Horizontal variability and regulation of bacterial production in a large, temperate lake

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Abstract

To clarify horizontal variability and regulation of bacterial production (BP), we investigated BP and environmental variables along three east-west transects (Lines 12, 15, and 17) covering inshore/offshore stations in Lake Biwa, Japan, during four seasons. In winter, surface BP along Line 12 (southern transect) was higher than Lines 15 and 17 (central and northern transects) and reflected the water-temperature distribution. Additionally, any nutrients and dissolved organic carbon did not correlate to BP, suggesting water temperature regulated BP in winter. In spring, BP was higher at eastern inshore stations, near large agricultural fields, and was correlated with phosphorus concentration rather than water temperature, suggesting that the limitation shifted to nutrient availability. As well as spring, surface BP in autumn was correlated with phosphorus. Additionally, a negative correlation with water temperature in autumn suggested that nutrient loadings through river and groundwater contributed to enhancing BP. In summer, surface BP at offshore stations along Lines 15 and 17 was notably lower than the other stations. Summer BP was correlated with phosphorus concentration, suggesting that allochthonous nutrient loading determines horizontal BP variations. Moreover, summer depth-integrated BP (DBP) at offshore stations was lower (32-52 mgC m-2 d-1) than inshore stations (43-110 mgC m-2 d-1) regardless of water depth. The average DBP at inshore stations in summer was 2.1 times that offshore stations, and the inshore/offshore DBP ratio was higher than the other seasons (0.58-1.0). The results suggest that inshore BP significantly contributes to whole-lake BP in productive seasons.

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12	Key Points:
13	• Horizontal heterogeneity of bacterial production (BP) was pronounced in a large lake,
14	showing different regulation factors among seasons
15	• BP varied along with a gradient of water temperature in winter and phosphorus in the
16	other seasons
17	• Depth-integrated BP in inshore area significantly contributed to the whole-lake BP
18	despite the lower water depths than offshore area
19	
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21 Abstract

22 To clarify horizontal variability and regulation of bacterial production (BP), we investigated BP and environmental variables along three east-west transects (Lines 12, 15, and 17) covering 23 24 inshore/offshore stations in Lake Biwa, Japan, during four seasons. In winter, surface BP along Line 12 (southern transect) was higher than Lines 15 and 17 (central and northern transects) and 25 reflected the water-temperature distribution. Additionally, any nutrients and dissolved organic 26 carbon did not correlate to BP, suggesting water temperature regulated BP in winter. In spring, 27 28 BP was higher at eastern inshore stations, near large agricultural fields, and was correlated with 29 phosphorus concentration rather than water temperature, suggesting that the limitation shifted to nutrient availability. As well as spring, surface BP in autumn was correlated with phosphorus. 30 Additionally, a negative correlation with water temperature in autumn suggested that nutrient 31 loadings through river and groundwater contributed to enhancing BP. In summer, surface BP at 32 33 offshore stations along Lines 15 and 17 was notably lower than the other stations. Summer BP 34 was correlated with phosphorus concentration, suggesting that allochthonous nutrient loading determines horizontal BP variations. Moreover, summer depth-integrated BP (DBP) at offshore 35 stations was lower (32–52 mgC $m^{-2} d^{-1}$) than inshore stations (43–110 mgC $m^{-2} d^{-1}$) regardless 36 of water depth. The average DBP at inshore stations in summer was 2.1 times that offshore 37 stations, and the inshore/offshore DBP ratio was higher than the other seasons (0.58–1.0). The 38 39 results suggest that inshore BP significantly contributes to whole-lake BP in productive seasons.

40

41 Plain Language Summary

42 Lakes receive large amounts of external nutrient and organic matter, affecting bacterial production (BP) and spatial distributions. To estimate a whole-lake carbon budget, we need to 43 understand how BP varies spatially within the lake ecosystem and what factors regulate it. We 44 45 conducted whole-lake samplings to quantify water quality and BP and examine their 46 relationships in Lake Biwa, Japan, during four seasons. In winter, surface BP at southern stations was higher than northern stations, and there was a positive relationship between the BP and 47 water temperature. Additionally, nutrients and organic matter did not correlate to BP, suggesting 48 49 water temperature regulated BP in winter. In the other seasons, surface BP was relatively high at inshore stations and correlated with phosphorus concentration, suggesting that nutrient loadings 50

- 51 determined horizontal BP variations. The BP integrated with the entire water column in summer
- 52 at offshore stations was lower than inshore stations, despite the higher water depth at offshore
- 53 stations, whereas similar situations were not observed in the other seasons. The results suggest
- 54 that inshore BP significantly contributes to whole-lake BP in productive seasons. This study
- 55 provides essential information for estimating material cycles in lake ecosystems, contributing to
- 56 lake management and future predictions associated with climate change.
- 57

