p) both showed pronounced losses of CDOM upon experimental irradiation for >300 hours, but at most small losses of DOC. There was a decrease in  $a_{350}$  by >70%, and strong decreases in SUVA<sub>254</sub> and increases in S<sub>275-295</sub> during both seasons. However, the NE Monsoon sample showed no change in DOC concentration (Fig. 7i); the SW Monsoon sample only showed a decrease in DOC concentration by 9 µmol L<sup>-1</sup> (Fig. 7m). Of the bulk DOC in this SW Monsoon

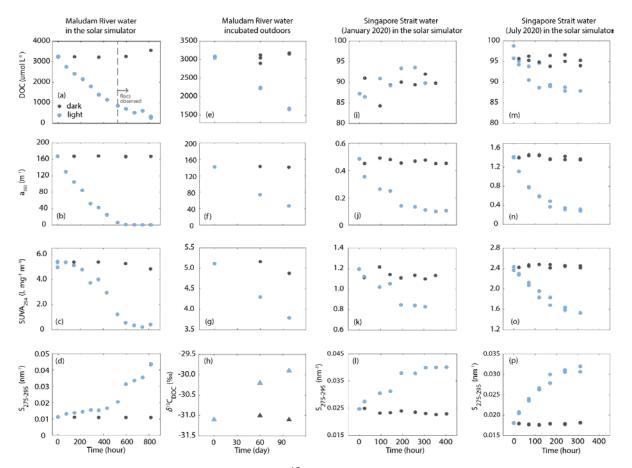
sample,  $36 \,\mu\text{mol}\,L^{-1}$  was tDOC (estimated from the  $\delta^{13}C_{DOC}$ ). Because the majority of the DOC pool during the NE Monsoon is of marine origin, it appears that the autochthonous marine DOC is not strongly photolabile. Assuming therefore that the observed decrease in DOC in the SW Monsoon experiment was entirely due to tDOC remineralization, our results would indicate that only about 25% (9 out of 36  $\mu$ mol  $L^{-1}$ ) of the peatland-derived tDOC that still remains unremineralized in the Singapore Strait is labile to direct photochemical remineralization.

Moreover, we found that the biodegradability of the SW Monsoon sample from the Singapore Strait was low, with <6% loss of DOC and CDOM  $a_{350}$  over the course of the 109-day incubation (Fig. 8). Although the leucine and glucose added as labile carbon to one treatment was rapidly consumed, no additional remineralization of DOC was induced in this treatment. Our results, consistent with the low biodegradability of tDOC from the Maludam River (Nichols & Martin, submitted), suggest that direct microbial remineralization is not a main driver of tDOC remineralization. Whether the observed biodegradation represents loss of tDOC at all or only of autochthonous DOC is unclear.

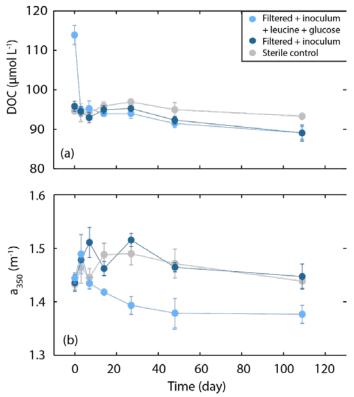
In summary, our experiments with the Maludam River samples indicate that around 74% of tDOC is directly photo-mineralizable, while our observational data from the Singapore Strait show that 60–70% of the peatland-derived tDOC is remineralized in the coastal waters of the Sunda Shelf. Our additional experiments then show that of the remaining 30–40% of this tDOC,

<25% is still photo-mineralizable, and that any biodegradation proceeds at most slowly.</p>
Depending on how much of the remaining tDOC really is biodegradable, these results imply that possibly around 20–30% of the total peatland tDOC from Sumatra is sufficiently refractory to resist remineralization on the Sunda Shelf and be exported to the Indian Ocean, given that the total water residence time in the Malacca Strait is approximately 2 years (Mayer et al., 2018).





**Figure 7.** Changes in DOC, CDOM and  $\delta^{13}C_{DOC}$  during four photodegradation experiments showing high photo-lability of DOC from the Maludam River but only limited photo-lability from the Singapore Strait. Flocs were observed in the water sample after incubation for 525 hours during the experiment with the Maludam River water in the solar simulator but were not observed in other experiments.



**Figure 8.** Changes in **(a)** DOC and **(b)** CDOM (a<sub>350</sub>) during the 109-day incubation showing limited biodegradability of DOC collected from the Singapore Strait.

Table 3. Results of the photodegradation experiments.

	DOC	a <sub>350</sub>	S <sub>275-295</sub>	SUVA <sub>254</sub>	$\delta^{13} C_{DOC}$	$\delta^{13}C_{DIC}**$
	(µmol L <sup>-1</sup> )	$(m^{-1})$	(nm <sup>-1</sup> )	$(L mg^{-1} m^{-1})$	(‰)	(‰)
Maludam River						
in solar simulator*						
before	3250	167.4	0.011	5.41		
after	850	5.9	0.021	1.23		
%loss	74%	96%				
Maludam River						
<b>Incubated outdoors</b>						
before	3069	140.88	0.012	5.12	-31.1	-
after	1674	46.41	0.015	3.79	-29.9	-32.5
%loss	45%	67%				
Singapore Strait,						
NE Monsoon						
(Jan 2020)						
before	87	0.49	0.025	1.20		
after	89	0.11	0.040	0.83		
%loss	-	78%				
Singapore Strait,						
SW Monsoon						
(July 2020)						
before	97	1.40	0.018	2.40		
after	88	0.29	0.032	1.53		
%loss	9%	79%				

<sup>\*</sup> Data beyond 525 hours were omitted because flocculation observed.

<sup>\*\*</sup>  $\delta^{13}$ C of the DIC produced from remineralization of the initial DOC.

### **4 Discussions**

707

708

709

710

711

712

713

714

715

716

717

718

719

720

721

722

### 4.1 Sources and remineralization of tDOC

The correlated seasonal variations of salinity, DOC,  $\delta^{13}C_{DOC}$ , CDOM absorption and spectral characteristics all indicate that our site receives a large input of tDOC during the SW Monsoon, and a much smaller tDOC input during the NE Monsoon. Although seasonal variation in rainfall can cause variation in tDOC flux from tropical peatlands (Rixen et al., 2016), the variation observed here is driven by the regional advection of coastal water that has mixed with peatland-draining river input according to the monsoon system (Van Maren & Gerritsen, 2012; Mayer & Pohlmann, 2014). The high riverine endmember DOC concentration (892 ± 104) umol L<sup>-1</sup>) predicted from the relationship between the total initial tDOC concentration and salinity (Fig. 4f) clearly points to peatlands as the source of the tDOC measured during the SW Monsoon. This seasonally variable advection of peatland tDOC explains the lack of a strong peatland signal in the carbonate system data measured by Hamzah et al. (2020) in the Karimata Strait to the southeast of our site: because they sampled during the SW Monsoon, the peatlandinfluenced water would have flowed northwards away from their transect and out to the South China Sea.

