

# Hillslopes in Headwaters of Qinghai-Tibetan Plateau as Hotspots for Subsurface Dissolved Organic Carbon Processing during Permafrost Thaw

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## Abstract

Climate warming has accelerated thawing of northern permafrost, resulting in changes to the supply of dissolved organic carbon (DOC) to inland waters with uncertain fate. Extensive surface – groundwater interactions occur in alpine permafrost watersheds and likely influence DOC processing differently than systems with limited interactions. Here, we quantify and characterize DOC in waters collected from eight water types sampled across a small (25 km<sup>2</sup>) alpine (elevation 2960 to 4820 m a.s.l.) watershed in the Qinghai-Tibetan Plateau (QTP) containing variably degraded permafrost. Three types of water (thermokarst ponds, red mud gully and seepage-I) contained high DOC concentrations (5.2 to 22.6 mg/L, n=38), with C contributions predominantly from frozen soil meltwater. Spatial patterns of DOC in stream (0.3 to 4.8 mg L<sup>-1</sup>, n=41), and subsurface waters (0.4 to 3.8 mg/L, n=34), all contained frozen soil meltwater C as constrained by  $\delta^{18}\text{O}$  and electrical conductivity, reflecting surface – groundwater exchanges in the upper-, mid- and lower stretches of the watershed. Further, patterns of increasing DOC loss ( $\Delta\text{DOC}$ ) in subsurface waters with decreased proportions of protein-like organic matter and SUVA<sub>254</sub>, suggest subsurface microbial processing. Using previously established biodegradation DOC kinetics (0.06 d<sup>-1</sup>) from the QTP, the groundwater transit time is estimated to be between 6 and 20 days based on  $\Delta\text{DOC}$  changes of 32% and 74% for July and September, respectively. Mass balance of DOC inputs and export fluxes demonstrate nearly half of all DOC was lost in this small watershed, indicating hillslopes are hotspots for DOC processing, with subsurface environments playing a key role.

# Hillslopes in Headwaters of Qinghai-Tibetan Plateau as Hotspots for Subsurface Dissolved Organic Carbon Processing during Permafrost Thaw

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## Key Points

- Surface and subsurface waters in an alpine watershed of Qinghai-Tibetan Plateau contain permafrost sourced bioavailable DOC.
- Hillslopes in headwaters of QTP are hotspots for subsurface DOC processing before it reaches the main river during seasonal thaw.
- New constraint is placed on hillslope groundwater mean transit time (~6 to 20 days) from the DOC loss and the biodegradation kinetics.

## Abstract

Climate warming has accelerated thawing of northern permafrost, resulting in changes to the supply of dissolved organic carbon (DOC) to inland waters with uncertain fate. Extensive surface – groundwater interactions occur in alpine permafrost watersheds and likely influence DOC processing differently than systems with limited interactions. Here, we quantify and characterize DOC in waters collected from eight water types sampled across a small (25 km<sup>2</sup>) alpine (elevation 2960 to 4820 m a.s.l) watershed in the Qinghai-Tibetan Plateau (QTP) containing variably degraded permafrost. Three types of water (thermokarst ponds, red mud gully and seepage-I) contained high DOC concentrations (5.2 to 22.6 mg/L, n=38), with C contributions predominantly from frozen soil meltwater. Spatial patterns of DOC in stream (0.3 to 4.8 mg L<sup>-1</sup>, n=41), and subsurface waters (0.4 to 3.8 mg/L, n=34), all contained frozen soil meltwater C as constrained by  $\delta^{18}\text{O}$  and electrical conductivity, reflecting surface – groundwater exchanges in the upper-, mid- and lower stretches of the watershed. Further, patterns of increasing DOC loss ( $\Delta\text{DOC}$ ) in subsurface waters with decreased proportions of protein-like organic matter and  $\text{SUVA}_{254}$ , suggest subsurface microbial processing. Using previously established biodegradation DOC kinetics (0.06 d<sup>-1</sup>) from the QTP, the groundwater transit time is estimated to be between 6 and 20 days based on  $\Delta\text{DOC}$  changes of 32% and 74% for July and September, respectively. Mass balance of DOC inputs and export fluxes demonstrate nearly half of all DOC was lost in this small watershed, indicating hillslopes are hotspots for DOC processing, with subsurface environments playing a key role.

## Plain Language Summary

Climate warming leads to thawing of the northern permafrost that has increased the release of organic carbon, previously regarded as “stable”, into streams and rivers. Recent laboratory studies of Arctic permafrost show the rapid biodegradation of permafrost-derived organic carbon, but is it true? For this, we turn to a small, alpine watershed in the Qinghai-Tibetan Plateau with a gradient of permafrost degradation. First, we provide rare field evidence for widespread permafrost sourced organic carbon in a variety of water types based upon its optical properties. Secondly, we use stable isotopes and electrical conductivity as “conservative” tracers to compare with dissolved organic carbon that are “reactive” to estimate the loss of organic carbon from its

upgradient source to the downslope sampling location. The extent of the loss of DOC in subsurface environment is dependent on the travel time of the groundwater along the hillslope. The DOC originating from permafrost soil is quickly dispersed in the watershed, shedding light on previously poorly constrained surface water – groundwater interaction in such settings. Finally, a mass budget finds a large loss of organic carbon within the watershed. Therefore, hillslopes act as hotspots for permafrost-derived organic carbon processing.

## **Keywords**

Qinghai-Tibetan Plateau, Dissolved organic carbon, Permafrost, Alpine watershed, Groundwater transit time, Fluorescence spectroscopy

## **1 Introduction**

Northern permafrost, including 42% of the areal extent of Qinghai-Tibetan Plateau (QTP), is estimated to account for >20% of total global soil organic carbon (SOC) pool of  $1832 \times 10^{12}$  kg [Tarnocai *et al.*, 2009; Wang *et al.*, 2020]. Models assessing biogeochemical cycles of soil C have raised the concern that thaw and degradation may reverse northern permafrost regions from a current C sink to a net source, under future warming trajectories [McGuire *et al.*, 2018; Turetsky *et al.*, 2020; Wang *et al.*, 2020]. Thawed permafrost SOC enters pore waters as dissolved organic carbon (DOC), a proportion of which is exported to aquatic systems to become components of in-stream carbon cycling. About 15% of permafrost SOC may enter aquatic systems over the next 300 years under future warming scenarios [McGuire *et al.*, 2018]. In the high latitude Arctic permafrost region, CO<sub>2</sub> evasion from inland waters is estimated to account for between  $40 \times 10^9$  to  $84 \times 10^9$  kg C yr<sup>-1</sup> [McGuire *et al.*, 2009], but little is known about CO<sub>2</sub> evasion from inland waters in high altitude permafrost regions such as the QTP.

Streams and rivers are known hotspots for DOC processing [Raymond *et al.*, 2013]. This notion is further reinforced in a compilation of DOC losses during passages from terrestrial sources to sea, highlighting microbially mediated respiration of DOC in response to retention time scales [Catalan *et al.*, 2016]. Globally, streams and rivers are responsible for  $1.8 \times 10^{12}$  kg C yr<sup>-1</sup> out of the total of  $2.1 \times 10^{12}$  kg C yr<sup>-1</sup> CO<sub>2</sub> evasion from inland waters [Raymond *et al.*, 2013]. The total evasion has been revised upward to  $3.9 \times 10^{12}$  kg C yr<sup>-1</sup> [Drake *et al.*, 2018],

owing to more observational data. However, this flux is likely an underestimation. Due to lack of hydrologic data in small-sized headwater streams, it has been suggested that C evasion from small streams may be substantial but remain unaccounted for [Drake *et al.*, 2018; Raymond *et al.*, 2013]. Here, we describe ambiguities in DOC processing associated with permafrost thawing and degradation first in lateral flow dominated Arctic watersheds, then in alpine watersheds with extensive surface water – groundwater interactions along the hillslopes.

The apparently wide variability in DOC processing observed in the Arctic permafrost regions can be reconciled by considering the evolving nature of the SOC sources and time it takes for the SOC leachate to infiltrate and move laterally into the stream (SOC to porewater DOC, then subsequently DOC in streams). All of these processes are likely to alter under ongoing climate change. Arctic river waters sampled at large basin scales (8000 to 855,000 km<sup>2</sup>) have been shown to contain highly aromatic and less biologically labile DOC with young (390-1440 yr BP) radiocarbon age [Balcarczyk *et al.*, 2009; Guo and Macdonald, 2006; Guo *et al.*, 2007; Kawahigashi *et al.*, 2004]. This likely reflects inputs from modern active-layer soil SOC, as well as previously degraded Holocene permafrost SOC [Heslop *et al.*, 2019], plus further degradation along transport pathways due to long residence time of water in large basins [Mann *et al.*, 2015; Spencer *et al.*, 2015; Striegl *et al.*, 2005]. When permafrost SOC sources are ancient (> 20,000 yr BP) [Drake *et al.*, 2015] and terrestrial thaw process abrupt [Turetsky *et al.*, 2020], stream DOC can become more biologically labile, contributing significantly to in-stream C processing as evidenced by one third to one half of DOC losses in first-order tributaries of the Kolyma River basin [Mann *et al.*, 2015; Spencer *et al.*, 2015; Vonk *et al.*, 2013]. Field observations of soil C has indicated that half of SOC loss is attributable to the lateral flow passing into the aquatic system of the Eight Mile Lake watershed, though the further fate of the transported C is highly uncertain [Plaza *et al.*, 2019]. The need to understand how flow and water exchange regulate the quantity and quality of DOC reaching the stream is also clear from soil column experiments showing substantial (> 70%) C loss in lateral leachate with artificial precipitation [Zhang *et al.*, 2017]. Given the uncertainties associated with C cycling in small sized, lower order watersheds [Raymond and Spencer, 2015; Raymond *et al.*, 2013], it is useful to examine quantity and quality of DOC along the SOC to stream DOC transport pathways.

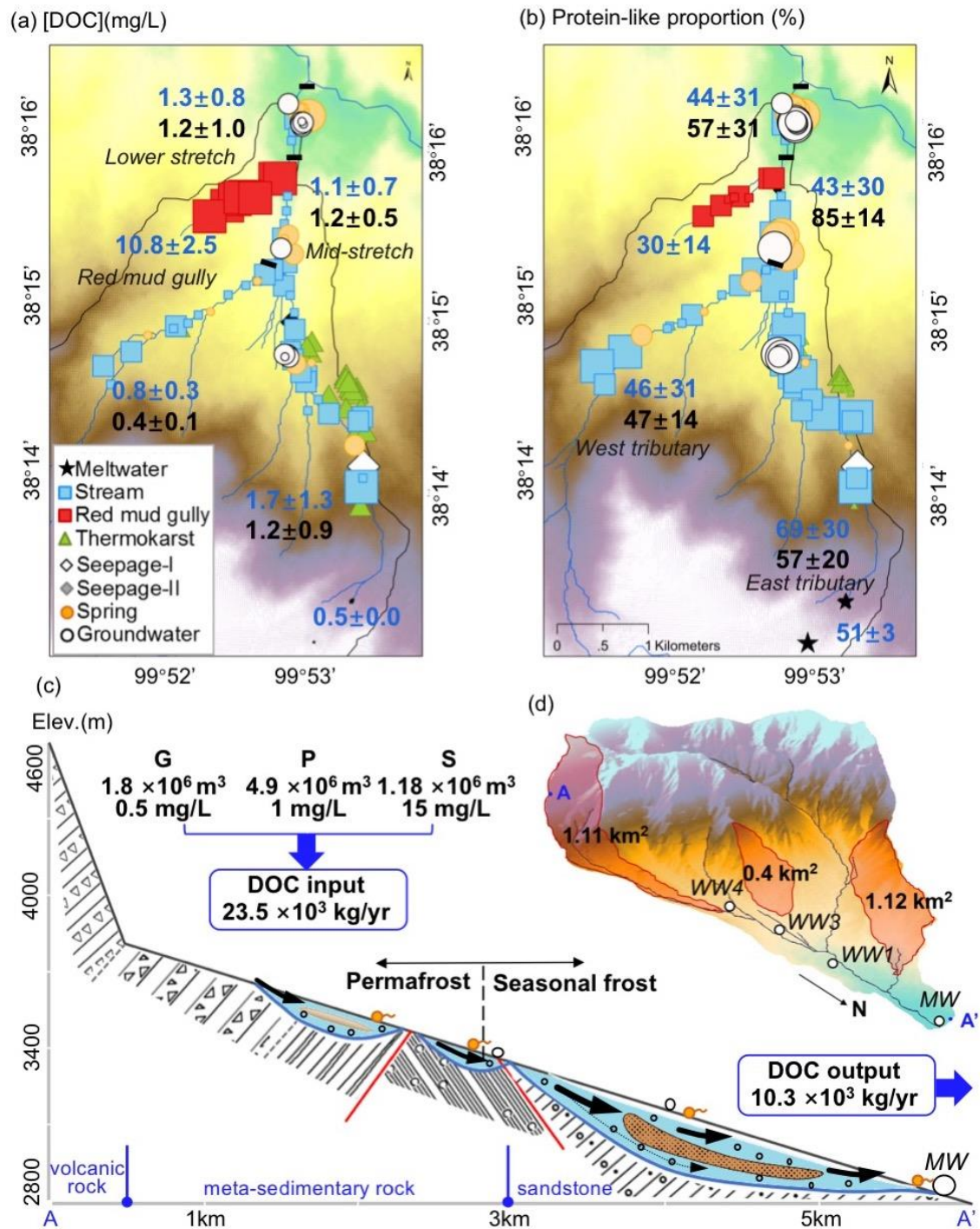
SOC transport pathways to streams are undergoing profound hydrological changes triggered by permafrost thawing [Mann *et al.*, 2015; O'Donnell *et al.*, 2012]. In Arctic permafrost

areas, there is an increasingly important role of groundwater linked with the increase of base flow and interaction between surface water and groundwater [Connolly *et al.*, 2020; Frey and McClelland, 2009; Vonk *et al.*, 2019]. Observations have identified accelerated input of older DOC exported as baseflow due to the deepening of flow path and increasing discharge [Aiken *et al.*, 2014; Barnes *et al.*, 2018; Neff *et al.*, 2006]. Warming is expected to reduce transit time, thus enhancing the propensity for lateral carbon transport along the terrestrial–aquatic continuum [Vonc *et al.*, 2019]. How changing hydrological processes will alter the in-stream C cycling in a warmer future is challenging to disentangle.