58 **1 Introduction**

Heterotrophic bacteria play an essential role in material and energy cycling in aquatic 59 ecosystems through the degradation and remineralization of organic matter (Azam, 1998). 60 Bacteria produce their biomass (= bacterial production; BP) using dissolved and particulate 61 organic matter; this production links to microbial and grazing food webs (Wylie & Currie, 1991). 62 Lake ecosystems receive large inputs of allochthonous organic matter: notably, bacterial use of 63 terrestrially-derived resources in some lake food webs can be as high as 40% to 94% (Tanentzap 64 et al., 2017). Substantial allochthonous organic matter and nutrients from surrounding 65 66 ecosystems can enter via rivers, groundwater and runoff. The relative intensity of external loading impact on ecosystems varies spatially, especially in larger lakes. Thus, horizontal 67 variability of bacterial dynamics and related environmental variables should be investigated to 68 estimate the carbon balance of a whole lake and understand the fate of allochthonous inputs. 69 Horizontal variability of BP has been studied in Lakes Michigan (Scavia & Laird, 1987), 70 Constance (Güde, 1990; Reichart & Simon, 1996), Nuesiedl (Reitner et al., 1999), Hallwil 71 (Filippini et al., 2008), and Mangueira (They & Margues, 2019). The maximum to minimum BP 72

ratios within a horizontal sampling, defined as the horizontal variability in lake ecosystems, vary
among seasons and lakes (1.6-19.5, Table 1). The BP varied along with inshore/offshore

75 gradients (allochthonous substrate dependent) (Scavia & Laird, 1987; Güde, 1990; Filippini et

al., 2008), similar gradients of primary production (autochthonous substrate dependent) (Güde,

1990; Reitner et al., 1999) and no clear gradients (Reichart & Simon, 1996; They & Marques,

2019). Although the horizontal variability of BP should have reflected their regulation factors,

there is little study available in which quantitative relationships between BP and environmental
variables have been shown in lacustrine ecosystems (e.g. a relationship between BP and PP in
Güde, 1990).

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Table 1 Ratios of the maximum (max) over the minimum (min) bacterial production in the
epilimnion (< 10-m depth) in horizontal variability. N represents the number of field
investigations.

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Lake	Stations	Season	max/min ratio	$mean \pm SD$	Ν	Reference
Michigan	3-5	Summer	1.6-3.3	2.5	2	Scavia and Laird (1987)
Constance	5	ND	11.1	11.1	1	Güde (1990)
	9	Spring, Summer	2.0-8.4	3.8 ± 2.0	12	Reichert and Simon (1996)
Neusiedl	3	Year-round	2.3-19.5	6.8 ± 4.8	22	Reitner et al. (1999)
Hallwil	3	Year-round	2.0-2.5	2.3 ± 0.3	4	Filippini et al. (2008)
Mangueira	7	Spring	1.9	1.9	1	They and Marques (2019)
Biwa	10-11	Year-round	2.9-9.5	5.2 ± 3.0	4	This study

86 87

Lake Biwa is the largest lake in Japan. The major part of this lake (670.3 km^2) is the 88 north basin, with 618 km² surface area and 46.5 m mean depth (104 m maximum depth; Fig. 1). 89 In the north basin, heterotrophic bacteria were severely limited by phosphorus (Gurung & Urabe, 90 1999). Thus, it is predictable that BP varies horizontally along a gradient of phosphorus. 91 However, since measurements of BP in this lake have been conducted only at fixed stations: Ie-1 92 (maximum depth approx. 73 m, Fig. 1; Nagata, 1987; Gurung et al., 2002; Ram et al., 2010) or 93 12B (maximum depth approx. 60 m, Fig. 1; Tsuchiya et al., 2020b), horizontal variability of BP 94 has not been clarified in this lake. Demonstrating the BP horizontal variability along with 95 gradients of phosphorus or the other environmental variables, the knowledge should contribute to 96 large-scale predictions of bacterial metabolism and understanding the biogeochemical cycling of 97 a whole lake. 98

We investigated the horizontal variabilities of BP and environmental variables in the north basin of Lake Biwa in 2018 in four seasons to elucidate the spatial and temporal heterogeneity in BP and the relationships between BP and regulation factors. Spatial distribution patterns were examined along three transects covering the inshore and offshore stations of the lake (Fig. 1). Its relations to environmental variables were analyzed in each of the four seasons.

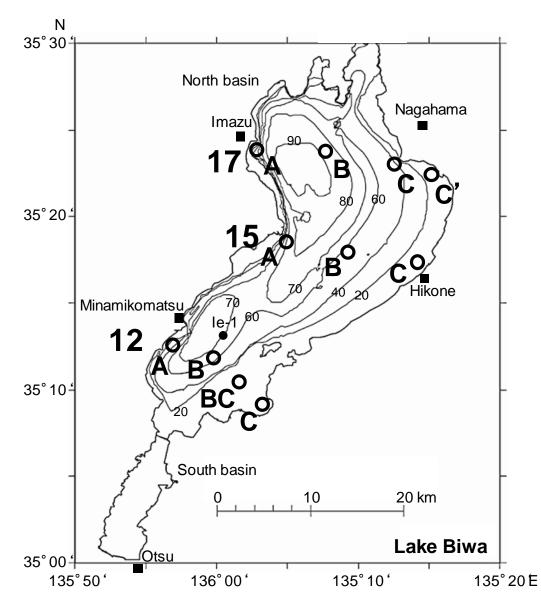


Fig. 1 Sampling stations in the north basin of Lake Biwa, Japan, along three transects (Lines 12,
15, and 17) with 3 or 4 stations each. Depth contours are in m. Square symbols represent weather
observation stations around Lake Biwa, Japan.