723

724

725

726

727

Our isotope mass balance calculations assume that the seasonal variation in our data is not driven by seasonal changes in autochthonous DOC remineralization. The relatively low chlorophyll-a concentrations year-round suggest that autochthonous organic matter production rates are moderate and, importantly, do not show strong seasonality. This is likely due to

persistent light limitation caused by strong vertical light attenuation (Chow et al., 2019; Morgan et al., 2020) and tidal mixing of the water column (Mayer & Pohlmann, 2014) which prevents vertical stratification. Moreover, the very small annual range in seawater temperature means that microbial physiological rates will not vary strongly. Therefore, the net community production rate is most likely relatively constant, and we can rule out the possibility that our observations are the result of seasonal changes in production and remineralization of marine autochthonous organic matter. Moreover, the seasonal decrease in  $\delta^{13}C_{DIC}$  is too large to be explained by the remineralization of marine DOC with a  $\delta^{13}C_{DIC}$  of around -22%. Even though we attributed all the excess DIC to remineralization of tDOC with  $\delta^{13}C$  of DIC production of -32%, the depletion in  $\delta^{13}C_{DIC}$  was so large that it could only be explained by inferring that an even greater quantity of tDOC had been remineralized and outgassed.

We further assume that the remineralized terrigenous carbon is tDOC rather than POC. The terrigenous organic carbon in peat-draining rivers in Southeast Asia is overwhelmingly in form of DOC year-round (typically  $\geq$ 95%) (Alkhatib et al., 2007; Baum et al., 2007; Moore et al., 2011; Müller et al., 2015), and thus any terrigenous POC input is expected to be small in this region. Part of any POC that is delivered by rivers is also likely to sink and accumulate in the sediment, thus decoupling the POC and its remineralization from the water carrying the DOC. Most importantly, the riverine endmember DOC concentration that we infer from our regression of total initial tDOC concentrations *versus* salinity (892  $\pm$  104  $\mu$ mol L<sup>-1</sup>) is also very close to the discharge-weighted average DOC concentration measured independently in the

main peatland-draining rivers on Sumatra, of 890  $\pm$  159  $\mu$ mol L<sup>-1</sup> (Wit et al., 2018). This indicates that additional remineralization of POC is not required to explain the observed variation in DIC and  $\delta^{13}C_{DIC}$ .

752

753

754

755

756

757

758

759

760

761

762

763

764

765

766

767

768

749

750

751

Remineralization of tDOC can occur either by microbial decomposition (i.e., biodegradation) or by photochemical decomposition. Biodegradation is increasingly recognized as an important pathway of tDOC decomposition in aquatic systems (Tranvik et al., 2009; Ward et al., 2013; Wickland et al., 2007). However, peatland organic matter especially in the tropics is enriched in polyphenolic compounds such as lignin derivatives, which are likely refractory to microbial respiration (Gandois et al., 2014; Hodgkins et al., 2018). Our data showed at most low biodegradability of the tDOC collected from the Singapore Strait, even when amended with labile carbon to stimulate a potential priming effect (Bianchi, 2011; Guenet et al., 2010; van Nugteren et al., 2009). Similarly, freshly collected tDOC from the peatland-draining Maludam River showed at most very limited biodegradability over 50 days, even when amended with dissolved nutrients or mixed into coastal seawater (Nichols and Martin, submitted). In contrast, we found that up to about 74% of peatland-derived tDOC can be photochemically remineralized. This is consistent with previous reports of high photo-lability of tropical peatland tDOC (Gandois et al., 2020; Martin et al., 2018; Spencer et al., 2009). Collectively, these results suggest that photodegradation rather than biodegradation likely drives the observed remineralization.

It is possible that partial photodegradation increases the biodegradability of the peatland tDOC (Cory & Kling, 2018; Moran et al., 2000; Moran & Zepp, 1997). For example, tDOC processing in Arctic freshwater systems depends on initial photodegradation that stimulates subsequent microbial remineralization (Cory et al., 2014), and 32% of the tDOC remineralization on the Louisiana Shelf was attributed to photochemically stimulated biodegradation (Fichot & Benner, 2014). Our data are consistent with preferential degradation of high-molecular-weight aromatic substances during irradiation, which is believed to increase tDOC biodegradability. Further studies are required to address the importance of photochemical–microbial interactions for Southeast Asian peatland tDOC decomposition.

Our data showing that the majority of tDOC is remineralized in the Sunda Shelf Sea are consistent with previous work showing that ocean margins are hotspots of tDOC processing (Fichot & Benner, 2014; Kaiser et al., 2017; Kitidis et al., 2019; Painter et al., 2018; Semiletov et al., 2016). This is also comparable to a recent estimate that 63% of Southeast Asian peatland carbon is remineralized, derived by scaling up the estimated sea-to-air CO<sub>2</sub> fluxes in the southern Malacca Strait and Karimata Strait (Wit et al., 2018). This study also inferred that the majority of the remaining tDOC would also be remineralized in the shelf. However, our incubations showed surprisingly low remineralizability of the remaining tDOC reaching our site by microbial and photo-chemical decomposition. Consequently, our data suggest that it is possible that roughly 20–30% of the tDOC from Sumatran peatlands is actually exported to the open Indian Ocean and ultimately contributes to the semi-refractory or even refractory oceanic

791 DOC pool.

792

793

794

795

796

797

798

799

800

801

802

803

804

805

806

807

808

809

810

## 4.2 CO<sub>2</sub> emissions from the Sunda Shelf driven by tDOC remineralization

The mass balance calculations showed that the proportion of the remineralized tDOC that had been removed by CO<sub>2</sub> outgassing increased throughout the SW Monsoon, indicating the ongoing removal of CO<sub>2</sub> to atmosphere. Our estimate that the Singapore Strait contributes a net sea-to-air CO<sub>2</sub> flux of 4.10–8.20 mol C m<sup>-2</sup> yr<sup>-1</sup> is consistent with the paradigm that tropical inner shelves tend to act as sources of atmospheric CO<sub>2</sub> (Cai, 2011; Chen et al., 2013; Chen & Borges, 2009). Our estimate is comparable to that of Wit et al. (2018) for the same region (4.12 mol C m<sup>-2</sup> yr<sup>-1</sup>), but is much higher than the estimates by Roobaert et al. (2019) (0–0.4 mol C m<sup>-2</sup> yr<sup>-1</sup>) and Hamzah et al. (2020) (1.09 mol C m<sup>-2</sup> yr<sup>-1</sup>). The estimates of Roobaert et al. (2019) were based on a global data product that lacked direct measurements from the Sunda Shelf Sea, while Hamzah et al. (2020) sampled in the southern Karimata Strait during the SW Monsoon, at which point the peatland tDOC is advected northeastwards into the South China Sea. The fact that the Sunda Shelf is clearly a stronger CO<sub>2</sub> source than estimated by some studies demonstrates the importance of collecting annual time series of carbon biogeochemistry in this region, and of increasing the data coverage for coastal waters in Southeast Asia. Given the potentially sizeable anthropogenic contribution to the peatland tDOC input to the Sunda Shelf, our study further helps to highlight the importance of downstream greenhouse gas emissions from tropical peatland disturbance.