Assessing complex hydrological processes on catchment scales is a starting point to tackle such challenges. The Qinghai Tibetan Plateau (QTP) is an ideal location to study these processes as it contains the most extensive permafrost cover at the mid- and low- latitudes, but also because the spatially heterogenous degradation of permafrost following the topographical variation also influences hydrology [Cheng and Wu, 2007; Yang *et al.*, 2010]. Increasing precipitation and glacier melting have led to an expansion of water storage ( $12.1 \pm 0.6 \text{ Gt yr}^{-1}$ ) [Yi *et al.*, 2016], especially that of groundwater storage ( $5.01 \pm 1.59 \text{ Gt yr}^{-1}$ ) since 2003 [Zhang *et al.*, 2017]. Given such significant changes in QTP’s hydrological cycle, it is likely that groundwater flow is affected [Ge *et al.*, 2011, Yao *et al.*, 2017], yet few studies have examined such multifaceted changes and its biogeochemical implications. Recent advances in hillslope hydrology, especially the notion of groundwater mean transit time (MTT), defined as the time that water parcels spend between the time entering the unsaturated zone and the time flowing out of the aquifer [Benettin *et al.*, 2015; McDonnell *et al.*, 2010], offer a unique perspective to shed light on DOC processing in northern, alpine permafrost zones. Because MTT is found to range only from days to weeks [Luo and Jiao, 2019], challenging the long-held view that distribution of transit time ranging from years to decades in alpine hillslope hydrological condition [McGuire *et al.*, 2005], this short time scale makes it worthwhile to integrate groundwater into the terrestrial SOC – aquatic DOC continuum. How groundwater – surface water interaction in hillslopes of headwater regions of QTP influences the fate of SOC derived DOC through regulating groundwater transit time, and in turn, the carbon loss in headwaters, remains largely unexplored.

To illuminate the emerging and likely significant role that hillslope hydrological processes play in biogeochemical C cycling, this study seeks to shed light on DOC processing in a small ( $25 \text{ km}^2$ ), alpine (elevation 2960 to 4820 m a.s.l) watershed named Hulugou (HLGW)

located in the northeastern QTP (Fig. 1). The conservative tracers of  $^{18}\text{O}$  and electrical conductivity are used in an end-member analysis to ascertain the contribution of frost soil to multiple types of waters and to illustrate the extensive and spatially variable surface water – groundwater interactions in HLGW. The spatial and temporal variations in quantity and quality of dissolved organic matter (DOM) are interpreted to indicate rapid and variable transit time in part caused by changes in groundwater flow paths in response to the freeze-thaw cycle. Finally, how the new constraints on rapid groundwater transit time in the alpine watersheds of the QTP sets the stage for DOC loss in headwaters and its implications for hillslopes acting as DOC processing hotspots are discussed.



**Figure 1.** (a) Concentrations of DOC in eight types of water in HLGW, Qinghai-Tibetan Plateau, with legends in inserted panel. Small, medium and large symbol sizes indicate low ( $< 0.7 \text{ mg L}^{-1}$ ), medium ( $0.7\text{--}3.6 \text{ mg L}^{-1}$ ) and high ( $> 3.6 \text{ mg L}^{-1}$ ) [DOC] according to its tertile values. Numbers are mean value  $\pm$  one standard deviation for [DOC] in stream (blue) and subsurface water (black). (b) Proportions of protein-like component identified by PARAFAC modeling, with small, medium and large symbol sizes indicating low ( $< 27.4\%$ ), medium ( $27.4\text{--}62.4\%$ ) and high ( $> 62.4\%$ ) proportions according to tertile values. Numbers are mean value  $\pm$  one standard



deviation for the proportion of protein-like compounds in stream (blue) and subsurface water (black). Maps of Other DOM optical properties SUVA<sub>254</sub>, FI are available in Fig. S1. (c) A schematic diagram of hillslope hydrological process in HLGW for the cross-section A-A' (inset d, three areas of permafrost and seasonal frost in HLGW are shown, with monitoring wells marked in white circles). Water fluxes and [DOC] in three end members (G: glacier-snow melt, P: precipitation, S: frozen soil meltwater) contributing to DOC input are shown. The mass balance of DOC input and out fluxes suggests that about half of DOC is processed in HLGW.

## 2 Material and Methods

### 2.1 Study Area and Sample Collection

The study was conducted in Hulugou watershed (HLGW), upper Heihe basin, NE Qinghai-Tibetan Plateau (99°50' – 99°54' E, 38°12' – 38°17' N; 2960 to 4820 m a.s.l, 25 km<sup>2</sup>). The HLGW consists of three geomorphic units of glacier-snow covered mountain, rocky hills, and meadow steppe. Glacier, permafrost and seasonal frost ground have experienced degradation, albeit to different extent within the HLGW [Li *et al.*, 2014]. A first-order stream network with extensive surface water and groundwater exchanges [Chang *et al.*, 2018; Evans *et al.*, 2015] is superimposed on permafrost (3400 m to 4500 m a.s.l) with abundant thermokarst ponds, seasonal frost ground (2900 m to 3400 m a.s.l), and degraded permafrost (3450 m to 3600 m a.s.l) with erosional channels (Fig. 1).

Water samples (n=129) were collected from HLGW for a total of five times in July 2012 (n=22), April (n=11), July (n=28) and September 2013 (n=30), and September 2018 (n=38), respectively. Eight types of water were sampled, including glacier-snow melt, streams, a thermo-erosional red mud gully, thermokarst ponds, seepage-I, seepage-II, spring and groundwater, with detailed sampling locations described in Table 1. All types of water samples except groundwater were collected from 2 cm below the surface. The meltwater sample was collected in triplicate at the same time from melting glacier-snow in bare gravel zone, and analyzed individually, with results reported as mean values of the triplicate samples that represent this one type of water. The upper-stretch of the stream network in HLGW includes the east and the west tributaries originating at elevation above 4765 m. After the east and the west tributaries converge, the mid-stretch flows through the permafrost zone into the seasonal frozen soil zones, and becomes the lower-stretch after the red mud gully joins in (Fig. 1). Red mud gully is an erosional tributary; formed by water cutting deeply into the seasonal frost ground in the alpine meadow between

3300 m to 3000 m a.s.l, and thus classified separately as a type of water. Thawing of permafrost results in many small thermokarst ponds (diameter < 1 m mostly), observed in the alpine meadow at an altitude ranging from 3351 m to 3548 m close to the east tributary. The aquifer lithology and the installation of the observation wells are described in a previous study [Hu *et al.*, 2019]. Groundwater samples were collected from the wells shown in Fig. 1d with well depths being WW1–25 m, WW3–5 m, WW3–10 m, WW3–20 m, WW3–30 m and MW–30 m (Fig. 1d), after the wells have been primed until stable readings of temperature and EC were reached.

Cation and anion samples were filtered with 0.22  $\mu\text{m}$  nylon filters. Stable isotopes  $\text{H}_2^{18}\text{O}$  and  $\text{D}_2\text{O}$  samples were filtered with 0.22  $\mu\text{m}$  nylon filters into 2 ml glass bottles then immediately sealed without head space. Dissolved organic matter (DOM) samples were filtered using glass fiber syringe filters (Whatman GF/F style) with a 0.7  $\mu\text{m}$  nominal pore size into pre-combusted (at 450  $^{\circ}\text{C}$  for 6 hours) 22 ml brown glass bottles and crimp sealed with Teflon-faced silicone septa. All samples were filtered immediately after sampling and stored on ice at 4 $^{\circ}\text{C}$  for shipment to laboratory.

## 2.2 Measurements of field and chemical parameters, stable isotopes and DOM

Temperature (T), electrical conductivity (EC), pH and alkalinity were measured in the field. Stable isotopes ( $\delta\text{D}$  and  $\delta^{18}\text{O}$ ) were measured on a water isotope spectrometer analyzer (Model PICARRO L2130-I) at Pri-ecoco, Beijing, China (see Text S1 of Supporting Information). Major anions and cations of samples in 2012 and 2013, were measured by ion chromatography (IC-1000, Dionex) and Inductively Coupled Plasma Atomic Emission Spectroscopy (ICPAES, Teledyne Leeman, Prodigy), respectively. Major ions of sample in 2018 were measured on IC (Aquion, Dionex) with CS18 analytical column for anions, and CS16 column for cations, respectively. DOC concentrations of all water samples and soil extraction solutions (see Text S2 and Fig. S2 of Supporting Information on soil incubation) were measured on a Shimadzu TOC Analyzer. Detailed procedures are in Text S1 of Supporting Information.

Absorbance of UV-Visible chromophore DOM (CDOM) was measured by a UV-Visible Spectrophotometer (Agilent 8453) scanning from 200 to 800 nm (1 nm increments) to acquire absorbance for samples collected in 2012 and 2013. Prior to analyzing samples, a quartz cuvette filled with Milli-Q water was used to establish a daily baseline. Excitation emission matrices

(EEMs) were employed by scanning over an excitation (ex) range of 240 to 450 nm at 10 nm increments, and an emission (em) range of 350 to 550 nm at 2nm increments on a JY-Horiba Fluoromax-3 spectrofluorometer (Queens College, CUNY) with instrument-specific corrections, Raman normalization, inner filter correction, and cuvette blank subtraction applied. Optical properties of samples collected in 2018 were analyzed using a Horiba Aqualog spectrofluorometer (Southern University of Science and Technology) following the same procedure. EEMs were generated over excitation wavelengths between 246.58 to 827.57 nm in about 1.2-nm interval and emission wavelengths between 220 to 800 nm in 1-nm interval. All DOM measurements were completed within two weeks of sampling.

To correct for minor effect of light scattering by particles and microbubbles, wavelength-independent correction is conducted by subtracting the mean absorbance at range of 600 to 800 nm from all spectral absorbance values [Green and Blough, 1994]. Specific UV absorbance (SUVA) represents relative DOM aromaticity [Weishaar *et al.*, 2003], and SUVA at 254 nm (SUVA<sub>254</sub>) was applied in this study following previously reported methods [Weishaar *et al.*, 2003]. Fluorescence index (FI) is calculated as the ratio of intensities emitted at 470 nm and 520 nm at an excitation wavelength of 370nm, with lower value (~1.2) usually indicating for more terrestrial derived DOM and higher (~1.8) for more microbial DOM sources [Cory and McKnight, 2005; McKnight *et al.*, 2001]. Freshness index (BIX) is calculated as the ratio of emission intensity at 380 nm to the maximum intensity between 420 nm and 435 nm at an excitation wavelength of 310 nm [Parlanti *et al.*, 2000]. Higher value of BIX represents a higher proportion of fresh DOM [Parlanti *et al.*, 2000].

**Table 1.** Description of water types, electrical conductivity (EC), isotopic compositions, DOC concentration and optical properties in eight types of water in HLGW sampled between 2012-2018.

Description	Elevation range (m)	Types of water in HLGW	n	EC ( $\mu\text{S cm}^{-1}$ )	$\delta^{18}\text{O}$ (‰)	$\delta\text{D}$ (‰)	DOC ( $\text{mg L}^{-1}$ )	Protein proportion (%)	SUVA <sub>254</sub> ( $\text{L mgC}^{-1} \text{m}^{-1}$ )	FI	BIX
Melting glacier-snow in bare gravel zone	4100	Meltwater	1	179±1	-10.6±0.5	-64.8±3.8	0.5±0.0	51±3	0.33±0.17	1.94±0.04	0.83±0.02
Upper-, mid-, and lower-stretch *	3620-2920	Stream	41	355±141	-9.1±0.4	-52.1±4.6	1.3±1.1	56±33	1.09±0.63	1.62±0.16	0.73±0.14
Erosional tributary cutting through the seasonal frost zone	3200-3050	Red mud gully	10	2881±399	-6.3±0.4	-32.8±2.5	10.8±2.5	30±14	2.84±0.78	1.50±0.14	0.71±0.09
Seasonal frost ground in alpine meadow	3620-3350	Thermokarst ponds	26	334±123	-4.5±0.7	-24.8±5.3	14.3±3.3	24±11	3.75±0.48	1.37±0.09	0.59±0.05
Emerging thermokarst	3610	Seepage-I	2	322±25	-6.4±0.1	-37.1±0.6	7.1±1.0	45±22	3.16±0.35	1.29±0.04	0.58±0.03
Slow discharging spring	3410-3060	Seepage-II	4	501±181	-9.1±0.4	-52.2±2.0	1.5±0.8	43±39	1.16±0.80	1.60±0.16	0.61±0.12
Fast discharging spring	3570-2950	Spring	17	450±196	-8.7±0.3	-50.1±2.2	1.1±1.0	59±31	1.14±0.60	1.75±0.21	0.74±0.21
Six wells <sup>‡</sup>	3300-2970	Groundwater	13	573±61	-8.5±0.3	-48.0±2.1	1.0±0.4	62±24	0.88±0.45	1.82±0.14	1.12±0.38

\* Upper-stretch includes east and west tributaries and is in permafrost zone, mid-stretch is in seasonal frost zone, and the lower-stretch is in seasonal frost zone

<sup>‡</sup> Wells shown in Fig. 1d with depths being WW1–25 m, WW3–5 m, WW3–10 m, WW3–20 m, WW3–30 m and MW–30

## 2.3 Parallel Factor Analysis (PARAFAC) of DOM Fluorescence Spectra

Fluorescence spectra, obtained as EEMs, are used to quantify the contribution of fluorescent DOM components through PARAFAC modeling [Coble, 1996; Murphy *et al.*, 2013]. PARAFAC was conducted following the procedures described in a previous algorithm to quantify the protein-like fluorophores and humic-like substances of DOM [Murphy *et al.*, 2013; Stedmon and Bro, 2008]. Prior to outlier tests, the intensity of EEMs of samples in 2018 was linearly interpolated to match the emission ranging 250 to 450 nm at an interval of 10 nm and excitation ranging 300 to 550 nm at an interval of 2 nm. To avoid highly fluorescent samples exerting significant leverages on the PARAFAC model, normalization of each EEM to its integrated fluorescence was applied before model fitting [Murphy *et al.*, 2013]. Normalized fluorescence of each EEM was reversed to its raw fluorescence after model fitting.

We applied two PARAFAC models with two datasets. The first model includes all 100 water samples and the second adds 47 soil extracted samples to the first dataset of 100 water samples (see Section 2.7 on soil extraction). A series of three to six component- models were fitted to the dataset, with non-negativity constraint of  $10^{-8}$  applied to each. The split-half validation was achieved through splitting the data in half and modeling each half separately, fitting 20 models with random starts, and by inspection towards lowest residuals [Murphy *et al.*, 2013]. The PARAFAC analysis resolved a four-components model comprising the EEMs dataset, explaining 97.8% of the total variance. The comparisons of two model exports are shown in Figure S6, suggesting consistency with additional samples. Spectra loadings of both excitation and emission mode for each component were matched to the OpenFluor database (<https://openfluor.lablicate.com/of/measurement/1104>), and the description of each component was interpreted from matched compounds identified from previous studies with similarity >95% [Murphy *et al.*, 2014]. The identified four fluorescence components (C1 to C4) are ubiquitous and common in marine environments [Catala *et al.*, 2015; Wunsch *et al.*, 2018], as well as cryosphere such as Arctic surface waters [Gonçalvesaraujo *et al.*, 2016] and ice cores in Arctic Canada [Brogi *et al.*, 2018], and ice in the Antarctic seas [Stedmon *et al.*, 2011]. The C1 and C3 components have a broader emission spectrum and two extraction spectra peaks, traditionally referred to as humic-like components (Fig. S3). The C2 (ex: 270 nm; em: 304 nm) and C4 (ex: 290 nm; em: 338) components have narrower spectra with excitation and emission maxima

below 350 nm (Fig. S3), and are similar to the spectra of tyrosine and tryptophan, respectively [Murphy *et al.*, 2008]. The characterization of C2 and C4 represents amino-acids, free or bound in proteins compounds. The proportion of protein-like fluorophores is calculated as the sum of C2 and C4 intensities divided by the bulk intensity of the sample. Representative EEMs of different water types are enclosed in Supporting Information (Figs. S4 and S5).