108

109 2 Materials and Methods

110 **2.1 Study site and samping**

111 The north basin of Lake Biwa is mesotrophic and monomictic, stratified from May to 112 November (seasonal thermocline) and well mixed from January to March (Hayakawa, 2004).

Many studies have been conducted in Lake Biwa, where environmental factors have been 113 observed over 40 years in a whole lake (Shiga-Prefecture, 2017). Additionally, since the external 114 loadings of organic matter and nutrients have been estimated in this lake (Sato et al. 2016), BP's 115 contributions to the lake's metabolism concerning the external loadings can be quantified. 116 Therefore, Lake Biwa is an appropriate lake to clarify the horizontal BP distribution and its 117 relationship with environmental variables for predicting the whole-lake carbon balance. We 118 sampled at a total of 11 stations along three transects (Fig. 1): Lines 17 (Sts. 17A, 17B, 17C, and 119 17C'), 15 (Sts. 15A, 15B, and 15C), and 12 (Sts. 12A, 12B, 12BC, and 12C) on 22 March, 11 120 May, 26 July and 21 November 2018 aboard R/V Biwakaze of the Lake Biwa Environmental 121 Research Institute, Shiga Prefecture. The water sample was collected from 3 depths (0, 20, and 122 40 m) with a bucket or a Niskin water sampler. If the water depth was shallower than the desired 123 124 collection depth, the water sample was collected 1 m above the lake bottom. In the water samples, we measured nutrients (total nitrogen TN, total phosphorus TP, total dissolved nitrogen 125 TDN, total dissolved phosphorus TDP, nitrate NO_3^- , nitrite NO_2^- , ammonium NH_4^+ and 126 phosphate PO_4^{3-}), dissolved organic carbon (DOC), and bacterial production and abundance. 127 128 Vertical profiles of water temperature, conductivity, and chlorophyll *a* concentration were determined with a conductivity-temperature-depth profiler equipped with oxygen, pH, and 129 130 chlorophyll sensors (AAQ-RINKO, JFE Advantech, Hyogo, Japan). The conversion from sensor-measured in situ chlorophyll a (chl a) to methanol-extracted chl a measured by 131 132 spectrophotometer was conducted using the following equation: [methanol-extracted chl a] = $1.08 \times [\text{sensor-measured chl } a] + 0.40 (n = 10, R^2 = 0.996, p < 0.001).$ 133

Water samples for dissolved nutrients (TDN, TDP, NO_3^- , NO_2^- , NH_4^+ and PO_4^{3-}) and DOC were immediately filtered through precombusted (450°C for 4 h) GF/F glass-fiber filters (Whatman) on the survey vessel, and the filtrate was frozen for later analysis of dissolved nutrients and DOC. Unfortunately, DOC samples stored in glass bottles collected in March were broken during the transport process, and 80% of DOC data in March was lost.

For measurements of bacterial production (BP), 40 mL of collected lake water was poured into 50-mL centrifuge tubes and incubated with 50 nM of $[^{15}N_5]$ -2'-deoxyadenosine ($^{15}N_{-141}$ dA, NLM-3895-PK, Cambridge Isotope Laboratories, Inc., Tewksbury, MA, USA) in the dark at *in situ* temperature for 3–8 hours depending on water temperature. The incubation was quenched

143 by adding 99.5% ethanol (special grade, Wako, Osaka, Japan) to the sample (final concentration,

>20%). After quenching, each water sample was filtered onto a 0.2-µm PTFE membrane filter

(Omnipore, Millipore, Burlington, MA, USA), and the filter was stored at -80°C until further
analysis.

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148 **2.2 Sample analysis**

Nutrients were analyzed in our laboratory using a continuous flow analyzer (SWAAT, 149 BLTEC) (Nojiri, 1987; Otsuki et al., 1993). DOC measurements were conducted as 150 nonpurgeable DOC with a TOC analyzer (TOC-V, Shimadzu) equipped with a Pt catalyst on 151 quartz wool. At least three measurements were made for each sample, and analytical precision 152 was typically less than 2%. Potassium hydrogen phthalate (Kanto Chemical) was used as a 153 standard. Additionally, molecular size distributions of DOC were measured by high-performance 154 size exclusion chromatography (HPSEC) with total organic carbon detectors, according to 155 Kawasaki et al., 2011; Shimotori et al., 2016. In the HPSEC analysis, there were two peaks of 156 weight-averaged molecular sizes (*Mw*) of $1.3 \pm 0.1 \times 10^5$ Da and $2.2 \pm 0.2 \times 10^3$ Da (*n* = 117), 157 and we defined them as high-molecular-weight DOC (HMW-DOC) and low-molecular-weight 158 DOC (LMW-DOC), respectively, in the present study. 159