# 4.3 Seasonal acidification driven by tDOC remineralization

During the SW Monsoon, 10–30 µmol kg<sup>-1</sup> of the remineralized tDOC is still present as DIC upon reaching the Singapore Strait, resulting in seasonal acidification by 0.60–0.10 pH units. Pronounced impacts of tDOC remineralization on the carbonate system were reported for the estuaries and coastal waters of eastern Sumatra (Wit et al., 2018). Our results further show that the seasonal advection of these waters subjects sensitive calcifying communities such as coral reefs, which are found further away from the peatland river estuaries around the islands to the east of Sumatra (UNEP-WCMC, 2010), to pronounced seasonal acidification.

Ocean acidification, which leads to decrease in the saturation state ( $\Omega$ ) of calcium carbonate, is thought to impair the growth of corals by reducing the calcification rates (Cooper et al., 2008; Langdon, 2005; Pisapia et al., 2019) and skeletal density (Manzello et al., 2008; Mollica et al., 2018), and cause degradation of the coral reefs such as coral bleaching (Anthony et al., 2008; DeCarlo et al., 2017; Fabricius et al., 2011), reduction of coral cover (Gardner et al., 2003; Hughes et al., 2018; Perry et al., 2018) and decrease in coral species diversity (Fabricius et al., 2011). Zero or negative carbonate accretion on coral reefs might be induced when the  $\Omega_{AR}$  decreases to  $\sim$ 3.3 or below (Hoegh-Guldberg et al., 2007; Kleypas et al., 1999), and large-scale reductions in calcification have been projected for future high-pCO<sub>2</sub> scenarios due to anthropogenic CO<sub>2</sub> emissions (Andersson et al., 2007; Anthony et al., 2008; Dove et al., 2013; Hoegh-Guldberg et al., 2007). The decrease in  $\Omega_{AR}$  to  $\sim$ 2.7 at our site during the SW Monsoon

might therefore reduce coral calcification and reef development during this period of the year, as has been shown in response to seasonal carbonate system variation in the Great Barrier Reef (Dove et al., 2013; Stoltenberg et al., 2021). Our analysis indicates that the reported increases in tDOC flux after peatland disturbance are sufficiently large to account for a significant part of the observed acidification. This demonstrates that anthropogenic disturbance of tropical peatlands might be impacting sensitive ecosystems such as coral reefs via increased fluvial tDOC export and remineralization.

#### **5 Conclusions**

854

855

856

857

858

859

860

861

862

863

864

865

866

867

868

869

870

Our results demonstrate that the large input of tDOC from peatlands on Sumatra affects regional DOC concentrations, CO<sub>2</sub> fluxes, and the carbonate system in the Sunda Shelf Sea. Isotope mass balance calculations indicate that 60–70% of this tDOC is remineralized in the coastal waters of shelf sea close to the peatlands, and incubation experiments suggest that photodegradation drives a major part of this remineralization. The increase in DIC concentration caused by this remineralization results in a seasonal decrease in seawater pH by up to 0.10 units, of which the remineralization of the putative anthropogenic tDOC fraction might account for a pH reduction of 0.01–0.04 units, potentially impacting coral physiology in the region. Most of the CO<sub>2</sub> produced from this tDOC remineralization is likely eventually lost to the atmosphere. Our data thus provide further evidence that there is a large indirect pathway of CO<sub>2</sub> emissions due to peatland disturbance via the remineralization of tDOC in coastal seas. Despite this rapid remineralization of a large fraction of tDOC, we also found low degradability of the residual tDOC pool. We conclude that the Sunda Shelf is an efficient sink of peatlandderived tDOC and a significant source of atmospheric CO<sub>2</sub>, but that a higher proportion of the peatland tDOC might be refractory and exported to the Indian Ocean than was previously thought. The long-term fate of this tDOC flux to the open ocean remains uncertain.

871

872

873

#### Acknowledgements

We thank Molly Moynihan, Nikita Kaushal, Robert Nichols, Ashleen Tan Su Ying, Chen Shuang, Woo Oon Yee, Kyle Morgan and Phyllis Kho Yu Yi for help during field work and laboratory analysis. We are very grateful to Nikita Kaushal for measuring  $\delta^{13}C_{DIC}$ , to Nathalie Goodkin for providing the moored CTD and pH sensors for this work, to Evelyn Lim for performing some of the photodegradation experiments, to Moritz Müller for collecting water samples from the Maludam River, and to Francis Yeo, Sapari and Surpato of *Dolphin Explorer* for enabling the sample collection. This research was funded by the National Research Foundation Singapore, Prime Minister's Office, as part of the Marine Science Research and Development Programme through grant MSRDP-P32 to P.M., and forms part of the Ph.D. research of Y.Z.

#### **Author Contributions**

P.M. conceptualized the research and acquired funding; Y.Z., Y.C., K.Y.W.C. and P.M. conducted field work; Y.Z. and Y.C. conducted degradation experiments; Y.Z., Y.C. and K.Y.W.C. analyzed samples; Y.Z. analyzed the data with support from C.D.E., P.M., Y.C. and K.Y.W.C.; Y.Z., P.M., C.D.E. and Y.C. wrote the manuscript. All authors edited the manuscript and approved the final version.

## **Competing Interests**

None of the authors have competing interests to declare.