## 2.4 End Member Analysis Based on Conservative Tracers

A three end-member mixing analysis constrained by conservative tracers  $\delta^{18}\text{O}$  and EC was used to calculate the fractions of glacier-snow ( $f_G$ ), precipitation ( $f_P$ ) and frozen soil melt water ( $f_S$ ) contributing to stream and subsurface waters in HLGW. The same three end members have been adopted in a hydrograph separation study of HLGW [Li *et al.*, 2014]. The analysis is based on the following assumptions: (1) the three water sources are the dominant sources over the ablation season and that any other sources are negligible; (2) shallow, organic layer of frozen soil is involved in lateral flow thus provides a signature needing representation.

$$f_G \times C_G^{18\text{O}} + f_P \times C_P^{18\text{O}} + f_S \times C_S^{18\text{O}} = C_i^{18\text{O}} \quad (1)$$

$$f_G \times C_G^{\text{EC}} + f_P \times C_P^{\text{EC}} + f_S \times C_S^{\text{EC}} = C_i^{\text{EC}} \quad (2)$$

$$f_G + f_P + f_S = 1 \quad (3)$$

where  $f$  represents the estimated fraction of a given endmember contributing to the specific sample  $i$ ; the subscripts  $G$ ,  $P$  and  $S$  represent the glacier-snow meltwater, precipitation and soil endmember, respectively;  $C^{18\text{O}}$  and  $C^{\text{EC}}$  represent the  $\delta^{18}\text{O}$  and EC of the sample specified in the subscript, respectively.

The  $\delta^{18}\text{O}$  and EC values to constrain the glacier-snow endmember composition rely on not only a meltwater sample collected in triplicate and analyzed individually in this study, but also two newly deposit snow samples collected in May and November of 2012 and one meltwater sample collected in July 2012 in the front of a glacier in HLGW [Li *et al.*, 2015] (Table 2). The elevation-weighted  $\delta^{18}\text{O}$  (-7.66‰) and the volume-weighted  $\delta^{18}\text{O}$  (-7.7‰) are comparable based on 65 rainwater samples collected weekly between June 28 to Sept 2, 2012 from 7 elevations (2960 to 4160 m a.s.l.) of the HLGW [Chang *et al.*, 2018]. The volume-weighted  $\delta^{18}\text{O}$  becomes more negative with higher elevation at a rate of 0.19‰ per 100 m in HLGW [Chang *et al.*, 2018]. The volume-weighted mean EC and  $\delta^{18}\text{O}$  are used to represent the precipitation endmember

(Table 2). In July 2013, pore waters were collected from piezometer installed to depths of 0.6 m to 1.5 m; groundwater was collected from monitoring well WW4 with a depth of 1 m. These 3 samples were regarded to represent frozen soil water [Li *et al.*, 2014; Ma *et al.*, 2017]. The EC and  $\delta^{18}\text{O}$  endmember compositions were averaged from these samples plus 87 samples collected underneath 7 soil profiles at elevation between 3620 to 2920 m analyzed similarly in Li *et al.* [Li *et al.*, 2014] (Table 2).

To estimate the uncertainty associated with the tracer-based end-member analysis, a classical Gaussian error propagation equation was employed [Genereux, 1998]. The calculation followed procedure described elsewhere [Chang *et al.*, 2018]. The fractions and associated uncertainties are reported in Table 2 and Table S1.

$$W_f = \left[ \left( \frac{\partial f}{\partial C_G^{18\text{O}}} W_G^{18\text{O}} \right)^2 + \left( \frac{\partial f}{\partial C_P^{18\text{O}}} W_P^{18\text{O}} \right)^2 + \left( \frac{\partial f}{\partial C_S^{18\text{O}}} W_S^{18\text{O}} \right)^2 + \left( \frac{\partial f}{\partial C_i^{18\text{O}}} W_i^{18\text{O}} \right)^2 + \left( \frac{\partial f}{\partial C_G^{\text{EC}}} W_G^{\text{EC}} \right)^2 + \left( \frac{\partial f}{\partial C_P^{\text{EC}}} W_P^{\text{EC}} \right)^2 + \left( \frac{\partial f}{\partial C_S^{\text{EC}}} W_S^{\text{EC}} \right)^2 + \left( \frac{\partial f}{\partial C_i^{\text{EC}}} W_i^{\text{EC}} \right)^2 \right]^{1/2} \quad (4)$$

where  $W$  represents the uncertainty in the variable specified in the subscript, i.e  $W_f$  represents the uncertainty of the contribution fraction for a given end member in a sample,  $W_G^{18\text{O}}$  represents the uncertainty of  $\delta^{18}\text{O}$  in glacier-snow endmember.

## 2.5 Estimation of DOC Loss: $\Delta\text{DOC}$

Because DOC is non-conservative, we first estimate an initial DOC ( $\text{DOC}_0$ ) through summation of DOC contributed from each endmember (equation 5).

$$\text{DOC}_0 = f_G \times \text{DOC}_G + f_P \times \text{DOC}_P + f_S \times \text{DOC}_S \quad (5)$$

The DOC concentration ( $0.5 \pm 0.02 \text{ mg L}^{-1}$ ) in meltwater at the highest elevation is taken to represent the glacier-snow endmember (Table 2). The DOC concentration ( $15 \pm 2.5 \text{ mg L}^{-1}$ ) of pore water samples collected by piezometers is taken to represent the soil endmember (Table 2). Though DOC of rainfall was not measured in HLGW, the volume-weighted mean DOC is  $0.9 \text{ mg L}^{-1}$  in three remote meteorological stations and  $1.1 \pm 0.5 \text{ mg L}^{-1}$  in Lhasa city of central QTP [Li *et al.*, 2017; Li *et al.*, 2018]. The DOC of  $1.0 \text{ mg L}^{-1}$  is taken to represent the precipitation endmember (Table 2).

The difference between initial DOC and measured DOC of a sample suggests the loss of carbon along the flow path of water to the sampling point. The DOC loss ( $\Delta\text{DOC}$ ) is calculated by subtracting measured DOC concentration ( $[\text{DOC}]$ ) of a given water sample from its initial DOC ( $\text{DOC}_0$ ) calculated above, where larger  $\Delta\text{DOC}$  corresponds to more loss of DOC during transport.

$$\Delta\text{DOC} = \text{DOC}_0 - [\text{DOC}] \quad (6)$$

**Table 2.** Mean and standard deviation (mean  $\pm$  std) of  $\delta^{18}\text{O}$  (‰) and electrical conductivity (EC,  $\mu\text{S}/\text{cm}$ ) used in computing end-member mixing fractions and their uncertainties ( $W$ ), and DOC (mg/L) in the three end members.

End-member	$\delta^{18}\text{O}$ (‰)				EC ( $\mu\text{S}/\text{cm}$ )				DOC (mg/L)
	mean $\pm$ std	n	$t^*$ (0.7)	$W^{18\text{o}}$ (0.7)	mean $\pm$ std	n	$t^*$ (0.7)	$W^{\text{EC}}$ (0.7)	mean $\pm$ std
Glacier-snow meltwater	$-14 \pm 3.9$	6	1	3.9	$160 \pm 27$	4	1	27	$0.5 \pm 0.02$
Precipitation	$-7.7 \pm 2.0$	65	0.85	1.7	$35 \pm 3.0$	65	0.85	2.5	1
Frozen soil meltwater	$-5.3 \pm 0.2$	3	1	0.2	$3487 \pm 542$	4 <sup>‡</sup>	1	542	$15 \pm 2.5$

\* The uncertainty of each tracer in a specific endmember is calculated as the standard deviation multiplying the  $t$  value under coefficient level at 0.7.

‡ The mean value of EC is averaged from three endmember samples and the average EC value of 87 soil samples from Li et al., [2004].

## 2.6 Estimation of Groundwater Mean Transit Time

First order kinetics have been widely used to describe biodegradation of DOC incubation experiments [Catala et al., 2015], allowing for calculation of the degradation rate constant ( $\lambda$ ) as in equations (7), where the  $\text{DOC}_0$  usually represents the starting point of the experiment.

$$\text{DOC} = \text{DOC}_0 \times e^{-\lambda t} \quad (7)$$

Because photodegradation is unlikely in subsurface environment, the biodegradation rate constant is assumed to regulate DOC degradation in groundwater. Re-arranging equation (7) to simulate the  $\Delta\text{DOC}$  change in groundwater observed at the outlet of HLGW in July and September result in equation (8), with the calculated time,  $t$ , regarded as representing groundwater mean transit time (MTT). The assumptions for the linkage between DOC loss behavior and transit time are (1) photodegradation is negligible in subsurface environment; (2)



the potential direct input from particle organic carbon along the pathway is excluded because particles would not be transported by groundwater; (3) physical removal of DOC due to retardation is expected to be fast, evenly distributed and hence not considered.

$$1 - \Delta DOC / DOC_0 = e^{-\lambda_{gw}t} \quad (8)$$

$$MTT \approx t = - \frac{\ln(1 - \Delta DOC / DOC_0)}{\lambda_{gw}} \quad (9)$$

where  $\lambda_{gw}$  represents DOC degradation constant in groundwater which has been corrected to observed groundwater temperature of 5 °C based on the Arrhenius equation [Catalan *et al.*, 2016] from the incubation experiment temperature of 20 °C.

## 2.7 Soil Incubation Experiment

A batch incubation experiment of SOC was initiated in the field to minimize sample storage artifacts. Four soil profiles were sampled at elevations from 2850 to 3600 m a.s.l and included seasonal frozen soil, thermokarst ponds and less degraded permafrost soil (Table S2). Approximately 5 g of soil was weighed and added to pre-combusted 20-ml glass bottles in triplicate, and filled with 15 – 19 ml Milli-Q water. Then the bottles were crimp sealed with Teflon-coated septa and aluminum cap right away. The bottles were stored in room temperature in dark before the supernatant was sampled at day 1, 3 and 40, sacrificing a bottle at a sampling time. The supernatant was filtered and measured for DOC concentrations, UV-visible absorbance and fluorescence spectra were performed within a week of sampling following the same protocol as in section 2.2.

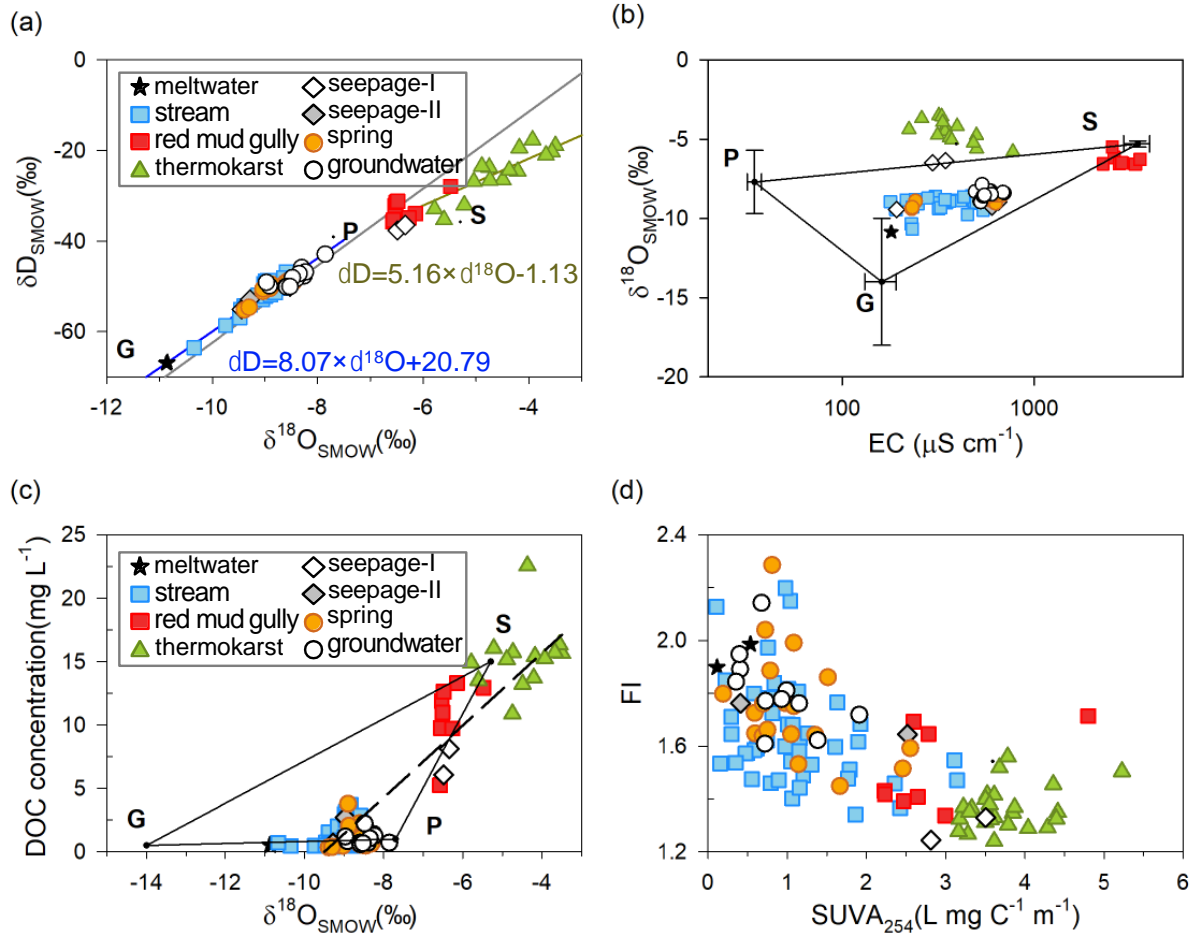
A second incubation experiment used a reactor approach similar to Drake *et al* to evaluate the biodegradation rate of soil derived DOC [Drake *et al.*, 2015] (Fig. S2). The shallow frost sample was collected by excavation to reach a depth of 50 cm within the active layer with thickness of 2 m nearby the well WW4. The sample was kept and transported on ice in a cooler to the lab. The details of incubation set up and sampling were described in Text S2 of supporting information.