For bacterial production (BP), bacterial DNA was extracted from the filter using the 160 Extrap Soil DNA Kit Plus ver. 2 (J-Bio21, NIPPON STEEL & SUMIKIN Eco-Tech Corp., 161 Chiba, Japan). Bacterial cells were broken by glass beads in a Fast Prep FP120 Cell Disrupter 162 (speed, 6.0; time, 40 s; MP Biomedicals, Santa Ana, CA, USA), and the released DNA was 163 purified by using magnetic beads, according to the manufacturer's protocols. The DNA 164 extraction efficiency was considered as 100% (Tsuchiya et al., 2019). After the DNA extraction, 165 the samples were processed according to Tsuchiya et al., (2015) and Tsuchiya et al., (2020a) for 166 quantifying the ¹⁵N-dA incorporation rates. Briefly. The DNA samples were hydrolyzed to 167 nucleosides by three enzymes (nuclease P1, Wako; phosphodiesterase I, Worthington 168 Biochemical Corp.; alkaline phosphatase, Promega Corp.). After the enzymatic hydrolysis, the 169 amount of ¹⁵N-dA incorporated during the incubation was analyzed by LC-MS/MS using 170

171 ${}^{13}C_{10}{}^{15}N_5$ -deoxyadenosine (CNLM-3896-CA, Cambridge Isotope Laboratories, Inc.) as a 172 surrogate (internal standard).

173	A cell-to-carbon conversion factor was obtained using the water sample collected at 5-m
174	depth of Sta. 12B in February 2018. Bacterial cells fixed with 1% glutaraldehyde were stained by
175	acridine orange, cell volumes were measured by epifluorescence microscopy (BX-40, Olympus,
176	Tokyo, Japan) with BX-FLA epifluorescence equipment, and the cell-to-carbon conversion
177	factor calculated using a conversion factor of 106 fgC μm^{-3} (Nagata, 1986), resulting in 11.8 fgC
178	cell ⁻¹ . The factor for converting 15 N-dA incorporation to cells produced was calculated as 1.83 \times
179	10^6 cells (mol ¹⁵ N-dA) ⁻¹ as determined by Tsuchiya et al., 2020b).
180	Bacterial abundance (BA) was estimated from measurements of 16S-rDNA
181	concentrations. The 16S-rDNA concentration was determined from the same extracted DNA
182	samples used for BP measurements. SYBR Green I PCR amplification was performed with a
183	LightCycler Instrument (Roche, Mannheim, Germany). For the real-time PCR assay of bacteria,
184	we used the primer set 357F (5'-CCTACGGGAGGCAGCAG-3'), 518R (5'-
185	GTATTACCGCGGCTGCTGG-3'). The 20-µL reaction mixture contained LightCycler 480
186	SYBR Green I Master (Roche), 4 pmol of each primer, and DNA extract. In the present study,
187	16S-rDNA concentration was converted to epifluorescence-enumerated bacterial cells (4',6-
188	diamidino-2-phenylindole, DAPI) by using a conversion factor of 0.31 cells $(16S-rDNA \text{ copy})^{-1}$
189	as previously determined (Tsuchiya et al., 2020b).

190

191 **3 Results**

192 **3.1. Bacterial production (BP)**

Bacterial production (BP) in 2018 varied from 0.057 μ gC L⁻¹ d⁻¹ at 40 m at Sta. 12B in November to 18.2 μ gC L⁻¹ d⁻¹ at 6 m at Sta. 12C in July (Fig. 2). BP was significantly positively correlated with water temperature (Spearman's rank correlation test, n = 104, p < 0.001, Fig. 3a). Variations of BP at certain water temperature levels (e.g., ca. 13–17°C and 28–32°C) were relatively high for the surface samples (< 10 m). In March, BP in the surface samples along Line 12 (1.48 ± 0.15 μ gC L⁻¹ d⁻¹, n = 3) was 2.4 times that of Lines 15 and 17 (0.62 ± 0.12 μ gC L⁻¹

 d^{-1} , n = 8). There was no clear trend of horizontal BP variations in the middle and deeper 199 samples (> 10 m). In May, BP in the surface samples was relatively high from the eastern 200 stations of Lines 12 and 15 (12BC, 12C, and 15C; $5.27 \pm 0.93 \ \mu \text{gC} \text{ L}^{-1} \text{ d}^{-1}$, n = 6). BP in the 201 surface samples along Line 17 did not show a clear horizontal trend (2.69 \pm 0.53 µgC L⁻¹ d⁻¹, n 202 = 4). Among the middle and deeper samples, BP at 20 m at western stations (12A, 15B, and 203 17A) was relatively high. In July in the surface samples, BP was notably lower at the central 204 stations of Lines 15 and 17 (15B and 17B); in other words, BP was relatively high at the stations 205 closer to shore. At all stations, BP at 20 m was higher than in the deeper samples (> 20 m). As in 206 May, BP at 19 m at Sta. 17A was highest among middle and deep samples from all stations. In 207 November in the surface samples, BP was relatively high at the eastern stations of Lines 12 and 208 15 (12C and 15C; $4.10 \pm 1.24 \ \mu \text{gC} \text{ L}^{-1} \text{ d}^{-1}$, n = 4); as in May, there was no clear horizontal trend 209 of BP along Line 17 (2.59 \pm 0.18 µgC L⁻¹ d⁻¹, n = 4). BP at 20 m increased compared to the 210

211 other months, but there was no clear horizontal trend.