896 Data availability 897 898 All processed data are available in the Supplementary Data Table. All raw data files and 899 analysis codes will be made available through the Nanyang Technological University Data 900 Repository upon acceptance, under doi:10.21979/N9/MVYDRU. 901 902 **Materials & Correspondence** 903 Correspondence and request for materials should be addressed to Yongli Zhou: 904 zhou0303@e.ntu.edu.sg or Patrick Martin: pmartin@ntu.edu.sg. 905 906 907 References Alkhatib, M., Jennerjahn, T. C., & Samiaji, J. (2007). Biogeochemistry of the Dumai River 908 909 estuary, Sumatra, Indonesia, a tropical blackwater river. Limnology and Oceanography, 910 52(6), 2410–2417. https://doi.org/10.4319/lo.2007.52.6.2410 Alling, V., Humborg, C., Mörth, C.-M., Rahm, L., & Pollehne, F. (2008). Tracing terrestrial 911 organic matter by  $\delta^{34}$  S and  $\delta^{13}$ C signatures in a subarctic estuary. Limnology and 912 913 Oceanography, 53(6), 2594–2602. https://doi.org/10.4319/lo.2008.53.6.2594 914 Andersson, A. J., Bates, N. R., & Mackenzie, F. T. (2007). Dissolution of carbonate 915 sediments under rising pCO<sub>2</sub> and ocean acidification: Observations from Devil's Hole, Bermuda. Aquatic Geochemistry, 13(3), 237–264. https://doi.org/10.1007/s10498-007-916

917

9018-8

918 Anthony, K. R. N., Kline, D. I., Diaz-Pulido, G., Dove, S., & Hoegh-Guldberg, O. (2008). 919 Ocean acidification causes bleaching and productivity loss in coral reef builders. 920 Proceedings of the National Academy of Sciences of the United States of America, 921 105(45), 17442–17446. https://doi.org/10.1073/pnas.0804478105 922 Assayag, N., Rivé, K., Ader, M., Jézéquel, D., & Agrinier, P. (2006). Improved method for 923 isotopic and quantitative analysis of dissolved inorganic carbon in natural water 924 samples. Rapid Communications in Mass Spectrometry, 20(15), 2243–2251. 925 https://doi.org/10.1002/rcm.2585 926 Baum, A. (2008). Tropical blackwater biogeochemistry: The Siak River in Central Sumatra, 927 Indonesia. PhD Thesis, University of Bremen, Bremen, Germany. Baum, A., Rixen, T., & Samiaji, J. (2007). Relevance of peat draining rivers in central 928 929 Sumatra for the riverine input of dissolved organic carbon into the ocean. Estuarine, 930 Coastal and Shelf Science, 73(3–4), 563–570. https://doi.org/10.1016/j.ecss.2007.02.012 931 Bauman, A. G., Hoey, A. S., Dunshea, G., Feary, D. A., Low, J., & Todd, P. A. (2017). 932 Macroalgal browsing on a heavily degraded, urbanized equatorial reef system. Scientific 933 Reports, 7(1), 1–8. https://doi.org/10.1038/s41598-017-08873-3 934 Beleites, C., & Sergo, V. (2012). hyperSpec: a package to handle hyperspectral data sets in R. 935 Journal of Statistical Software. 936 Bianchi, T. S. (2011). The role of terrestrially derived organic carbon in the coastal ocean: A 937 changing paradigm and the priming effect. Proceedings of the National Academy of Sciences, 108(49), 19473–19481. https://doi.org/10.1073/pnas.1017982108 938

939 Cai, W.-J. (2011). Estuarine and Coastal Ocean Carbon Paradox: CO<sub>2</sub> Sinks or Sites of Terrestrial Carbon Incineration? Annual Review of Marine Science, 3(1), 123–145. 940 941 https://doi.org/10.1146/annurev-marine-120709-142723 942 Cao, X., Aiken, G. R., Butler, K. D., Huntington, T. G., Balch, W. M., Mao, J., & Schmidt-943 Rohr, K. (2018). Evidence for major input of riverine organic matter into the ocean. 944 Organic Geochemistry, 116, 62–76. https://doi.org/10.1016/j.orggeochem.2017.11.001 945 Chen, C.-T. A., & Borges, A. V. (2009). Reconciling opposing views on carbon cycling in 946 the coastal ocean: Continental shelves as sinks and near-shore ecosystems as sources of 947 atmospheric CO<sub>2</sub>. Deep Sea Research Part II: Topical Studies in Oceanography, 56(8– 10), 578–590. https://doi.org/10.1016/j.dsr2.2009.01.001 948 Chen, C.-T. A., Huang, T.-H., Chen, Y.-C., Bai, Y., He, X., & Kang, Y. (2013). Air-sea 949 950 exchanges of CO<sub>2</sub> in the world's coastal seas. *Biogeosciences*, 10(10), 6509–6544. 951 https://doi.org/10.5194/bg-10-6509-2013 952 Cheng, L., Normandeau, C., Bowden, R., Doucett, R., Gallagher, B., Gillikin, D. P., et al. 953 (2019). An international intercomparison of stable carbon isotope composition 954 measurements of dissolved inorganic carbon in seawater. Limnology and Oceanography: Methods, 17(3), 200–209. https://doi.org/10.1002/lom3.10300 955 956 Chow, G. S. E., Chan, Y. K. S., Jain, S. S., & Huang, D. (2019). Light limitation selects for 957 depth generalists in urbanised reef coral communities. Marine Environmental Research, 958 147(February), 101–112. https://doi.org/10.1016/j.marenvres.2019.04.010 Chupakova, A. A., Chupakov, A. V., Neverova, N. V., Shirokova, L. S., & Pokrovsky, O. S. 959

960 (2018). Photodegradation of river dissolved organic matter and trace metals in the largest European Arctic estuary. Science of The Total Environment, 622-623, 1343-961 962 1352. https://doi.org/10.1016/j.scitotenv.2017.12.030 963 Cooper, T. F., De'ath, G., Fabricius, K. E., & Lough, J. M. (2008). Declining coral 964 calcification in massive Porites in two nearshore regions of the northern Great Barrier 965 Reef. Global Change Biology, 14(3), 529-538. https://doi.org/10.1111/j.1365-966 2486.2007.01520.x Cory, R. M., & Kling, G. W. (2018). Interactions between sunlight and microorganisms 967 968 influence dissolved organic matter degradation along the aquatic continuum. Limnology and Oceanography Letters. https://doi.org/10.1002/lol2.10060 969 Cory, R. M., Ward, C. P., Crump, R. C., & Kling, G. W. (2014). Sunlight controls water 970 971 column processing of carbon in arctic fresh waters, 345(6199). 972 CYGNSS. (2017). CYGNSS Level 2 Science Data Record Version 2.1. NASA Physical 973 Oceanography DAAC. https://doi.org/10.5067/CYGNS-L2X21 974 DeCarlo, T. M., Cohen, A. L., Wong, G. T. F., Shiah, F.-K., Lentz, S. J., Davis, K. A., et al. 975 (2017). Community production modulates coral reef pH and the sensitivity of ecosystem calcification to ocean acidification. Journal of Geophysical Research: Oceans, 122(1), 976 977 745–761. https://doi.org/10.1002/2016JC012326 978 Dommain, R., Couwenberg, J., Glaser, P. H., Joosten, H., & Suryadiputra, I. N. N. (2014). 979 Carbon storage and release in Indonesian peatlands since the last deglaciation. Quaternary Science Reviews, 97, 1–32. https://doi.org/10.1016/j.quascirev.2014.05.002 980