### 3 Results

#### 3.1 Frozen Soil Meltwater Contributes to All Water Types

Stable isotope characteristics of eight types of water indicate the influence of not only glacier-snow meltwater (G) and precipitation (P) but also frozen soil melt water (S) endmembers (Figs. 2a and 2b). Stream and subsurface water (groundwater, spring and seepage-II) samples fall within the triangle defined by the G, P and S endmembers (Figs. 2a and 2b). The regression line of meltwater, stream, seepage-II, spring and groundwater ( $8.07 \times \delta^{18}\text{O} + 20.79$ ,  $r^2=0.92$ ,  $n=71$ , blue line in Fig. 2a) is similar to the local meteoric water line (LMWL:  $8.5 \times \delta^{18}\text{O} + 22.6$ , gray line in Fig. 2a) of HLGW [Ma *et al.*, 2017]. In contrast, thermokarst pond waters, two seepage-I samples and red mud gully waters exhibit more positive isotopic compositions and fall below the local meteoric water line (LMWL) on a different trend line ( $5.16 \times \delta^{18}\text{O} - 1.13$ ,  $r^2=0.83$ ,  $n=20$ , black line in Fig. 2a), indicating fractionation due to evaporation [Froehlich *et al.*, 2008]. The red mud gully waters are most similar to the frozen soil melt water endmember in compositions (Fig. 2b); and this is interpreted to indicate that red mud gully waters are derived from the frozen soil melt water. The two seepage-I samples (emerging thermokarst) represent mixing between roughly  $25\% \pm 1\%$  of the soil endmember, and  $75\% \pm 1\%$  of the precipitation endmember, respectively (Fig. 2b). The thermokarst pond waters are also likely a mixture of the S and P endmembers, although the  $\delta^{18}\text{O}$  values have become more positive due to evaporation. In summary, red mud gully, emerging thermokarst (seepage-I) and thermokarst ponds all have significant frozen soil meltwater contribution.

Using EC and  $\delta^{18}\text{O}$  to un-mix among the three endmembers for stream and subsurface water (groundwater, spring, and seepage-II) samples falling within the triangle, precipitation contributes on average about  $62\% \pm 7\%$ ,  $64\% \pm 4\%$ ,  $61\% \pm 4\%$  and  $63\%$  of the water in stream, groundwater, spring, and seepage-II, respectively (Fig. 2b). Glacier-snow meltwater ranked second for stream ( $28\% \pm 6\%$ ), groundwater ( $18\% \pm 4\%$ ) spring ( $24\% \pm 4\%$ ) and seepage-II ( $26\%$ ) contributions. Frozen soil meltwater contributed the least to the stream waters ( $10\% \pm 4\%$ ), but contributed  $11\%$ ,  $15\% \pm 5\%$ , and  $17\% \pm 2\%$  in seepage-II, spring, and groundwater samples, respectively.



**Figure 2.** (a) Stable isotope compositions ( $\delta^{18}O$  and  $\delta D$ ) of eight types of water in HLGW. Three endmembers: glacier-snow meltwater (G), precipitation (P) and frozen soil meltwater (S) are marked according to their compositions. Local meteoric water line (LMWL:  $8.5 \times \delta^{18}O + 22.6$ ) is shown as a gray line [Ma et al., 2017]. (b) The  $\delta^{18}O$  vs electrical conductivity (EC, log scale) of eight types of water samples from HLGW. The triangle outlines the three end members with the error bar representing standard deviation for the two tracers. Uncertainties in  $\delta^{18}O$  and EC of the three endmembers are reported in Table 2. (c) The  $\delta^{18}O$  vs DOC concentrations for all eight types of water within HLGW are shown with a positive correlation ( $p < 0.01$ ;  $n = 103$ ). The triangle outlines the three end members according to their  $\delta^{18}O$  and DOC concentrations. (d) The  $SUVA_{254}$  vs FI for all eight types of water with a negative correlation ( $r = -0.59$ ,  $p = 0.01$ ).

### 3.2 Frozen Soil Thaw Supplies the Majority of DOM to Surface and Subsurface Waters

Several lines of evidence suggest that DOM in surface water has been influenced by DOM from thawing of seasonal frost and/or permafrost. Three types of water show high DOC levels (Table 1): thermokarst ponds ( $14.3 \pm 3.3 mg L^{-1}$ ), red mud gully ( $10.8 \pm 2.5 mg L^{-1}$ ) and seepage-I ( $7.1 \pm 1.4 mg L^{-1}$ ). The elevated DOC levels of these three types of water are consistent

with the large contribution of frozen soil melt water endmember (Fig. 2b). These waters also show the most positive  $\delta^{18}\text{O}$ , with a correlation with DOC concentrations (Fig. 2c). Because DOC concentrations in glacier-snow melt ( $0.5 \text{ mg L}^{-1}$ ) and precipitation ( $1 \text{ mg L}^{-1}$ ) endmembers are low, this means that DOC in the stream ( $1.3 \pm 1.1 \text{ mg L}^{-1}$ , spring ( $1.1 \pm 1.1 \text{ mg L}^{-1}$ ), groundwater ( $1.1 \pm 0.4 \text{ mg L}^{-1}$ ) and seepage-II ( $1.5 \pm 0.8 \text{ mg L}^{-1}$ ) must include a source from thawing of frozen soil, consistent with the water source analysis results above. When the DOC concentrations of all samples are divided according to the two tertile values, with one third of samples each belonging to low, medium and high levels equally (Fig. 1a), stream waters ( $n = 29$ ) with low DOC level ( $0.7 \pm 0.3 \text{ mg L}^{-1}$ ) display the most negative  $\delta^{18}\text{O}$  values of  $-9.20 \pm 0.48 \text{ ‰}$  while stream waters ( $n = 12$ ) with medium DOC level ( $2.9 \pm 0.7 \text{ mg L}^{-1}$ ) show in-between  $\delta^{18}\text{O}$  of  $-8.82 \pm 0.14 \text{ ‰}$ , indicating input of frozen soil-derived DOC to streams.

Further support for the importance of thawing supplied DOM in the watershed is based on characterization of the quality of DOM by UV-visible and fluorescence spectroscopy because of its ability to absorb light and fluoresce. The optical properties including  $\text{SUVA}_{254}$ , FI and BIX all point to various degrees of influence by DOM from thawing of seasonal frost and/or permafrost soil. Thermokarst, red mud gully and seepage-I (emerging thermokarst) waters characterized by elevated DOC concentration display the highest  $\text{SUVA}_{254}$  and the lowest FI values (Fig. 2d and Table 1). This indicates overwhelming influence by terrestrial plant-soil sourced DOM with high aromaticity from organic matter produced some time ago. The glacier-snow meltwater is the least likely to be influenced by any DOM from frozen soil melt, and thus shows the lowest  $\text{SUVA}_{254}$  ( $0.33 \pm 0.17 \text{ L mgC}^{-1} \text{ m}^{-1}$ ) and the highest FI ( $1.94 \pm 0.04$ ). Stream, spring and groundwater display  $\text{SUVA}_{254}$  and FI values between the aforementioned two DOM optical “endmembers” (Fig. 2d). Subsurface waters (spring and groundwater) show higher BIX and FI values than stream water, indicating recent microbially processed DOM [Parlanti *et al.*, 2000], consistent with the interaction with microbes in subsurface environment. The BIX values of meltwater, groundwater, and most spring water lie above the median BIX value of 0.69 of the all eight types of water, while those of most stream, red mud gully, seepage-I and thermokarst pond waters are below (Fig. S6).

### 3.3 Differences in DOM Quality in Surface and Subsurface Waters Support Subsurface Processing

The DOM of meltwater collected at the origin of the east tributary displays low SUVA<sub>254</sub> of  $0.33 \pm 0.17 \text{ L mg C}^{-1} \text{ m}^{-1}$  and  $51 \pm 3\%$  of protein-like fluorophores based on a four-component PARAFAC model quantifying fluorescent DOM composition (Figs. 1b and Table 1). These values are comparable to that of glacier-snow meltwater ( $n=2$ ) in southwest QTP, with SUVA<sub>254</sub> of  $0.60 \text{ L mg C}^{-1} \text{ m}^{-1}$  and  $>50\%$  of amino-acids [Spencer *et al.*, 2014; Xu *et al.*, 2013]. Although the meltwater can be a source of bioavailable DOC based on FT-ICR-MS analysis with high proportions of protein-like fluorophores [Feng *et al.*, 2016], its DOC concentration is also the lowest among the eight types of water. Thus, there must be additional DOC with such optical properties from other sources. Three types of water with significant S contribution, including thermokarst, red mud gully and seepage-I, contain high DOC with substantial (though variable) proportions of protein-like fluorophores of  $24 \pm 11\%$  ( $n=12$ ),  $30 \pm 14\%$  ( $n=8$ ), and  $45 \pm 22\%$  ( $n=2$ ), respectively (Table 3). The permafrost SOC fingerprints evident in these three types of water are also observed in stream, spring and groundwater with DOM containing substantial though spatially variable proportions of protein-like fluorophores (Fig. 1b), intermediate SUVA<sub>254</sub> of  $\sim 1 \text{ L mgC}^{-1} \text{ m}^{-1}$  and medium-to-high FI of  $> 1.6$  (Fig. S6).

Several lines of evidence support that differences in water – permafrost soil hydrological interactions in the upper-, mid-, and lower-stretches of the HLGW lead to spatial variations in DOM quality in surface and subsurface waters. In the upper-stretch, the quantity and quality of DOC in surface water exhibit notable differences between the east and the west tributaries. Concentrations of DOC and proportions of protein-like components are on average higher in the east tributary ( $1.7 \pm 1.3 \text{ mg L}^{-1}$ ,  $69 \pm 30\%$ ) that drains two areas of permafrost and seasonal frost areas than those in the west tributary ( $0.8 \pm 0.3 \text{ mg L}^{-1}$ ,  $46 \pm 31\%$ ) that drains only one smaller area of permafrost (Fig. 1d and Table 3). Subsurface waters of the upper stretch display lower DOC concentration than surface water (Fig. 1a), with slightly less protein-like proportions and higher SUVA<sub>254</sub> (Fig. 1b and Table 3).

From the mid-stretch to the outlet of the HLGW, the quality of stream DOC reflects mixing of upstream waters from the east and west tributaries, and after convergence of red mud gully, with this additional input (Fig. 1a and Table 1). The comparable subsurface water DOC

mean concentrations between the upper-stretch east tributary ( $1.2 \pm 0.9 \text{ mg L}^{-1}$ ) and lower- ( $1.2 \pm 1.0 \text{ mg L}^{-1}$ ) stretches (Table 3) suggests that a groundwater flow path may connect the highest elevation area in the east with the lowest elevation area (Figs. 1a and 1b). The mid-stretch subsurface water may have been influenced more by a cluster of thermokarst ponds nearby because the proportion of protein-like fluorophores ( $85 \pm 14 \%$ ,  $n=4$ ) in four subsurface water samples from the mid-stretch nearly double that of stream water, with higher  $\text{SUVA}_{254}$  values as well (Fig. 1b and Table 3). This may be attributed to less photodegradation of permafrost sourced DOM in subsurface environment than in surface water.

In each stretch of the HLGW stream network, the mean value of FI in subsurface waters is substantially higher than that in the surface waters (Table 3), indicating more microbially processed DOM in subsurface environment, except for the mid-stretch where subsurface processing may be weak. When protein-like proportion is divided into three groups again according to the two tertile values, all of 12 groundwater samples, and 10 out of 13 spring samples belong to medium and high levels (Fig. 1b). The spring waters also have the highest bulk fluorescence intensity averaging  $3.85 \pm 6.58 \text{ RU}$  (Dataset S1). Taken together, the differences in DOM quality in surface and subsurface waters, along with the spatially variable distribution of optical properties of DOM, suggest that subsurface environment actively participate in DOM processing in headwaters of the QTP, with microbially driven biodegradation likely being important (see section 4.1).

**Table 3.** DOC concentrations, optical properties of DOM, proportions of glacier-snow ( $f_G$ ), precipitation ( $f_P$ ) and soil water ( $f_S$ ) contributing to stream and subsurface waters and associated DOC loss for the upper, mid and lower stretches of HLGW.

Category	Type*	n	DOC (mg/L)	Protein proportion (%)	$\text{SUVA}_{254}$ ( $\text{L mg}^{-1} \text{C}^{-1} \text{m}^{-1}$ )	FI	$f_G$	$f_P$	$f_S$	$\text{DOC}_0$ (mg/L)	$\Delta\text{DOC}$ (mg/L)
Upper stretch: east tributary (3620-3190 m)	surface	21	$1.7 \pm 1.3$	$69 \pm 30\%$	$1.18 \pm 0.60$	$1.60 \pm 0.14$	$29 \pm 9\%$	$62 \pm 9\%$	$10 \pm 4\%$	$2.2 \pm 0.6$	$1.2 \pm 0.8$
	subsurface	9	$1.2 \pm 0.9$	$57 \pm 20\%$	$1.38 \pm 0.73$	$1.71 \pm 0.11$	$19 \pm 2\%$	$64 \pm 4\%$	$17 \pm 2\%$	$3.3 \pm 0.3$	$2.3 \pm 0.5$
Upper stretch: west tributary (3500-3180 m)	surface	13	$0.8 \pm 0.3$	$46 \pm 31\%$	$1.12 \pm 0.75$	$1.62 \pm 0.20$	$27 \pm 3\%$	$65 \pm 3\%$	$8 \pm 2\%$	$2.0 \pm 0.3$	$1.2 \pm 0.5$
	subsurface	3	$0.4 \pm 0.0$	$47 \pm 14\%$	$1.00 \pm 0.37$	$1.85 \pm 0.15$	$27 \pm 4\%$	$59 \pm 5\%$	$13 \pm 6\%$	$2.7 \pm 0.8$	$2.3 \pm 0.8$
Mid-stretch (3140-3040 m)	surface	8	$1.1 \pm 0.7$	$43 \pm 30\%$	$0.77 \pm 0.24$	$1.66 \pm 0.10$	$27 \pm 3\%$	$58 \pm 3\%$	$16 \pm 2\%$	$3.0 \pm 0.3$	$1.9 \pm 0.5$
	subsurface	4	$1.2 \pm 0.5$	$85 \pm 14\%$	$1.00 \pm 0.16$	$1.69 \pm 0.18$	$22 \pm 3\%$	$64 \pm 3\%$	$14 \pm 5\%$	$2.9 \pm 0.8$	$1.7 \pm 0.9$
	surface	3	$1.3 \pm 0.8$	$44 \pm 31\%$	$0.76 \pm 0.21$	$1.67 \pm 0.12$	$26 \pm 2\%$	$60 \pm 5\%$	$14 \pm 3\%$	$2.8 \pm 0.4$	$2.4 \pm 0.1$

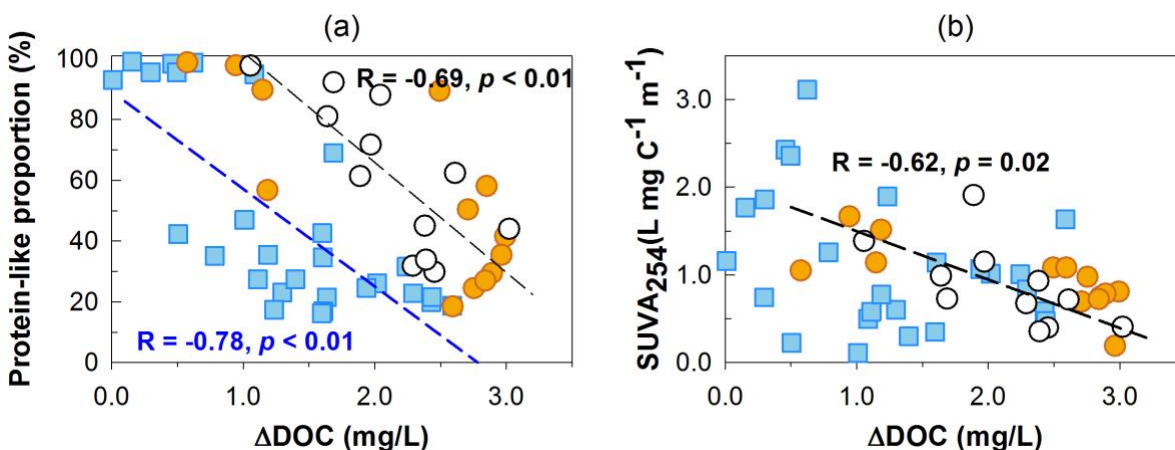
Lower stretch (3000-2940 m)	subsurface	13	1.2±1.0	57±31%	0.81±0.40	1.84±0.21	22±4%	62±4%	15±3%	3.1±0.5	2.2±0.7
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\*Note. Surface represents stream and subsurface includes spring and groundwater. Data are reported as averages  $\pm$  standard deviation for each category.