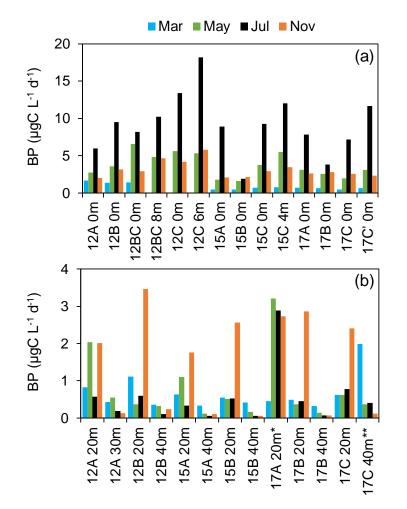


Fig. 2 Bacterial production (BP) in the north basin of Lake Biwa. Sampling depths were < 10 m
(a) and > 10 m (b). Blue, green, black, and orange bars represent the samples collected in March,

May, July, and November 2018, respectively. * and ** indicate that sampling depths were 19 m in July and 39 m in March, respectively.

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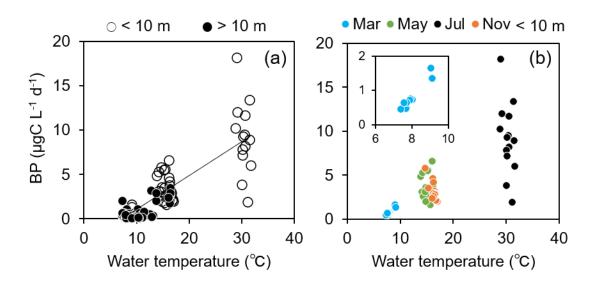




Fig. 3 Bacterial production (BP) versus water temperature in the north basin of Lake Biwa,
Japan. (a) Open and filled circles represent < 10 m and > 10 m depths, respectively. (b) Blue,
green, black, and orange circles represent the samples collected in March, May, July, and
November 2018, respectively, in the surface waters (< 10-m depth).

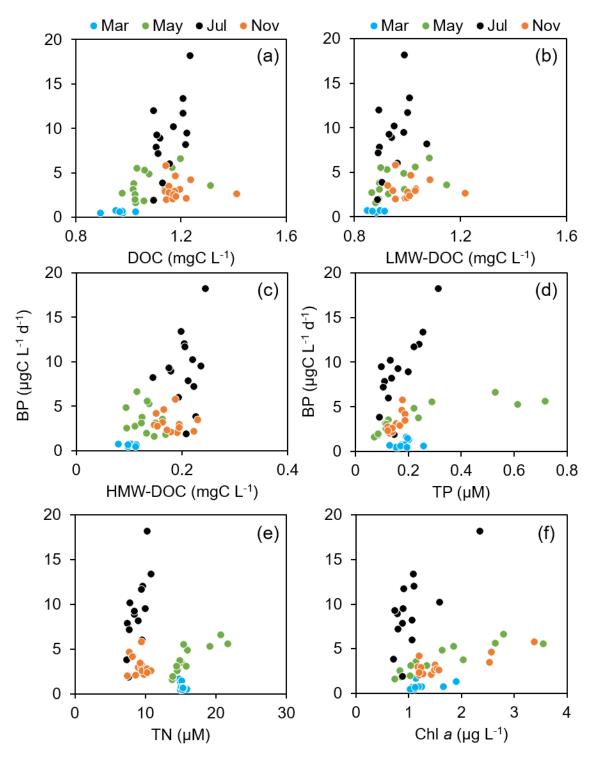
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3.2 Relationships between bacterial production and environmental variables at the surface

In the surface samples in March, BP was significantly positively correlated with water temperature (p < 0.01, Fig. 3b, Table 2), whereas BP was negatively correlated to water temperature in November. In other seasons, there were no significant relationships between BP and water temperature. As to DOC, LMW-DOC, and HMW-DOC, BP was significantly positively correlated to them in year-round (Fig. 4a-c, Table 2). However, BP in each season did not correlate to them significantly. In the surface samples, BP was significantly correlated with TP in May, July, and November (Fig. 4d, Table 2). Although BP did not significantly correlate to

- 232 TP in year-round, there was a significant relationship between BP and TP when winter data were
- excluded (Year-round without winter, Table 2). BP was correlated to TN in May and July (Fig.
- 4e, Table 2). BP was correlated to Chl *a* in March, May, and July (Fig. 4f, Table 2).
- There were significantly positive regressions between BA and BP in each season's
- surface samples (Fig. 5). The slopes of the linear regressions were steepest in March (0.81),
- almost the same in May and November (0.30 and 0.29, respectively), and gentlest in July (0.12).
- 238
- Table 2 Summary of spearman's rank correlation coefficients (rs) between bacterial production
- (BP) and environmental variables in the epilimnion (< 10-m depth). WT; water temperature,
- 241 DOC; dissolved organic carbon, LMW-DOC; low-molecular-weight DOC, HMW-DOC; high-
- 242 molecular-weight DOC, TP; total phosphorus, TN; total nitrogen, Chl *a*: chlorophyll *a*, n.s.; not
- 243 significant