981 Dove, S. G., Kline, D. I., Pantos, O., Angly, F. E., Tyson, G. W., & Hoegh-Guldberg, O. (2013). Future reef decalcification under a business-as-usual CO<sub>2</sub> emission scenario. 982 983 Proceedings of the National Academy of Sciences of the United States of America, 984 110(38), 15342–15347. https://doi.org/10.1073/pnas.1302701110 985 Evans, C. D., Page, S. E., Jones, T., Moore, S., Gauci, V., Laiho, R., et al. (2014). 986 Contrasting vulnerability of drained tropical and high-latitude peatlands to fluvial loss of 987 stored carbon. Global Biogeochemical Cycles, 28(11), 1215–1234. https://doi.org/10.1002/2013GB004782 988 989 Fabricius, K. E., Langdon, C., Uthicke, S., Humphrey, C., Noonan, S., De'ath, G., et al. 990 (2011). Losers and winners in coral reefs acclimatized to elevated carbon dioxide 991 concentrations. *Nature Climate Change*, 1(3), 165–169. 992 https://doi.org/10.1038/nclimate1122 993 Fichot, C. G., & Benner, R. (2014). The fate of terrigenous dissolved organic carbon in a 994 river-influenced ocean margin. Global Biogeochemical Cycles, 28(3), 300–318. 995 https://doi.org/10.1002/2013GB004670 Follett, C. L., Repeta, D. J., Rothman, D. H., Xu, L., & Santinelli, C. (2014). Hidden cycle of 996 997 dissolved organic carbon in the deep ocean. Proceedings of the National Academy of 998 Sciences of the United States of America, 111(47), 16706–16711. 999 https://doi.org/10.1073/pnas.1407445111 1000 Gandois, L., Teisserenc, R., Cobb, A. R., Chieng, H. I., Lim, L. B. L., Kamariah, A. S., et al. 1001 (2014). Origin, composition, and transformation of dissolved organic matter in tropical

1002 peatlands. Geochimica et Cosmochimica Acta, 137, 35-47. 1003 https://doi.org/10.1016/j.gca.2014.03.012 1004 Gandois, L., Hoyt, A. M., Mounier, S., Le Roux, G., Harvey, C. F., Claustres, A., et al. 1005 (2020). From canals to the coast: dissolved organic matter and trace metal composition 1006 in rivers draining degraded tropical peatlands in Indonesia. *Biogeosciences*, 17(7), 1007 1897–1909. https://doi.org/10.5194/bg-17-1897-2020 1008 Gardner, T. A., Côté, I. M., Gill, J. A., Grant, A., & Watkinson, A. R. (2003). Long-term 1009 region-wide declines in Caribbean corals. Science, 301(5635), 958–960. 1010 https://doi.org/10.1126/science.1086050 Gordon, A. L., Huber, B. A., Metzger, E. J., Susanto, R. D., Hurlburt, H. E., & Adi, T. R. 1011 1012 (2012). South China Sea throughflow impact on the Indonesian throughflow. 1013 Geophysical Research Letters, 39(11), 1–7. https://doi.org/10.1029/2012GL052021 1014 Gran, G. (1952). Determination of the equivalence point in potentiometric titrations. Part II. 1015 The Analyst. https://doi.org/10.1039/AN9527700661 1016 Guenet, B., Danger, M., Abbadie, L., & Lacroix, G. (2010). Priming effect: bridging the gap 1017 between terrestrial and aquatic ecology. Ecology, 91(10), 2850–2861. 1018 https://doi.org/10.1890/09-1968.1 1019 Hamzah, F., Agustiadi, T., Susanto, R. D., Wei, Z., Guo, L., Cao, Z., & Dai, M. (2020). 1020 Dynamics of the Carbonate System in the Western Indonesian Seas During the 1021 Southeast Monsoon. *Journal of Geophysical Research: Oceans*, 125(1), 1–18. 1022 https://doi.org/10.1029/2018JC014912

1023 Helms, J. R., Stubbins, A., Ritchie, J. D., Minor, E. C., Kieber, D. J., & Mopper, K. (2008). 1024 Absorption spectral slopes and slope ratios as indicators of molecular weight, source, 1025 and photobleaching of chromophoric dissolved organic matter. Limnology and 1026 Oceanography, 53(3), 955–969. https://doi.org/10.4319/lo.2008.53.3.0955 1027 van Heuven, S., Pierrot, D., Rae, J. W. B., Lewis, E., & Wallace, D. W. R. (2011). MATLAB 1028 Program Developed for CO<sub>2</sub> System Calculations. ORNL/CDIAC-105b. ORNL/CDIAC-1029 105b. Carbon Dioxide Information Analysis Center, Oak Ridge National Laboratory, 1030 U.S. Department of Energy, Oak Ridge, Tennessee. 1031 https://doi.org/10.3334/CDIAC/otg.CO2SYS\_MATLAB\_v1.1 1032 Hodgkins, S. B., Richardson, C. J., Dommain, R., Wang, H., Glaser, P. H., Verbeke, B., et al. 1033 (2018). Tropical peatland carbon storage linked to global latitudinal trends in peat 1034 recalcitrance. Nature Communications, 9(1), 1–13. https://doi.org/10.1038/s41467-018-1035 06050-2 1036 Hoegh-Guldberg, O., Mumby, P. J., Hooten, A. J., Steneck, R. S., Greenfield, P., Gomez, E., 1037 et al. (2007). Coral Reefs Under Rapid Climate Change and Ocean Acidification. 1038 Science, 318(5857), 1737–1742. https://doi.org/10.1126/science.1152509 1039 Huang, D., Tun, K. P. P., Chou, L. M., & Todd, P. a. (2009). An inventory of zooxanthellate 1040 scleractinian corals in Singapore, including 33 new records. Raffles Bulletin of Zoology, 1041 22(22), 69–80. Retrieved from http://rmbr.nus.edu.sg/rbz/biblio/s22/s22rbz069-080.pdf 1042 Hughes, T. P., Kerry, J. T., Baird, A. H., Connolly, S. R., Dietzel, A., Eakin, C. M., et al. 1043 (2018). Global warming transforms coral reef assemblages. Nature, 556(7702), 492–