### 3.4 Concurrent DOC Quantity and Quality Changes in Stream and Subsurface Waters

The loss of DOC ( $\Delta$ DOC) in stream and subsurface waters, calculated by subtracting the measured DOC concentration from an estimated initial DOC concentration based on mixing of three-endmembers, represents DOC consumption along the flow paths in HLGW. In subsurface waters where the effect of photodegradation is minimal, simultaneous decreases in the proportions of protein-like fluorophores and SUVA<sub>254</sub> values are observed with increasing  $\Delta$ DOC (Fig. 3). For the stream waters, only the proportion of protein-like fluorophores is correlated with  $\Delta$ DOC, not SUVA<sub>254</sub> (Fig. 3). If only the upper stretches of the stream network in HLGW are considered, the correlations between the proportions of protein-like fluorophores and  $\Delta$ DOC improve in the about 2-km long east ( $r = -0.73$ ,  $p = 0.002$ ) and west ( $r = -0.78$ ,  $p = 0.003$ ) tributaries, respectively. The lack of correlation between stream water SUVA<sub>254</sub> and  $\Delta$ DOC (Fig. 3) is likely due to stronger effect of photodegradation in surface water than in groundwater due to aromatic C as light absorbing chromophores [Ward and Cory, 2016].

The concurrent changes in DOC quantity and quality are observed in the batch experiments of four types of frozen soils collected from the HLGW. Consistent with the concurrent changes in DOM quantity and quality in HLGW waters (Fig. 3), anoxic incubation of four types of soil reveals utilization of protein-like components and aromatic carbon. Decrease of SUVA<sub>254</sub> from  $> 4$  to about  $1 \text{ L mg C}^{-1} \text{ m}^{-1}$  and halving of protein-like fluorescent DOM is found in 8 soil samples collected from the active layers of permafrost, the degraded permafrost and thermokarst soils (Table S2). However, the soil incubations are compounded by simultaneous release of DOM from SOC mobilization, and adsorption may also contribute to DOC loss, making quantitative assessment impossible at this time, with details in Supporting Information. When the first-order kinetic equation was applied to fit the SOC biodegradation curves for the decrease in DOC in the second incubation experiment of the frost soil in the reactor, the biodegradation constant of SOC ( $\lambda_{\text{SOC}}$ ) is estimated to be  $0.32 \text{ d}^{-1}$  at  $20^\circ\text{C}$ , or a half-time of 5 hours (Fig. S7).



**Figure 3.** (a) The proportion of protein-like component vs DOC loss ( $\Delta\text{DOC}$ ) in surface (blue) and subsurface (white and orange) water. (b)  $\text{SUVA}_{254}$  vs DOC loss ( $\Delta\text{DOC}$ ) in surface (blue) and subsurface (white and orange) water. The blue and black dash lines represent the regression lines for stream and groundwater, respectively, with correlation coefficients marked next to the lines.

## 4 Discussion

### 4.1 Biodegradation Responsible for DOC Processing in Subsurface Environment

$\Delta\text{DOC}$  that represents DOC loss is estimated to evaluate to what extent DOM in HLGW is processed, based on the constraints of end-member analysis. Although glacier-snow and precipitation endmembers exhibit substantial uncertainties in their compositions (Table 2) due to altitudinal and temporal effects on isotopic compositions [Chang *et al.*, 2018; Li *et al.*, 2015], their effect on uncertainties of  $\Delta\text{DOC}$  estimation is insignificant due to their low DOC content. In comparison, because the frozen soil endmember contains much more DOC than the glacier-snow and precipitation endmembers do, the uncertainty in  $\Delta\text{DOC}$  estimation is mostly driven by the very small uncertainty of the soil endmember, and fortunately, is small (Table S1).

Both photodegradation and biodegradation can be important for mineralization of permafrost sourced DOC in high-altitude surface waters due to availability of sunlight [Wang *et al.*, 2018]. However, subsurface DOM is much less likely to undergo photo-oxidation or mineralization by sunlight, the DOC loss is therefore considered to mostly reflect biodegradation. The concurrent decreases of protein-like fluorophores and  $\text{SUVA}_{254}$  with increasing DOC loss (Fig. 3) support the notion that aromatic DOM with high protein-like fluorophores, most likely



sourced from thawing of frozen soil, is subject to biodegradation in subsurface environment of the HLGW. Further, the DOM quality changes in batch soil incubation support the utilization of protein-like and aromatic C. While we cannot entirely rule out the possibility of photodegradation due to frequent surface and subsurface water exchanges in HLGW, the transit time of subsurface water should be longer than that of stream water, allowing for microbial processing time of DOM in subsurface environment. The longer it takes for water to flow through subsurface, the higher the DOC loss by microbial activities will be along this flow path. We take advantage of this linkage to calculate MTT of groundwater using its  $\Delta\text{DOC}$  obtained at the outlet of HLGW as discussed in 4.2 next. The MTT reflects the flow pathways and water dynamics of the whole system [McGuire *et al.*, 2005].

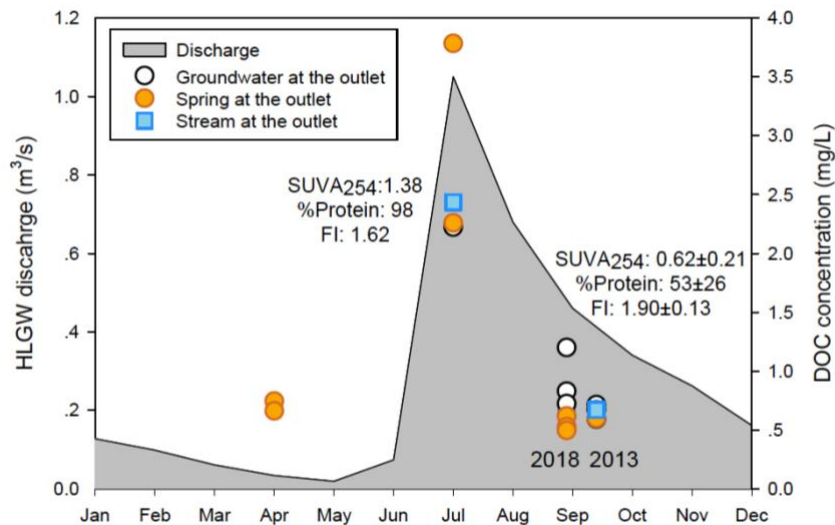
#### 4.2 Groundwater Mean Transit Time (MTT) in Response to Freeze-Thaw Cycles

In the Arctic permafrost region, groundwater discharge is driven by soil freeze – thaw processes with long water retention time [Ameli *et al.*, 2017; Ireson *et al.*, 2013; Walvoord and Striegl, 2007] and horizontal flow path [Aiken *et al.*, 2014; O'Donnell *et al.*, 2012]. This longer retention time explains the variable but usually low percentages of protein-like fluorophores: 0.3–22% in eight streams extending 250 km at Yenisei basin [Kawahigashi *et al.*, 2004]; 1–12% over 500 km at Yukon basin and its small tributaries [O'Donnell *et al.*, 2010; Wickland *et al.*, 2012]. HLGW is small (25 km<sup>2</sup>) and alpine (11° topographical gradient) so the groundwater MTT is expected to be short though not previously quantified. Only a handful of studies have attempted to quantify mean transit time in hillslopes. MTT of days to weeks (10 – 25 days) has been obtained for storm events, based on hydrometric and isotopic tracer approach in an alpine watershed of Oregon [McGuire and McDonnell, 2010]. Very short transit time of 2 to 12 days is observed for a mountain spring in Hong Kong constrained by radium (Ra) and radon (Rn) isotopes along a steep slope [Luo and Jiao, 2019].

In the following, we first discuss qualitative evidence for MTT of groundwater in HLGW responding to the seasonal freeze-thaw cycles followed by an attempt to quantify the MTT in low and high discharge periods.

First, DOC concentrations of groundwater and spring collected at the same locations close to the outlet of HLGW reach a maximum in July during peak discharge, higher than those

in September when discharge is much less (Fig. 4). The  $\Delta\text{DOC}$  of these subsurface waters are  $1.0 \pm 0.1 \text{ mg L}^{-1}$  in July and  $2.5 \pm 0.4 \text{ mg L}^{-1}$  in September, respectively, suggesting lower loss in July than in September. This contrast is neither attributable to a difference in  $\text{DOC}_0$  (July:  $3.2 \pm 0.04 \text{ mg L}^{-1}$ ; September:  $3.2 \pm 0.2 \text{ mg L}^{-1}$ ) nor to water temperature ( $5^\circ\text{C}$ ). It is unlikely due to adsorption alone because DOM adsorption is rapid, and usually reaches equilibrium within a few minutes to hours [Gu et al., 1994; Kalbitz and Wennrich, 1998]. Further, retardation of DOM due to sorption only “delays” the arrival of influent DOC by 1.5 to 3 pore volumes in sandy to clayey columns [Li and Shuman, 1997]. Therefore, this retardation of DOC transport alone is unlikely to account for large changes in DOC concentrations at the outlet over 2 months (Fig. 4).



**Figure 4.** Monthly average stream discharge (left y-axis) in 2013 displayed in gray shade recorded at the gauging station (2960 m a.s.l) at the outlet of HLGW. Concentrations of DOC (right y-axis) in surface water (blue square) and springs (orange circles) and groundwater (white circles) from a monitoring well (MW – 30 m depth in Fig. 1d), all close to the gauging station, are higher in July than in September. Numbers are values of  $\text{SUVA}_{254}$  ( $\text{L mg C}^{-1} \text{ m}^{-1}$ ), proportion of protein-like compound (%) and FI for groundwater from the MW.

Second, the protein-like proportion and  $\text{SUVA}_{254}$  of all subsurface waters at the outlet are higher in July ( $98 \pm 0.5\%$ ,  $1.25 \pm 0.41 \text{ L mg C}^{-1} \text{ m}^{-1}$ ,  $n=3$ ) and lower in September ( $45 \pm 23\%$ ,  $0.66 \pm 0.26 \text{ L mg C}^{-1} \text{ m}^{-1}$ ,  $n=10$ ). The inferred shorter MTT in July allows for more DOM with stronger permafrost DOM fingerprint to be detected in the subsurface environment. This change is especially clear when groundwaters repeatedly sampled from a single monitoring well screened at 30 m depth below ground (MW, see Fig.1d) in July 2013, September 2013 and 2018

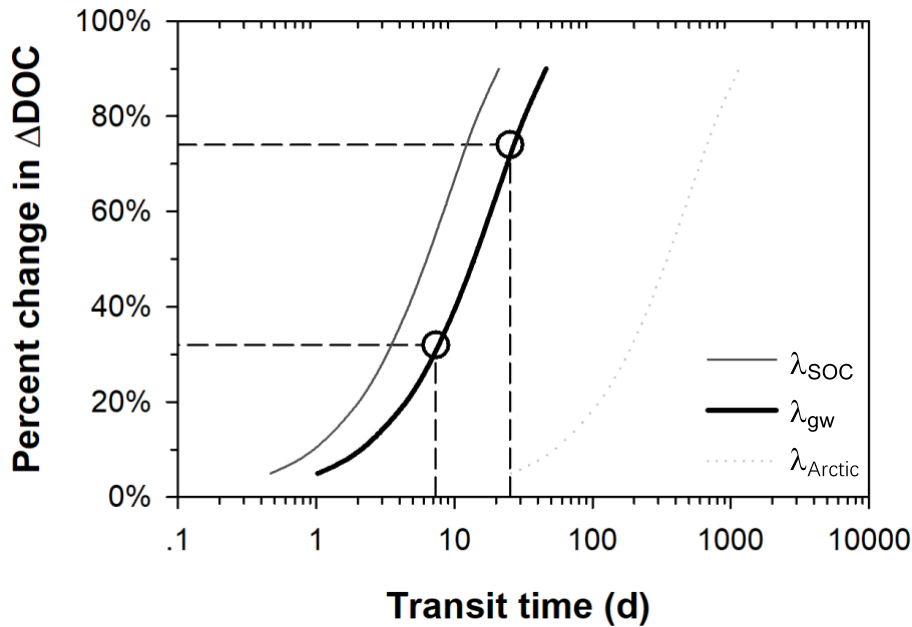
are considered (Fig. 4). Moreover, the DOC contrast in surface water is also consistent with this difference in transit time (Fig. 4). The higher discharge and thus more extensive surface water and groundwater interaction in July compared to September accelerates the already rapid downward flow of groundwater in HLGW, also supported by field observation of the spring discharge at the outlet of HLGW.

Third, an attempt is made to estimate groundwater MTT based on the DOC dynamics described above, assuming subsurface biodegradation of DOC with first order kinetics ( $\lambda$ ) that is known to vary so the rationale for our choice is described. Incubation of 12 riverine DOM samples yield  $\lambda$  from 0.147 to 0.781 d<sup>-1</sup> in headwater regions of the Yangtze, Yellow and Lantsang -Mekong rivers [Ma *et al.*, 2018]. We adopt the average  $\lambda$  value of 0.25 d<sup>-1</sup> from 2 riverine DOM samples in 2 small alpine watersheds (94.3304°E, 35.75439° N) of the Yellow River in Qinghai as the most likely for HLGW, given the similar topography, landscape (glacier and permafrost), proximity (~ 500 km from HLGW), and comparable DOC concentration of 2.5±0.4 mg L<sup>-1</sup> and discharge of 1.27 m<sup>3</sup>/s [Ma *et al.*, 2018]. The  $\lambda$  becomes 0.06 d<sup>-1</sup> after correcting to HLGW groundwater temperature of 5°C following the Arrhenius equation [Catalan *et al.*, 2016]. Using 0.06 d<sup>-1</sup>, the MTT is estimated to be approximately 6 days and 20 days in July and September, respectively, corresponding to the changes in  $\Delta$ DOC ( $\Delta$ DOC/DOC<sub>0</sub>) observed for groundwater of monitoring well at HLGW outlet of 32% in July and 74% in September (Fig. 5). Because most DOC entered the aquatic environment in the upper stretch of HLGW (Fig. 1), this estimation of reaction time based on DOC degradation is thus indicative of MTT for groundwater in HLGW.