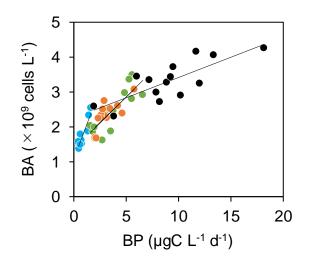
Season		WТ	DOC	LMW-DOC	HMW-DOC	TP	TN	Chl a
Winter	rs	0.939	0.143	0.314	-0.200	0.318	-0.518	0.794
(March)	n	10	6	6	6	11	11	10
	р	0.0048	n.s.	n.s.	n.s.	n.s.	n.s.	0.0172
Spring	rs	0.015	0.486	0.530	0.068	0.930	0.925	0.925
(May)	n	14	14	14	14	14	14	14
	р	n.s.	n.s.	n.s.	n.s.	<0.001	<0.001	<0.001
Summer	rs	-0.305	0.486	0.530	0.068	0.710	0.758	0.675
(July)	n	14	14	14	14	14	14	14
	р	n.s.	n.s.	n.s.	n.s.	0.0105	0.0063	0.0150
Autumn	rs	-0.727	-0.054	0.110	-0.214	0.811	-0.182	0.464
(November)	n	14	14	13	13	14	14	14
	р	0.0087	n.s.	n.s.	n.s.	0.0035	n.s.	n.s.
Year-round	rs	0.689	0.498	0.394	0.509	0.176	-0.382	0.067
	n	52	48	47	47	53	53	52
	р	<0.001	<0.001	0.0076	<0.001	n.s.	0.0059	n.s.
Year-round	rs	0.412	0.268	0.164	0.302	0.477	-0.098	0.078
without winter	n	42	42	41	41	42	42	42
	р	0.0083	n.s.	n.s.	n.s.	0.0022	n.s.	n.s.



245

Fig. 4 Bacterial production (BP) versus (a) dissolved organic carbon (DOC) concentration, (b)
low-molecular-weight DOC (LMW-DOC) concentration, (c) high-molecular-weight DOC
(HMW-DOC) concentration, (d) total phosphorus (TP) concentration, (e) total nitrogen (TN)
concentration, and (f) chlorophyll *a* concentration (Chl *a*) in the surface waters (< 10-m depth) in

- the north basin of Lake Biwa, Japan. Blue, green, black, and orange circles represent the samples
- collected in March, May, July, and November 2018, respectively.
- 252



253

Fig. 5 Bacterial abundance (BA) versus bacterial production (BP) in surface waters of the north basin of Lake Biwa, Japan, in March, May, July and November 2018 (blue, green, black and orange circles, respectively). Regression lines for March: [BA] = $0.81 \times [BP] + 1.1$ ($n = 10, R^2 =$ 0.84, p < 0.001); May: [BA] = $0.30 \times [BP] + 1.4$ ($n = 14, R^2 = 0.68, p < 0.001$); July: [BA] = $0.12 \times [BP] + 2.3$ ($n = 14, R^2 = 0.62, p < 0.001$); and November: [BA] = $0.29 \times [BP] + 1.4$ (n = $14, R^2 = 0.60, p < 0.01$). The regression lines for May and November are almost indistinguishable.

262 **3.3 Depth-integrated BP**

Depth-integrated BP (DBP) ranged from 2.0 mgC $m^{-2} d^{-1}$ at Sta. 17C' in March to 110 263 mgC $m^{-2} d^{-1}$ at Sta. 12B in July (Table 3). In March and November, DBP significantly increased 264 with maximum water depth (Fig. 6a, d). Although the regression of DBP on maximum depth for 265 May was not significant, when the three transects were analyzed together, the regressions were 266 significant when the Line 12 data were analyzed separately from those for Lines 15 and 17 (Fig. 267 6b). For July, there was no significant regression (Fig. 6c). No clear trends were found in the 268 relationship between DBP and distance from shore in March, May, and November (Fig. 6e, f, h). 269 In July, DBP at Sts. 15B and 17B (> 4 km offshore) were relatively low compared to the stations 270 closer to shore (Table 3, Fig. 6g). The inshore/offshore ratio of DBP was lowest in November 271

272 (0.58) and highest in July (2.1) (Table 3).

273

- Table 3 Depth-integrated bacterial production (BP) in Lake Biwa in 2018. Stations 15B and 17B
- are classified as offshore (> 4 km from shore) and the others as inshore.