1044 496. https://doi.org/10.1038/s41586-018-0041-2 1045 Humborg, C., Geibel, M. C., Anderson, L. G., Björk, G., Mörth, C.-M., Sundbom, M., et al. 1046 (2017). Sea-air exchange patterns along the central and outer East Siberian Arctic Shelf 1047 as inferred from continuous CO<sub>2</sub>, stable isotope, and bulk chemistry measurements. 1048 Global Biogeochemical Cycles, 31(7), 1173–1191. 1049 https://doi.org/10.1002/2017GB005656 1050 Humphreys, M. P., Greatrix, F. M., Tynan, E., Achterberg, E. P., Griffiths, A. M., Fry, C. H., 1051 et al. (2016). Stable carbon isotopes of dissolved inorganic carbon for a zonal transect 1052 across the subpolar North Atlantic Ocean in summer 2014. Earth System Science Data, 1053 8(1), 221–233. https://doi.org/10.5194/essd-8-221-2016 1054 Januchowski-Hartley, F. A., Bauman, A. G., Morgan, K. M., Seah, J. C. L., Huang, D., & 1055 Todd, P. A. (2020). Accreting coral reefs in a highly urbanized environment. Coral 1056 Reefs, 39(3), 717–731. https://doi.org/10.1007/s00338-020-01953-3 1057 Kaiser, K., Benner, R., & Amon, R. M. W. (2017). The fate of terrigenous dissolved organic 1058 carbon on the Eurasian shelves and export to the North Atlantic. Journal of Geophysical 1059 Research: Oceans, 122(1), 4–22. https://doi.org/10.1002/2016JC012380 1060 Kitidis, V., Shutler, J. D., Ashton, I., Warren, M., Brown, I., Findlay, H., et al. (2019). Winter 1061 weather controls net influx of atmospheric CO<sub>2</sub> on the north-west European shelf. 1062 Scientific Reports, 9(1), 1–11. https://doi.org/10.1038/s41598-019-56363-5 1063 Kleypas, J. A., McManu, J. W., & Mene, L. A. B. (1999). Environmental limits to coral reef 1064 development: Where do we draw the line? American Zoologist, 39(1), 146–159.

1065 https://doi.org/10.1093/icb/39.1.146 Langdon, C. (2005). Effect of elevated pCO<sub>2</sub> on photosynthesis and calcification of corals 1066 and interactions with seasonal change in temperature/irradiance and nutrient enrichment. 1067 1068 Journal of Geophysical Research, 110(C9), C09S07. 1069 https://doi.org/10.1029/2004JC002576 1070 Letscher, R. T., Hansell, D. A., & Kadko, D. (2011). Rapid removal of terrigenous dissolved 1071 organic carbon over the Eurasian shelves of the Arctic Ocean. *Marine Chemistry*, 1072 123(1-4), 78-87. https://doi.org/10.1016/j.marchem.2010.10.002 1073 Lønborg, C., Calleja, M. L., Fabricius, K. E., Smith, J. N., & Achterberg, E. P. (2019). The 1074 Great Barrier Reef: A source of CO<sub>2</sub> to the atmosphere. *Marine Chemistry*, 210, 24–33. 1075 https://doi.org/10.1016/j.marchem.2019.02.003 1076 Manzello, D. P., Kleypas, J. A., Budd, D. A., Eakin, C. M., Glynn, P. W., & Langdon, C. 1077 (2008). Poorly cemented coral reefs of the eastern tropical Pacific: Possible insights into 1078 reef development in a high-CO<sub>2</sub> world. Proceedings of the National Academy of 1079 Sciences, 105(30), 10450–10455. https://doi.org/10.1073/pnas.0712167105 1080 Van Maren, D. S., & Gerritsen, H. (2012). Residual flow and tidal asymmetry in the 1081 Singapore Strait, with implications for resuspension and residual transport of sediment. 1082 Journal of Geophysical Research: Oceans, 117(4), 1–18. 1083 https://doi.org/10.1029/2011JC007615 1084 Martin, P., Cherukuru, N., Tan, A. S. Y., Sanwlani, N., Mujahid, A., & Müller, M. (2018). 1085 Distribution and cycling of terrigenous dissolved organic carbon in peatland-draining

1086 rivers and coastal waters of Sarawak, Borneo. *Biogeosciences*, 15(22), 6847–6865. 1087 https://doi.org/10.5194/bg-15-6847-2018 1088 Mayer, B., & Pohlmann, T. (2014). Simulation of organic pollutants: First step towards an 1089 adaptation to the malacca strait. Asian Journal of Water, Environment and Pollution, 1090 *11*(1), 75–86. 1091 Mayer, B., Rixen, T., & Pohlmann, T. (2018). The spatial and temporal variability of air-sea 1092 CO<sub>2</sub> fluxes and the effect of net coral reef calcification in the Indonesian Seas: A 1093 numerical sensitivity study. Frontiers in Marine Science, 5(APR), 1–19. 1094 https://doi.org/10.3389/fmars.2018.00116 1095 Medeiros, P. M., Seidel, M., Ward, N. D., Carpenter, E. J., Gomes, H. R., Niggemann, J., et 1096 al. (2015). Fate of the Amazon River dissolved organic matter in the tropical Atlantic 1097 Ocean. Global Biogeochemical Cycles, 29(5), 677-690. 1098 https://doi.org/10.1002/2015GB005115 1099 Medeiros, P. M., Seidel, M., Niggemann, J., Spencer, R. G. M., Hernes, P. J., Yager, P. L., et 1100 al. (2016). A novel molecular approach for tracing terrigenous dissolved organic matter 1101 into the deep ocean. Global Biogeochemical Cycles, 30(5), 689–699. 1102 https://doi.org/10.1002/2015GB005320 1103 Meybeck, M. (1982). Carbon, nitrogen, and phosphorus transport by world rivers. American 1104 Journal of Science, 282(4), 401–450. https://doi.org/10.2475/ajs.282.4.401 1105 Meyers-Schulte, K. J., & Hedges, J. I. (1986). Molecular evidence for a terrestrial component 1106 of organic matter dissolved in ocean water. *Nature*, 321(6065), 61–63.

1107 https://doi.org/10.1038/321061a0 1108 Miettinen, J., Shi, C., & Liew, S. C. (2016). Land cover distribution in the peatlands of 1109 Peninsular Malaysia, Sumatra and Borneo in 2015 with changes since 1990. Global 1110 Ecology and Conservation, 6, 67–78. https://doi.org/10.1016/j.gecco.2016.02.004 1111 Mollica, N. R., Guo, W., Cohen, A. L., Huang, K. F., Foster, G. L., Donald, H. K., & Solow, 1112 A. R. (2018). Ocean acidification affects coral growth by reducing skeletal density. 1113 Proceedings of the National Academy of Sciences of the United States of America, 1114 115(8), 1754–1759. https://doi.org/10.1073/pnas.1712806115 1115 Moore, S., Gauci, V., Evans, C. D., & Page, S. E. (2011). Fluvial organic carbon losses from 1116 a Bornean blackwater river. Biogeosciences, 8(4), 901–909. https://doi.org/10.5194/bg-1117 8-901-2011 1118 Moore, S., Evans, C. D., Page, S. E., Garnett, M. H., Jones, T. G., Freeman, C., et al. (2013). 1119 Deep instability of deforested tropical peatlands revealed by fluvial organic carbon 1120 fluxes. *Nature*, 493(7434), 660–663. https://doi.org/10.1038/nature11818 1121 Moran, M. A., & Zepp, R. G. (1997). Role of photoreactions in the formation of biologically 1122 labile compounds from dissolved organic matter. Limnology and Oceanography, 42(6), 1123 1307–1316. https://doi.org/10.4319/lo.1997.42.6.1307 1124 Moran, M. A., Sheldon, W. M., & Zepp, R. G. (2000). Carbon loss and optical property 1125 changes during long-term photochemical and biological degradation of estuarine 1126 dissolved organic matter. Limnology and Oceanography, 45(6), 1254–1264. https://doi.org/10.4319/lo.2000.45.6.1254 1127