Fourth, the sensitivity of MTT estimate to variable  $\lambda$  is described as follows. The SOC degradation constant of 0.32 d<sup>-1</sup> established from our HLGW soil at 20°C is within the range of  $\lambda$  from the 12 riverine DOM sample. However, incubation studies of Arctic water DOM have found variable  $\lambda$  of 5×10<sup>-3</sup> to 0.15 d<sup>-1</sup> [Balcarczyk *et al.*, 2009; Fellman *et al.*, 2008; Fellman *et al.*, 2009; Spencer *et al.*, 2015]. The highest  $\lambda$  of 0.15 d<sup>-1</sup> is established from the first-order streams of the Kolyma River, most likely to be representative of highly labile and freshly released DOC in Arctic headwaters [Spencer *et al.*, 2015], and thus relevant. The slowest  $\lambda$  of 5×10<sup>-3</sup> d<sup>-1</sup> is reported for incubation of DOM in the stream waters from large basins with long retention time and Holocene deposit in Alaska at 4°C with in situ nutrients [Balcarczyk *et al.*, 2009].

Although these very long rates are unlikely, we have estimated the shortest and longest MTT using the faster  $\lambda_{\text{SOC}}$  and slowest  $\lambda$ . The  $\lambda_{\text{SOC}}$  becomes  $0.11 \text{ d}^{-1}$  at  $5^{\circ}\text{C}$  after temperature correction, and the MTT is estimated to be 3.5 d and 12.5 d for July and September, respectively (Fig. 5). Based on the slowest  $\lambda$  of  $5 \times 10^{-3} \text{ d}^{-1}$  the MTT can be as long as 200 d for July and 700 d for September, respectively (Fig. 5).

There are several limitations to our MTT estimation. Given the degrees of spatial and temporal variability in DOM quantity and quality, biodegradation kinetics of DOM warrants further investigation. Perhaps more useful would be an independent assessment of groundwater MTT using Ra-Rn isotopes [Luo and Jiao, 2019]. Further, changes in DOC and SUVA<sub>254</sub> from our soil incubation experiments suggest release of aromatic carbon (Table S2). This raises the yet to be assessed possibility that preferential sorption of highly aromatic carbon or humic-like compounds onto soil or sediment may contribute to DOC loss along the groundwater flow path [Jin and Zimmerman, 2010]. Finally, long-term observations of DOM quantity and quality are desirable to reveal DOM dynamics that will allow separation of effects of gradual, seasonal freeze-thaw cycles and abrupt, accelerated thawing of permafrost. These limitations are unlikely to challenge the finding that subsurface DOM processing is primarily driven by biodegradation, and the rate constants are similar in July and September. If so, the results are interpreted to suggest that MTT in hillslopes vary in response to discharge corresponding to season freeze-thaw cycles.



**Figure 5.** Percent changes in  $\Delta\text{DOC}$  vs MTT (d) shown in log scale. Black line is for the most likely biodegradation rate constant ( $\lambda_{\text{gw}}$ ) of  $0.06 \text{ d}^{-1}$ . The different groundwater MTT is shown as white circles to reflect percent changes in  $\Delta\text{DOC}$  observed at MW in July (32%) and September (74%), respectively. The gray line to the left indicates a constant ( $\lambda_{\text{soc}}$ ) of  $0.11 \text{ d}^{-1}$  at  $5^\circ\text{C}$  based on HLGW soil incubation experiment. The dotted gray line ( $\lambda_{\text{Arctic}}$ ) to the right indicates that MTT estimates using the lowest observed  $\lambda$  of  $5 \times 10^{-3} \text{ d}^{-1}$  based on incubation of Arctic stream water samples [Balcarczyk *et al.*, 2009].

#### 4.3 Hillslopes Act as Hotspots of Permafrost Derived DOC Processing in Subsurface Environment

Although better constraints on DOC degradation kinetics and repeated monthly sampling from June to December of groundwater would likely result in improved estimates of mean transit time in HLGW, the results nevertheless demonstrate that DOM in permafrost regions of the QTP can be used to shed light on hillslope hydrological process in its headwaters. A long-held view is that aquifer is dominated by water with older ages ( $> 3$  months) compared to riverine systems of much younger water ( $< 3$  months) [Jasechko *et al.*, 2017]. Recent studies suggest a component of groundwater is very young in age [Gleeson *et al.*, 2016], yet unraveling the age distribution is currently challenging with long-term tracer observations [Luo and Jiao, 2019; McDonnell *et al.*, 2010]. Though fraught with uncertainties, the estimation above is addressing a challenging

problem, and represents the first attempt to quantify hillslope groundwater transit time in the QTP. A global evaluation has shown that the DOC decomposition rate in inland waters ranges from 0.0003 to 9 d<sup>-1</sup> corresponding to a water retention time of 0.04 day to 42 years [Catalan *et al.*, 2016]. This study supports the notion that hillslopes are hotspots for DOC processing with subsurface environment playing an important role, expanding the coverage of the aforementioned global evaluation to include an important inland water system in QTP.

Existing and newly gained insights on groundwater flow system in HLGW suggest extensive surface water and groundwater interaction, supported by modeling [Evans *et al.*, 2015], isotopic [Ma *et al.*, 2017], hydrochemical [Li *et al.*, 2014; Li *et al.*, 2016] and now DOM quantity and quality data. These studies have identified shallow groundwater sourced primarily from glacier-snow and precipitation [Chang *et al.*, 2018], and flows down gradient above permafrost layer and later above the clay layer (Fig. 1d) [Evans *et al.*, 2015; Ma *et al.*, 2017]. Hydrogeological studies have shown that the subsurface water's radiocarbon age is modern and contains tritium (<sup>3</sup>H) at shallow depth (<20 m) in permafrost and seasonal frost zones of the HLGW [Ma *et al.*, 2017]. This understanding of groundwater flow together with its extensive interaction with surface water is taken as representative of headwater watersheds with permafrost of alpine hillslopes in the entire QTP. Considering the significantly lower DOC loss in July than that in September based on differences in ΔDOC, the DOC fluxes were estimated for July and other months, June to December except July. This assumes that September is representative of other months which have lower discharges (Fig. 4), and the input and export of water is mass balanced thus the same. Fluxes between January and May are not considered because the discharge is very low and the soil is frozen.

We put this all together to estimate DOC input and output fluxes to illustrate the extent of DOC loss in HLGW. In July, the DOC export is estimated to be 6.8×10<sup>3</sup> kg by multiplying monthly discharge and DOC concentration of 2.4 mg L<sup>-1</sup> from the outlet stream water of HLGW (Table 4). We assume that the discharge of the HLGW outlet equals to the total of water recharge from the three end-members to meet the volume balance. We take the DOC mass of glacier-snow melting water as an example to present the estimation of DOC input. The input water volume is calculated as total discharge of 2.8 ×10<sup>6</sup> m<sup>3</sup> multiplying its contribution fraction of 23% in the outlet water sample. The input DOC mass of 0.3×10<sup>3</sup> kg is based on the DOC value in glacier

end member of 0.5 mg L<sup>-1</sup> multiplying the water volume. Therefore, the DOC input flux is 8.4×10<sup>3</sup> kg, with the glacier-snow meltwater, precipitation, and soil meltwater contributing 0.3×10<sup>3</sup>, 1.7×10<sup>3</sup> and 6.4×10<sup>3</sup> kg, respectively (Table 4). Between June to December excluding July, the DOC concentration of 0.7 mg L<sup>-1</sup> in September is representative for these months. The DOC export is 3.5×10<sup>3</sup> kg, and the DOC input is 15.1×10<sup>3</sup> kg with the glacier-snow meltwater, precipitation, and soil melt water contributing 0.6×10<sup>3</sup> kg, 3.1×10<sup>3</sup> kg and 11.4×10<sup>3</sup> kg, respectively (Table 4). The annual DOC input flux is 23.5×10<sup>3</sup> kg yr<sup>-1</sup>, with an annual DOC export flux of 10.3 ×10<sup>3</sup> kg yr<sup>-1</sup>. The difference between input and export DOC fluxes is taken to represent a “lost” DOC flux of 13.2 ×10<sup>3</sup> kg yr<sup>-1</sup> within HLGW before transporting into large rivers (Fig. 1c and Table 4). About half of the DOC is processed in hillslopes of small alpine watersheds, comprising a substantial fraction in C cycling in permafrost region under warming and warrant further attention.

**Table 4.** Input, export and respired DOC fluxes from HLGW.

Input DOC from three endmembers							
End membe rs	DOC (mg/L)	Frac <sub>tion</sub>	July		June-Dec w/o July *		Annual flux (10 <sup>3</sup> kg/yr)
			Water vol. (10 <sup>6</sup> m <sup>3</sup> )	DOC mass (10 <sup>3</sup> kg)	Water vol. (10 <sup>6</sup> m <sup>3</sup> )	DOC mass (10 <sup>3</sup> kg)	
G	0.5	23%	0.65	0.3	1.16	0.6	
P	1.0	62%	1.74	1.7	3.12	3.1	
S	15.0	15%	0.42	6.4	0.76	11.4	
Input DOC (x10 <sup>3</sup> kg)			8.4		15.1		23.5
Export DOC at the outlet of HLGW							
Stream at outlet	July			June-Dec w/o July			Annual flux (10 <sup>3</sup> kg/yr)
	DOC (mg/L)	Water vol. (10 <sup>6</sup> m <sup>3</sup> )	DOC mass (10 <sup>3</sup> kg)	DOC (mg/L)	Water vol. (10 <sup>6</sup> m <sup>3</sup> )	DOC mass (10 <sup>3</sup> kg)	
Export DOC (x10 <sup>3</sup> kg)	2.4	2.8	6.8	0.7	5.0	3.5	10.3
Loss of DOC during in-stream processing							
Loss of DOC (10 <sup>3</sup> kg)	July			June-Dec w/o July			Annual
	1.6			11.6			13.2
% lost DOC	19%			77%			56%

\* Input and output DOC between January and May are not considered due to seasonal freeze.

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*[JGR Biogeosciences]*

Supporting Information for

**[Hillslopes in Headwaters of Qinghai-Tibetan Plateau as Hotspots for Subsurface Dissolved Organic Carbon Processing during Permafrost Thaw]**

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**Additional Supporting Information (Files uploaded separately)**

Captions for Datasets S1 to S2

**Introduction**

Supporting information contains descriptions of sampling protocols, field and laboratory measurement details (Text S1), soil incubation experimental procedure (Text S2) and results (Text S3). Figures S1 and S6 illustrates the variations of SUVA<sub>254</sub> FI and BIX. Figure S2 depicts the soil incubation experiment set up. Figures S3 – S5 show fluorescent characteristics of the DOM samples and PARAFAC modeling results. Tables S1 reports uncertainties of endmember compositions and analysis. Table S3 and Figure S7 report soil incubation experimental data. Dataset S1 to S2 (separate electronic files) are original data of field and laboratory measurements of water chemistry parameters for all samples.

### Text S1. Sampling and Measurements.

Temperature (T), electrical conductivity (EC) and pH were measured in the field using WTW-COND-3301 instrument in 2012 and 2013, and Thermo 520M-01A instrument in 2018. Alkalinity was measured using titration method in the field [Gran, 1952]. All samples were stored on ice at 4°C for shipment to laboratory.

Stable isotopes ( $\delta\text{D}$  and  $\delta^{18}\text{O}$ ) were measured for samples collected in July 2013, and September of 2013 and 2018 on a water isotope spectrometer analyzer (Model PICARRO L2130-I) at Pri-ecoco, Beijing, China. Three standard solutions (GBW58, GBW59, and GBW60) were employed as daily QA/QC. Each sample was analyzed 6 times and the result was calculated as the average of the last three injections. The relative standard deviation (RSD) for duplicate analysis was < 0.8%. The isotope compositions are expressed as the  $\delta$ -notation related to Vienna Standard Mean Ocean Water (VSMOW) in ‰.

DOC concentrations for samples collected in 2012 and 2013 were measured on a TOC-5000 Analyzer (Shimadzu) at both the Environmental Engineering Laboratory, Peking University and Queens College, CUNY according to a Non Purgeable Organic Carbon (NPOC) method. Samples collected in 2018 were analyzed by a TOC-L Analyzer (Shimadzu) at Southern University of Science and Technology following the same protocol. Samples were analyzed in triplicate with the RSD less than 3% as a threshold for accepting the result.

Measurement of cations was performed using Inductively Coupled Plasma Atomic Emission Spectrometry (ICP-AES) at Peking University for samples collected in 2012 and 2013. Anions of nitrate ( $\text{NO}_3$ ), sulfate ( $\text{SO}_4$ ) and chloride ( $\text{Cl}$ ) were measured using ion chromatography (IC) at Peking University. Major anions and cations concentrations were measured using IC at Southern University of Science and Technology for samples collected in 2018.

### Text S2. Experimental Procedure of Soil Incubation.

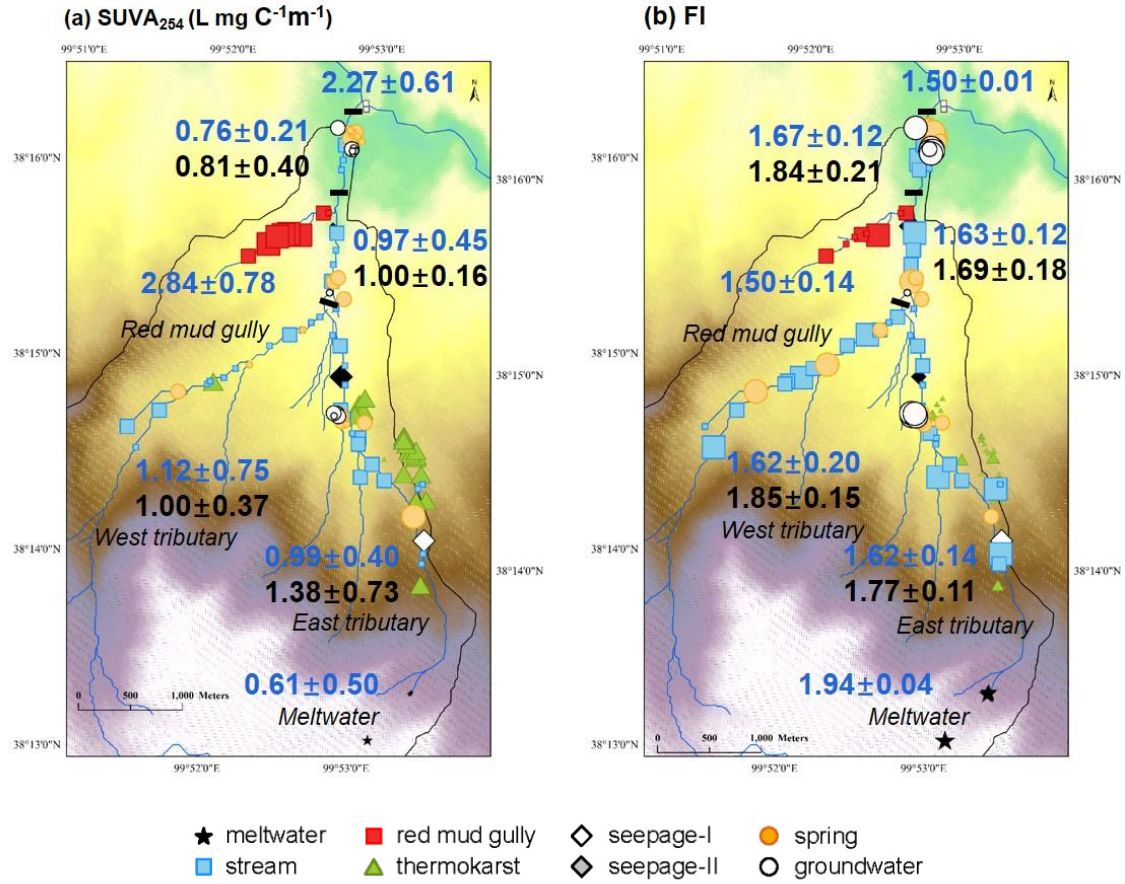
At the start of the second incubation in the reactor, 100 g of wet soil was weighed and added to a glass reactor and filled with 5L  $\text{CO}_2$ -free MQ water without headspace at room temperature. The reactor was kept in dark for 80 hours, with a peristaltic pump running at 29 ml/min to ensure mixing. The inlet and outlet of the reactor were connected with 6M NaOH solution to maintain  $\text{CO}_2$ -free headspace (Fig. S1). A total of 29 samples were collected at an interval of 30 min in the initial 4.6 hours, at an interval of 60 min till 12 hours, and at an interval of 180 min until the end of incubation that lasted 80 hours. At each sampling time, approximately 75 ml of water was removed. Water volume and pH were measured immediately before filtering with 0.22  $\mu\text{m}$  SPE filters. Filtered samples were analyzed for DOC concentrations, UV-vis and fluorescence as in section 2.2 of the main text.