Station	Average	Distance from	Depth-ir	ntegrated Bl	P (mgC m⁻	² d ⁻¹)
Station	depth (m)	shore (km)	Mar	May	Jul	Nov
12A	33.1	0.8	32.3	80.5	70.0	51.1
12B	60.5	3.2	46.5	86.7	110.4	103.7
12BC	8.9	1.6	12.8	49.1	84.7	35.0
12C	6.9	1.0	ND	40.6	109.1	32.2
15A	76.0	1.3	33.4	31.5	98.2	58.4
15B	62.5	6.7	28.6	43.4	31.8	75.0
15C	6.4	0.6	3.9	25.7	85.7	24.7
17A	26.3	0.9	23.0	36.7	104.0	51.5
17B	88.4	8.0	35.4	41.8	51.6	92.0
17C	41.9	0.6	38.6	37.3	92.3	74.3
17C'	3.9	0.5	2.0	12.9	43.2	8.2
Offshore (15B & 17B)	average	32.0	42.6	41.7	83.5
Inshore av	erage	-	24.1	44.6	88.6	48.8
Inshore/Of	ffshore ratio		0.75	1.0	2.1	0.58

ND: No data

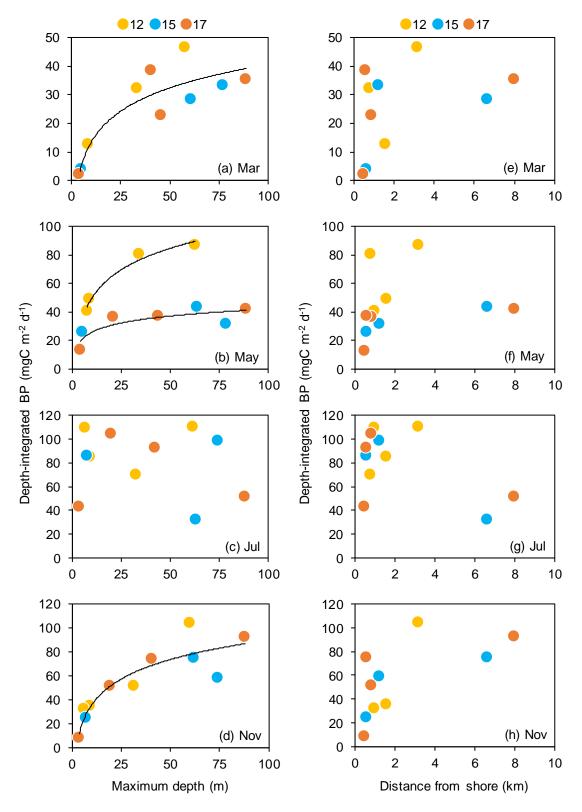




Fig. 6 Depth-integrated bacterial production (DBP) in each season versus maximum depth (left)
or distance from shore (right) in the north basin of Lake Biwa, Japan. Yellow, blue and orange

plots represent Line 12, 15 and 17 transects, respectively. The regression curves represent (a)

281 [DBP] = $11.7 \times \ln [\text{depth}] - 13.4 (n = 10, R^2 = 0.81, p < 0.001);$ (b) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81, p < 0.001);$ (b) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81, p < 0.001);$ (b) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81, p < 0.001);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81, p < 0.001);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81, p < 0.001);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81, p < 0.001);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81, p < 0.001);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81, p < 0.001);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81, p < 0.001);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81, p < 0.001);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81, p < 0.001);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81, p < 0.001);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81, p < 0.001);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81, p < 0.001);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81, p < 0.001);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10, R^2 = 0.81);$ (c) [DBP_Line12] = $21.7 \times \ln (n = 10,$

282 [depth] + 0.0885 ($n = 4, R^2 = 0.98, p < 0.05$) and [DPB_Lines15&17] = 7.02 × ln [depth] + 9.78

283 $(n = 7, R^2 = 0.71, p < 0.05)$; and (d) [DBP] = 23.4 × ln [depth] - 18.1 $(n = 11, R^2 = 0.82, p < 1.5)$

284 0.001).

285

286 4 Discussion

The present study examined spatial and seasonal variations of BP and environmental 287 variables in the Lake Biwa and found a significantly positive relationship between BP and water 288 temperature in year-round data, suggesting that water temperature was a primary BP regulation 289 factor. This relationship agrees with previous studies (Goosen et al., 1997; Pomeroy & Wiebe, 290 2001). However, the coefficient of variance (CV) of BP (0.35-0.49) was much larger than that of 291 water temperature (0.030-0.075) at the surface in each season; in other words, the BP variation 292 was more considerable than that of water temperature. The large BP variations in each season 293 were attributed to the horizontal distributions. The DOC, LMW-DOC, and HMW-DOC 294 concentrations correlated to BP in year-round data, whereas there were no significant 295 relationships in each season (Table 2), suggesting that DOC could not be an explanatory factor 296 for horizontal BP variations. The other environmental variables showed significant correlations 297 298 to BP in each season and can be explanatory factors for BP horizontal variations.