1128 Morgan, K. M., Moynihan, M. A., Sanwlani, N., & Switzer, A. D. (2020). Light Limitation 1129 and Depth-Variable Sedimentation Drives Vertical Reef Compression on Turbid Coral 1130 Reefs. Frontiers in Marine Science, 7(November), 1–13. 1131 https://doi.org/10.3389/fmars.2020.571256 1132 Müller-Dum, D., Warneke, T., Rixen, T., Müller, M., Baum, A., Christodoulou, A., et al. 1133 (2019). Impact of peatlands on carbon dioxide (CO<sub>2</sub>) emissions from the Rajang River 1134 and Estuary, Malaysia. *Biogeosciences*, 16(1), 17–32. https://doi.org/10.5194/bg-16-17-1135 2019 1136 Müller, D., Warneke, T., Rixen, T., Müller, M., Jamahari, S., Denis, N., et al. (2015). Lateral 1137 carbon fluxes and CO<sub>2</sub> outgassing from a tropical peat-draining river. *Biogeosciences*, 1138 12(20), 5967–5979. https://doi.org/10.5194/bg-12-5967-2015 1139 van Nugteren, P., Moodley, L., Brummer, G. J., Heip, C. H. R., Herman, P. M. J., & 1140 Middelburg, J. J. (2009). Seafloor ecosystem functioning: The importance of organic 1141 matter priming. Marine Biology, 156(11), 2277–2287. https://doi.org/10.1007/s00227-1142 009-1255-5 1143 Opsahl, S., & Benner, R. (1997). Distribution and cycling of terrigenous dissolved organic 1144 matter in the ocean. Nature, 386(6624), 480–482. https://doi.org/10.1038/386480a0 1145 Opsahl, S., & Zepp, R. G. (2001). Photochemically-induced alteration of stable carbon 1146 isotope ratios (δ13C) in terrigeneous dissolved organic carbon. Geophysical Research 1147 Letters, 28(12), 2417–2420. https://doi.org/10.1029/2000GL012686 1148 Osburn, C. L., Morris, D. P., Thorn, K. A., & Moeller, R. E. (2001). Chemical and optical

1149 changes in freshwater dissolved organic matter exposed to solar radiation. Biogeochemistry, 54(3), 251–278. https://doi.org/10.1023/A:1010657428418 1150 1151 Page, S. E., Rieley, J. O., & Banks, C. J. (2011). Global and regional importance of the 1152 tropical peatland carbon pool. Global Change Biology, 17(2), 798–818. 1153 https://doi.org/10.1111/j.1365-2486.2010.02279.x 1154 Painter, S. C., Lapworth, D. J., Woodward, E. M. S., Kroeger, S., Evans, C. D., Mayor, D. J., 1155 & Sanders, R. J. (2018). Terrestrial dissolved organic matter distribution in the North 1156 Sea. Science of the Total Environment, 630, 630–647. 1157 https://doi.org/10.1016/j.scitotenv.2018.02.237 Perry, C. T., Alvarez-Filip, L., Graham, N. A. J., Mumby, P. J., Wilson, S. K., Kench, P. S., 1158 1159 et al. (2018). Loss of coral reef growth capacity to track future increases in sea level. 1160 Nature, 558(7710), 396–400. https://doi.org/10.1038/s41586-018-0194-z 1161 Pisapia, C., Hochberg, E. J., & Carpenter, R. (2019). Multi-Decadal Change in Reef-Scale 1162 Production and Calcification Associated With Recent Disturbances on a Lizard Island 1163 Reef Flat. Frontiers in Marine Science, 6(September), 1–10. 1164 https://doi.org/10.3389/fmars.2019.00575 1165 Rixen, T., Baum, A., Pohlmann, T., Balzer, W., Samiaji, J., & Jose, C. (2008). The Siak, a 1166 tropical black water river in central Sumatra on the verge of anoxia. *Biogeochemistry*, 1167 90(2), 129–140. https://doi.org/10.1007/s10533-008-9239-y 1168 Rixen, T., Baum, A., Wit, F., & Samiaji, J. (2016). Carbon leaching from tropical peat soils 1169 and consequences for carbon balances. Frontiers in Earth Science, 4(July).

1170 https://doi.org/10.3389/feart.2016.00074 1171 Roobaert, A., Laruelle, G. G., Landschützer, P., Gruber, N., Chou, L., & Regnier, P. (2019). 1172 The Spatiotemporal Dynamics of the Sources and Sinks of CO<sub>2</sub> in the Global Coastal 1173 Ocean. Global Biogeochemical Cycles, 2, 1–22. https://doi.org/10.1029/2019GB006239 1174 Ruf, C. S., Gleason, S., & McKague, D. S. (2019). Assessment of CYGNSS Wind Speed 1175 Retrieval Uncertainty. IEEE Journal of Selected Topics in Applied Earth Observations 1176 and Remote Sensing, 12(1), 87–97. https://doi.org/10.1109/JSTARS.2018.2825948 1177 Samanta, S., Dalai, T. K., Pattanaik, J. K., Rai, S. K., & Mazumdar, A. (2015). Dissolved 1178 inorganic carbon (DIC) and its  $\delta^{13}$ C in the Ganga (Hooghly) River estuary, India: 1179 Evidence of DIC generation via organic carbon degradation and carbonate dissolution. 1180 Geochimica et Cosmochimica Acta, 165, 226–248. 1181 https://doi.org/10.1016/j.gca.2015.05.040 Semiletov, I., Pipko, I., Gustafsson, Ö., Anderson, L. G., Sergienko, V., Pugach, S., et al. 1182 1183 (2016). Acidification of East Siberian Arctic Shelf waters through addition of freshwater 1184 and terrestrial carbon. *Nature Geoscience*, 9(5), 361–365. 1185 https://doi.org/10.1038/NEGO2695 1186 Shirokova, L. S., Chupakov, A. V., Zabelina, S. A., Neverova, N. V., Payandi-Rolland, D., 1187 Causserand, C., et al. (2019). Humic surface waters of frozen peat bogs (permafrost 1188 zone) are highly resistant to bio- and photodegradation. Biogeosciences, 16(12), 2511– 1189 2526. https://doi.org/10.5194/bg-16-2511-2019 1190 Siegel, H., Gerth, M., Stottmeister, I., Baum, A., & Samiaji, J. (2019). Remote Sensing of