### Text S3. Soil Incubation Results.

During the 80-hours dark incubation of the frost soil in the reactor, the carbon mass of DOC decreases from 30 mg to 12 mg in the initial 12 hours (Fig. S6a). The utilization of aromatic DOC is supported by the doubling of  $\text{SUVA}_{254}$  values concurrent with a 60% decrease in DOC mass in the system between 0 to 12 hours (Fig. S6). From 12 hours to 80 hours,  $\text{SUVA}_{254}$  values instead decreased with an increase of DOC mass from 12 mg to 18 mg (Fig. S6c). The EEMs of all incubation samples show the main fluorescent peak at excitation of 230 nm and emission of 240 nm, suggesting that protein-like fluorophores are significant.

77  
78       The first-order kinetic equation was applied to fit the SOC degradation curves for the  
79 decrease in DOC mass in the first 12 hrs (Fig. S6c). The half-time of soil derived DOC degradation  
80 is estimated to be 5 hours, and the degradation constant is calculated as dividing  $-\ln(0.5)$  by half-  
81 time. Therefore, the degradation constant following first-order kinetics of DOC derived from  
82 permafrost soil is estimated to be  $0.32 \text{ d}^{-1}$  at  $20^{\circ}\text{C}$ , and is corrected to  $0.11 \text{ d}^{-1}$  for  $5^{\circ}\text{C}$ .  
83

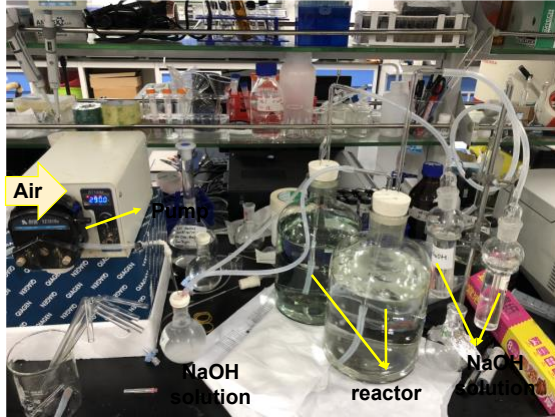




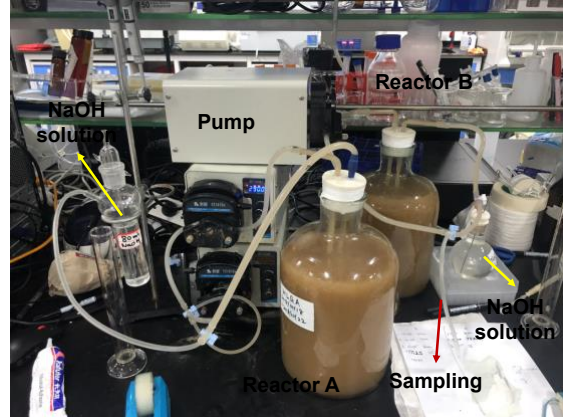
**Figure S1.** (a) SUVA<sub>254</sub> for eight types of water in HLGW, Qinghai-Tibetan Plateau. Small, medium and large symbol sizes indicate low (< 0.91 L mg C<sup>-1</sup> m<sup>-1</sup>), medium (0.91–2.46 L mg C<sup>-1</sup> m<sup>-1</sup>) and high (>2.46 L mg C<sup>-1</sup> m<sup>-1</sup>) SUVA<sub>254</sub> according to tertile values of the entire dataset. Numbers are mean value ± one standard deviation for SUVA<sub>254</sub> in stream (blue) and subsurface water (black). (b) FI with small, medium and large symbol sizes indicate low (<1.59), medium (1.59–1.69) and high (>1.69) according to tertile values. Numbers are mean value ± one standard deviation for FI in stream (blue) and subsurface water (black).

93

(a) Preparation of CO<sub>2</sub>-free MQ water

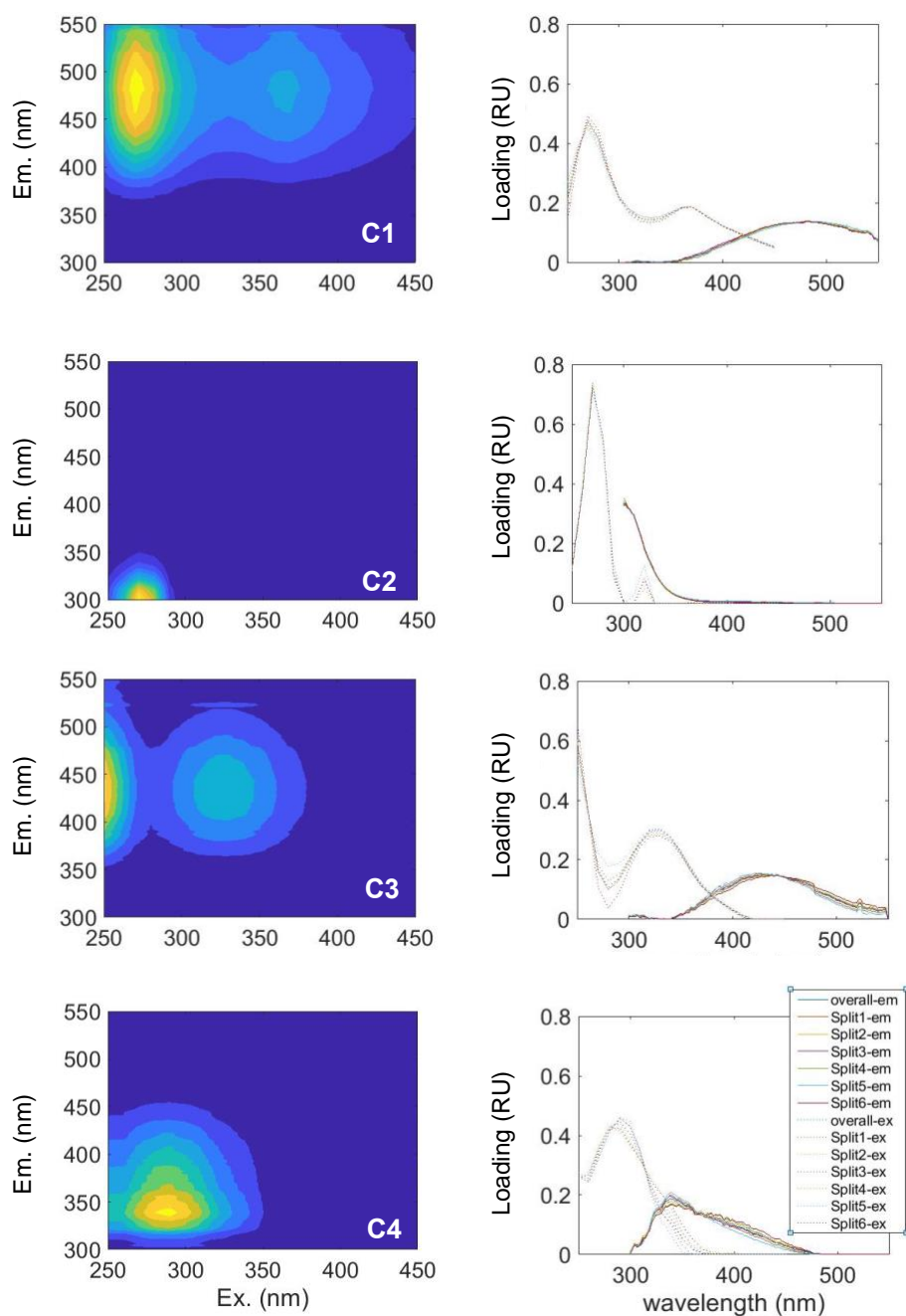


(b) Incubation reactor setting up and sampling



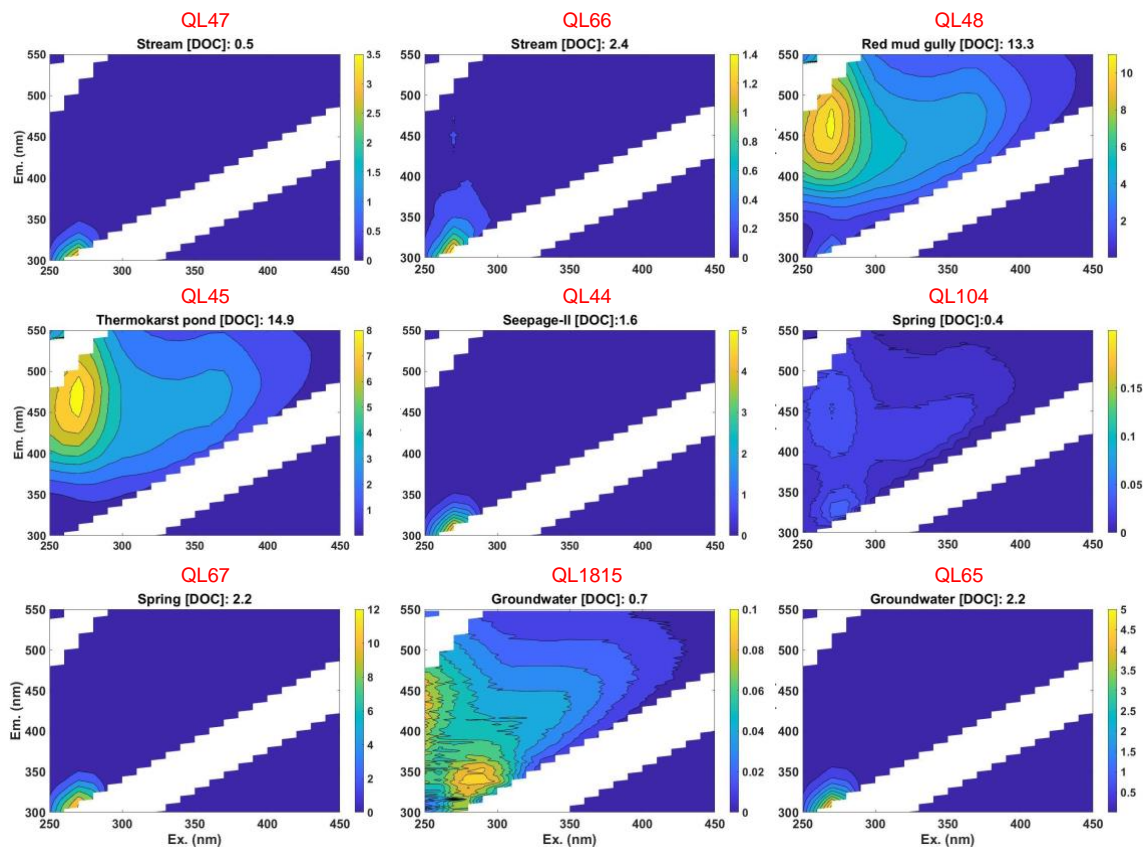
94

95 **Figure S2.** Figure S1. Experimental set up of frost soil reactor incubation. Left photo shows the  
96 preparation of CO<sub>2</sub>-free water for the two reactors. Right photo shows ongoing experiment with  
97 sampling port location during incubation. The reactors used 5-L glass containers with inlet and  
98 outlet ports. Continuous mixing is ensured by pumping water at a rate of 29 mL min<sup>-1</sup> from the  
99 reactors by a peristaltic pump through a CO<sub>2</sub>-free chamber. Samples were collected by a syringe  
100 at the sampling port with a three-way valve.



**Figure S3.** Fluorescence matrixes of the four identified components (left) and loadings of excitation (the dot lines) and emission (the solid lines) wavelength (right), respectively. Component 1 (ex: 270/370 nm, em: 470 nm) and component 3 (ex: <250/330 nm, em:420 nm) represent two humic-like components. Fluorescence characteristic of C2 (ex: 270 nm, em: 304 nm) represents tyrosine-like material [D'Andrilli *et al.*, 2019] (20), and C4 (ex: 290 nm, em: 338 nm) represents amino-acids, free or bound in proteins compounds [Catala *et al.*, 2015; Murphy *et al.*, 2008].

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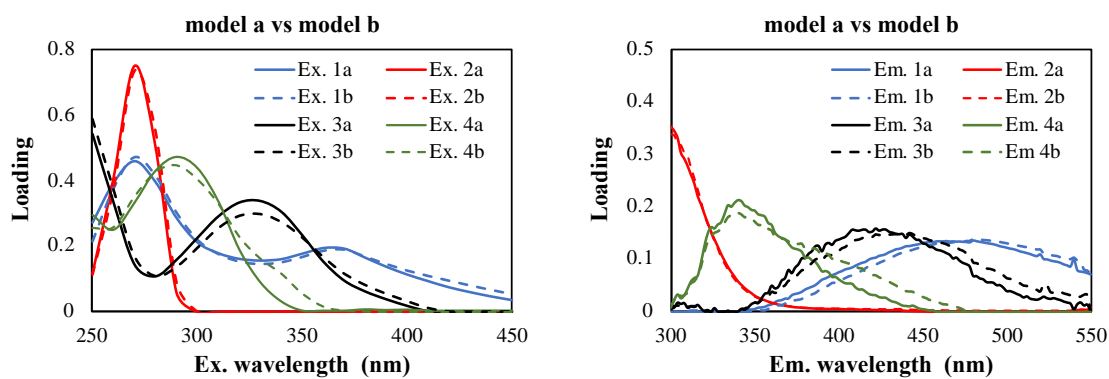


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112 **Figure S4.** EEMs plots of typical stream, red mud gully, thermokarst pond, seepage-II, spring and  
113 groundwater samples in HLGW. Sample ID is written in red (see Data set S1 for details), and the  
114 DOC concentration ( $\text{mg L}^{-1}$ ) is written in black above each EEMs. Color bar represents the  
115 fluorescence intensity.

116

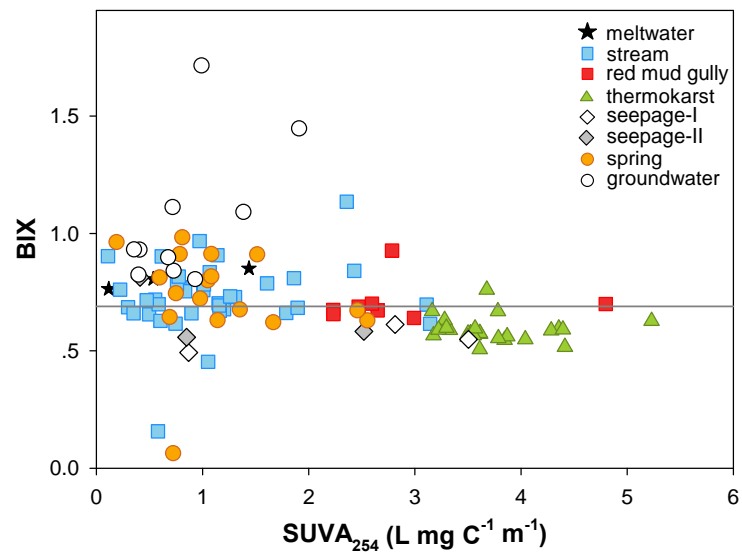
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118

119 **Figure S5.** Comparison of excitation (Ex.) and emission (Em.) loadings of the four-components  
 120 models.

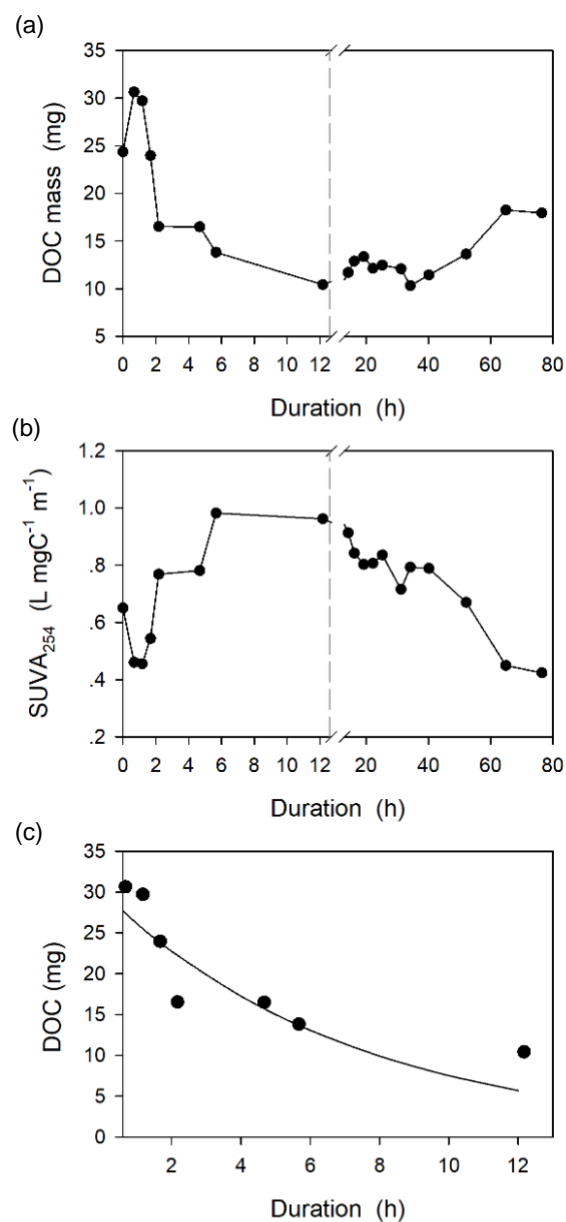
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123

124 **Figure S6.** SUVA<sub>254</sub> vs BIX for eight types of water samples with gray line indicating the median  
125 value of BIX for entire dataset.

126



129 **Figure S7.** Changes of (a) DOC expressed as carbon mass in supernatant and (b) SUVA<sub>254</sub> values  
130 over 80 hrs for the reactor soil incubation experiment in dark. The first-order kinetics fit to the  
131 soil carbon derived DOC degradation data in the first 12 hrs is shown in (c).  
132

Sample ID	Type	$f_G$	$f_P$	$f_S$	$W_G$	$W_P$	$W_S$	Sample ID	Type	$f_G$	$f_P$	$f_S$	$W_G$	$W_P$	$W_S$
QL41	stream	0.20	0.73	0.08	0.48	0.46	0.02	QL67	spring	0.23	0.61	0.17	0.37	0.36	0.03
QL42	stream	0.26	0.68	0.06	0.35	0.34	0.02	QL71	spring	0.23	0.71	0.06	0.36	0.35	0.03
QL43	stream	0.31	0.65	0.04	0.49	0.47	0.02	QL72	spring	0.29	0.66	0.05	0.39	0.38	0.02
QL47	stream	0.26	0.68	0.06	0.42	0.41	0.02	QL88	spring	0.27	0.56	0.17	0.42	0.41	0.03
QL58	stream	0.22	0.73	0.05	0.38	0.36	0.02	QL104	spring	0.30	0.53	0.18	0.45	0.44	0.03
QL61	stream	0.24	0.73	0.04	0.40	0.38	0.02	QL1812	spring	0.22	0.61	0.17	0.36	0.35	0.03
QL64	stream	0.23	0.72	0.05	0.39	0.37	0.02	QL1813	spring	0.18	0.66	0.16	0.32	0.31	0.03
QL66	stream	0.24	0.66	0.10	0.39	0.38	0.02	QL1818	spring	0.23	0.58	0.19	0.37	0.36	0.03
QL68	stream	0.21	0.72	0.07	0.37	0.35	0.02	QL1819	spring	0.22	0.59	0.19	0.36	0.35	0.03
QL73	stream	0.23	0.59	0.18	0.37	0.36	0.03	QL1820	spring	0.21	0.61	0.18	0.35	0.34	0.03
QL83	stream	0.27	0.59	0.13	0.43	0.42	0.02	QL1821	spring	0.22	0.61	0.16	0.37	0.36	0.03
QL84	stream	0.25	0.64	0.11	0.40	0.39	0.02	QL1830	spring	0.31	0.64	0.05	0.48	0.46	0.02
QL85	stream	0.27	0.60	0.13	0.42	0.41	0.02	QL1831	spring	0.22	0.61	0.17	0.46	0.45	0.02
QL86	stream	0.26	0.61	0.13	0.42	0.40	0.02	QL65	groundwater	0.21	0.62	0.17	0.35	0.34	0.03
QL87	stream	0.22	0.62	0.16	0.36	0.35	0.02	QL94	groundwater	0.27	0.59	0.14	0.42	0.41	0.03
QL90	stream	0.30	0.54	0.16	0.47	0.45	0.03	QL95	groundwater	0.27	0.58	0.14	0.43	0.42	0.03
QL92	stream	0.29	0.55	0.16	0.44	0.43	0.03	QL98	groundwater	0.11	0.74	0.15	0.27	0.26	0.02
QL99	stream	0.26	0.66	0.08	0.42	0.40	0.02	QL1808	groundwater	0.17	0.66	0.17	0.31	0.31	0.03
QL100	stream	0.26	0.65	0.09	0.42	0.40	0.02	QL1809	groundwater	0.17	0.67	0.17	0.31	0.30	0.03
QL101	stream	0.25	0.66	0.09	0.41	0.40	0.02	QL1810	groundwater	0.21	0.59	0.20	0.34	0.33	0.03
QL102	stream	0.25	0.66	0.09	0.40	0.39	0.02	QL1811	groundwater	0.17	0.69	0.14	0.32	0.31	0.02
QL103	stream	0.25	0.66	0.09	0.40	0.39	0.02	QL1815	groundwater	0.20	0.60	0.20	0.34	0.33	0.03
QL106	stream	0.22	0.67	0.12	0.36	0.35	0.02	QL1840	groundwater	0.22	0.63	0.15	0.37	0.35	0.03
QL1814	stream	0.29	0.57	0.14	0.44	0.43	0.03	QL1841	groundwater	0.22	0.63	0.15	0.36	0.35	0.03
QL1816	stream	0.27	0.59	0.14	0.43	0.42	0.03	QL1842	groundwater	0.21	0.64	0.15	0.35	0.34	0.03
QL1817	stream	0.26	0.59	0.15	0.42	0.40	0.03								
QL1829	stream	0.29	0.66	0.05	0.46	0.44	0.02								
QL1832	stream	0.32	0.61	0.08	0.49	0.47	0.02								
QL1833	stream	0.31	0.61	0.08	0.48	0.46	0.02								
QL1834	stream	0.25	0.60	0.15	0.40	0.39	0.03								
QL1836	stream	0.45	0.50	0.04	0.68	0.66	0.03								
QL1837	stream	0.38	0.50	0.12	0.58	0.56	0.03								
QL1838	stream	0.23	0.68	0.09	0.38	0.37	0.02								
QL1839	stream	0.35	0.50	0.15	0.54	0.52	0.03								
QL1844	stream	0.50	0.46	0.04	0.75	0.72	0.03								

134

135 **Table S1.** Unmixed fractions of glacier-snow ( $f_G$ ), precipitation ( $f_P$ ) and soil ( $f_S$ ) endmember  
136 contributions and the associated uncertainties for glacier-snow ( $W_G$ ), precipitation ( $W_P$ ) and soil  
137 ( $W_S$ ). For isotopic and EC “conservative” tracer concentrations and other chemical data of each  
138 sample, see Dataset S1 and S2.

139



Depth b.s.l	Time (d)	DOC (mg/ L solution)	DOC (mg/g wet soil)	SUVA <sub>254</sub> (L mg C <sup>-1</sup> m <sup>-1</sup> )	Protein- like percent	FI	BIX	Protein- like intensity (RU)
profile 1: thawed soil in seasonal permafrost zone near red mud gully (38.2630N, 99.8777E, 2850 m a.s.l)								
5 cm	3	13.1	0.05	3.48		1.33	0.48	
	40	282.8	1.10	0.88		1.41	0.47	
15 cm	3	10.0	0.04	3.53	22%	1.29	0.52	0.12
	40	125.4	0.35	0.98	11%	1.53	0.52	0.10
30 cm	3	13.6	0.02	3.79	29%	1.38	0.60	0.25
	40	16.1		3.87	13%	1.50	0.52	0.12
50 cm	3	27.8	0.05	1.71	42%	1.54	0.69	0.45
	40	33.1	0.06	1.88	13%	1.50	0.52	0.12
profile 2: dried thermokarst sediment (38.2423N, 99.8859 E, 3500 m a.s.l)								
10 cm	3	15.4	0.24	4.07	22%	1.33	0.53	0.22
	40	34.4	0.13	6.13	7%	1.41	0.48	0.17
30 cm	3	15.6	0.03	2.92	33%	1.51	0.60	0.34
	40	23.6	0.05	4.27	19%	1.43	0.54	0.13
80 cm	3	16.6	0.03	2.83	33%	1.47	0.58	0.39
	40	34.0	0.05	2.98	22%	1.35	0.53	0.18
profile 3: wet thermokarst sediment (38.2443N, 99.8879 E, 3500 m a.s.l)								
10 cm	1	18.7	0.06	1.48	20%	1.43	0.48	0.13
	3	44.8	0.14	1.33	10%	1.38	0.46	0.05
	40	46.8	0.16	4.18	6%	1.42	0.49	0.07
20 cm	1	28.0	0.07	2.08	19%	1.43	0.50	0.17
	3	53.3	0.14	1.93	9%	1.31	0.44	0.06
	40	42.5	0.12	4.40	9%	1.34	0.50	0.10
30 cm	1	23.1	0.06	2.51	19%	1.45	0.49	0.20
	3	63.5	0.16	1.80	9%	1.41	0.50	0.09
	40	38.9	0.11	4.44	7%	1.36	0.47	0.10
profile 4: permafrost zone (38.2414N, 99.8862E, elevation: 3600 m)								
5 cm	1	12.4	0.09	3.64	18%	1.29	0.42	0.11
	3	24.4	0.20	2.91	61%	1.35	0.48	1.17
	40	129.5	0.80	1.29	40%	1.45	0.55	0.58
15 cm	1	13.4	0.06	3.80	17%	1.30	0.42	0.11
	3	15.3	0.06	4.45	15%	1.29	0.48	0.14
	40	55.4	0.27	2.42	13%	1.40	0.50	0.11
25 cm	1	15.8	0.08	2.92	21%	1.27	0.43	0.13
	3	18.0	0.09	3.20	13%	1.30	0.45	0.11
	40	99.2	0.57	1.64	20%	1.38	0.55	0.21
35 cm	1	11.6	0.05	5.05	20%	1.32	0.43	0.15
	3	18.9	0.07	4.30	19%	1.27	0.48	0.20
	40	32.5	0.11	3.38	23%	1.38	0.56	0.15

**Table S2.** DOC concentrations and fluorescence properties for four soil profiles in soil batch incubation experiment after 1, 3 and 40 extraction days.

**Dataset S1.** Detailed sampling information including water types, sampling date (year/month/date), locations (Lat., Long, and Elev.), electrical conductivity (EC,  $\mu\text{S}/\text{cm}$ ). Calculations including proportions of glacier-snow meltwater, precipitation, and frozen soil meltwater end members (G, P, S) based on stable isotopes and EC, the expected initial DOC ( $\text{DOC}_0$ ) and utilized DOC ( $\Delta\text{DOC}$ ). DOC concentrations ( $\text{mg L}^{-1}$ ) and DOM optical parameters including  $\text{SUVA}_{254}$  ( $\text{L mg C}^{-1}\text{m}^{-1}$ ), fluorescence index (FI), freshness index (BIX), percentage proportions of protein-like component (%) derived from PARAFAC.

\* Only DOC concentrations were available for samples collected in Apr 2012. One thermokarst sample collected near the west tributary was excluded in the data analysis.

† The 7 groundwater samples were collected from the same monitoring well screened at 30 m at the outlet (see MW in Fig.1d and Fig 3).

† The four groundwater samples were collected from the WW3 well (see WW3 in Fig.1d) located in piedmont sloping plain dominated by seasonal frost, with screened intervals being 5, 10, 20 and 30 m underground, respectively.

§ The groundwater/well water was collected from the WW4 with depth at 1m without pumping.

† The groundwater sample was collected from the WW1 well screened at 25 m (WW1 in Fig.1d).

**Dataset S2.** Additional field measurements including water temperature, pH, alkalinity (Alk,  $\text{meq L}^{-1}$ ), dissolved oxygen (DO,  $\text{mg L}^{-1}$ ). Concentrations of major ions and total nitrogen (TN,  $\text{mg L}^{-1}$ ). The intensity loadings of the four components derived from PARAFAC (C1 to C4).

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