1191 Coastal Discharge of SE Sumatra (Indonesia). In Remote Sensing of the Asian Seas (pp. 1192 359–376). Cham: Springer International Publishing. https://doi.org/10.1007/978-3-319-1193 94067-0 20 1194 Spencer, R. G. M., Stubbins, A., Hernes, P. J., Baker, A., Mopper, K., Aufdenkampe, A. K., 1195 et al. (2009). Photochemical degradation of dissolved organic matter and dissolved 1196 lignin phenols from the Congo River. Journal of Geophysical Research, 114(G03010). 1197 https://doi.org/10.1029/2009JG000968 1198 Stoltenberg, L., Schulz, K. G., Lantz, C. A., Cyronak, T., & Eyre, B. D. (2021). Late 1199 afternoon seasonal transition to dissolution in a coral reef: An early warning of a net 1200 dissolving ecosystem? Geophysical Research Letters. 1201 https://doi.org/10.1029/2020gl090811 1202 Stubbins, A., Mann, P. J., Powers, L., Bittar, T. B., Dittmar, T., McIntyre, C. P., et al. (2017). 1203 Low photolability of yedoma permafrost dissolved organic carbon. Journal of 1204 Geophysical Research: Biogeosciences, 122(1), 200–211. 1205 https://doi.org/10.1002/2016JG003688 1206 Tranvik, L. J., Downing, J. A., Cotner, J. B., Loiselle, S. A., Striegl, R. G., Ballatore, T. J., et 1207 al. (2009). Lakes and reservoirs as regulators of carbon cycling and climate. *Limnology* 1208 and Oceanography, 54(6part2), 2298-2314. 1209 https://doi.org/10.4319/lo.2009.54.6\_part\_2.2298 1210 UNEP-WCMC. (2010). Global distribution of warm-water coral reefs, compiled from 1211 multiple sources including the Millennium Coral Reef Mapping Project, Version 1.3.

1212 Wanninkhof, R. (1992). Relationship between wind speed and gas exchange over the ocean. 1213 Journal of Geophysical Research, 97(C5), 7373. https://doi.org/10.1029/92JC00188 1214 Wanninkhof, R., Asher, W. E., Ho, D. T., Sweeney, C., & McGillis, W. R. (2009). Advances 1215 in Quantifying Air-Sea Gas Exchange and Environmental Forcing. Annual Review of 1216 Marine Science, 1(1), 213–244. https://doi.org/10.1146/annurev.marine.010908.163742 1217 Ward, N. D., Keil, R. G., Medeiros, P. M., Brito, D. C., Cunha, A. C., Dittmar, T., et al. 1218 (2013). Degradation of terrestrially derived macromolecules in the Amazon River. 1219 *Nature Geoscience*, 6(7), 530–533. https://doi.org/10.1038/ngeo1817 1220 Weishaar, J. L., Aiken, G. R., Bergamaschi, B. A., Fram, M. S., Fujii, R., & Mopper, K. 1221 (2003). Evaluation of Specific Ultraviolet Absorbance as an Indicator of the Chemical 1222 Composition and Reactivity of Dissolved Organic Carbon. Environmental Science & 1223 Technology, 37(20), 4702–4708. https://doi.org/10.1021/es030360x 1224 Weiss, R. F. (1974). Carbon dioxide in water and seawater: the solubility of a non-ideal gas. 1225 Marine Chemistry, 2(3), 203–215. https://doi.org/10.1016/0304-4203(74)90015-2 1226 Welschmeyer, N. A. (1994). Fluorometric analysis of chlorophyll a in the presence of 1227 chlorophyll b and pheopigments. Limnology and Oceanography, 39(8), 1985–1992. https://doi.org/10.4319/lo.1994.39.8.1985 1228 1229 Wickland, K. P., Neff, J. C., & Aiken, G. R. (2007). Dissolved organic carbon in Alaskan 1230 boreal forest: Sources, chemical characteristics, and biodegradability. *Ecosystems*, 10(8), 1231 1323-1340. https://doi.org/10.1007/s10021-007-9101-4 Wit, F., Müller, D., Baum, A., Warneke, T., Pranowo, W. S., Müller, M., & Rixen, T. (2015). 1232

1233 The impact of disturbed peatlands on river outgassing in Southeast Asia. *Nature* 1234 Communications, 6, 10155. https://doi.org/10.1038/ncomms10155 1235 Wit, F., Rixen, T., Baum, A., Pranowo, W. S., & Hutahaean, A. A. (2018). The Invisible 1236 Carbon Footprint as a hidden impact of peatland degradation inducing marine carbonate 1237 dissolution in Sumatra, Indonesia. Scientific Reports, 8(1), 17403. 1238 https://doi.org/10.1038/s41598-018-35769-7 1239 Yupi, H. M., Inoue, T., Bathgate, J., & Putra, R. (2016). Concentrations, loads and yields of 1240 organic carbon from two tropical peat swamp forest streams in Riau province, Sumatra, 1241 Indonesia. Mires and Peat, 18, 1–15. https://doi.org/10.19189/MaP.2015.OMB.181 Zeebe, R. E., & Wolf-Gladrow, D. A. (2001). CO<sub>2</sub> in seawater: equilibrium, kinetics, 1242 1243 isotopes. Elsevier Oceanography Series. 1244 Zhou, Y., Martin, P., & Müller, M. (2019). Composition and cycling of dissolved organic 1245 matter from tropical peatlands of coastal Sarawak, Borneo, revealed by fluorescence 1246 spectroscopy and parallel factor analysis. *Biogeosciences*, 16(13), 2733–2749. 1247 https://doi.org/10.5194/bg-16-2733-2019 1248 Zhu, Z.-Y., Oakes, J., Eyre, B., Hao, Y., Sia, E. S. A., Jiang, S., et al. (2020). The 1249 nonconservative distribution pattern of organic matter in the Rajang, a tropical river with 1250 peatland in its estuary. Biogeosciences, 17(9), 2473–2485. https://doi.org/10.5194/bg-1251 17-2473-2020 1252 Zigah, P. K., McNichol, A. P., Xu, L., Johnson, C., Santinelli, C., Karl, D. M., & Repeta, D. 1253 J. (2017). Allochthonous sources and dynamic cycling of ocean dissolved organic

1254	carbon revealed by carbon isotopes. <i>Geophysical Research Letters</i> , 44(5), 2407–2415.
1255	https://doi.org/10.1002/2016GL071348
1256	