In March, BP in the surface samples sensitively responded to variations in water 299 300 temperature (Fig. 4a, Table 2); however, BP was correlated with neither TP nor TN concentrations. The chl *a* concentration was correlated to water temperature (rs = 0.79, n = 10, p 301 < 0.05) as well as BP, suggesting that water temperature could be the primary limiting factor for 302 both bacterioplankton and phytoplankton in winter. The importance of water temperature 303 affecting bacterial growth, especially during winter, was also emphasized in estuarine and marine 304 environments (e.g. Shiah & Ducklow, 1994a, b). The slope of the linear regression of BA on BP 305 was highest in March (Fig. 5). The relationship between BA and BP has been used to evaluate 306 bottom-up and top-down regulation (Billen et al., 1990; Pace & Cole, 1994). The regressions of 307 308 BA as a function of BP should have a steep slope if BA is strongly controlled by bottom-up

factors such as resources and water temperature. Alternatively, when other factors such as
mortality are most important in regulating BA, there should be no relationships or shallower
slopes between them. The ingestion rates of bacteria by heterotrophic nanoflagellates, *Daphnia galeata*, and *Bosmina longirostris* in Lake Biwa were relatively low in March (Nakano et al.,
1998). Therefore, in March, bacteria should have been regulated by bottom-up control, especially
by water temperature.

The latitudinal difference of water temperature (Line 12 > Lines 15 and 17) could be 315 316 attributed to Lake Biwa's climate conditions. Air temperatures in 20-23 March 2018, including sampling days, at weather observation stations of the Japan Meteorological Agency around Lake 317 318 Biwa (https://www.jma.go.jp/jma/, last accessed on 6 March 2020) varied along a latitudinal gradient, i.e., Line 12 > Line 15 > Line 17 in this period (see Supporting Information, Fig. A1). 319 The results suggest that climate conditions such as air temperature clearly linked to the 320 horizontal distribution of water temperature and consequently affected biological productivity 321 within the lake in winter. 322

In May, BP was significantly positively correlated with TP, TN, and chl a concentrations 323 (Fig. 4, Table 2). These results suggest that the effect of resource limitation on BP became more 324 pronounced. The spatial distribution of the loading of nutrients, organic matter, or both affects 325 the horizontal BP variation. It is related to higher BP at eastern stations of Lines 12 and 15 in the 326 327 surface samples. Nutrient discharges into Lake Biwa by rivers flowing into the eastern area of the lake were larger than those in the western area, which is attributed to the high population 328 densities in the urbanized areas and many agricultural fields east of the lake (Taniguchi & Tase, 329 330 1999). Since TP and TN concentrations included bacterial biomass as a particulate form and 331 there was a significantly positive relationship between BP and BA (Fig. 5), the correlations with TP and TN did not necessarily indicate limiting factors for BP. The relationship between BP and 332 333 various forms of N and P (dissolved/particulate and inorganic/organic) was significant only with particulate nitrogen for N, but with dissolved total/organic phosphorus, PO4³⁻, and particulate 334 phosphorus for P (Table A1). The results suggest that the horizontal variations of BP could have 335 been regulated by phosphorus as their resource. 336

In July, the BP variations in the surface samples were larger than that in other seasons 337 (Fig. 3). A similar larger BP variation was observed in another lake (Lake Kasumigaura, the 338 second largest lake in Japan) at higher water temperatures; the variations of BP in Lake 339 Kasumigaura were attributed to autochthonous substrate supply of dissolved organic carbon from 340 primary production (Tsuchiya et al., 2019). In the present study, BP was significantly correlated 341 not to DOC, but rather to TP and TN concentration, suggesting that the large BP variations 342 should be attributed to phosphorus and/or nitrogen limitations. Since PO₄³⁻ concentrations were 343 below the detection limit in this season except for Sts. 12BC and 12C, we examined whether 344 particulate or dissolved P was the dominant source for BP (Fig. 7). Particulate P explained 64% 345 of the BP variations, although particulate P partially comprises bacteria, while dissolved P 346 explained 31% of the variation. However, we found obvious outliers (the two lowest BP values) 347 in the relationship between BP and dissolved P at Sts. 15B and 17B. When the two outliers were 348 excluded from the regression analysis, dissolved P also explained BP's variations (69%). The 349 results for these outliers suggest that particulate P should be a dominant P source for BP at the 350 surface of these offshore stations (15B and 17B). Besides, dissolved P could be transformed into 351 352 refractory substrates by photochemical and/or biological reactions. Exposure of surface-water dissolved organic matter (DOM) to sunlight led to a 75% reduction of BP in marine ecosystems, 353 354 suggesting that the photomineralization of bioreactive DOM likely contributed to the decrease in BP (Benner & Biddanda, 1998). Further study is needed to clarify the relationship between BP 355 356 and refractory dissolved organic P and material cycling in such strongly P-limited environments. We uniformly observed relatively high BP in inshore stations, which we attribute to nutrient 357 loading via some combination of river inflow, groundwater seepage, and elution from sediments. 358 As for nitrogen, there were no significant relationships between BP and any dissolved forms of N 359 other than particulate nitrogen as well as the case in spring (Table A1), suggesting that N was not 360 likely to be a limiting factor for BP. The result is consistent with a previous study demonstrating 361 that bacterial growth at the surface in Lake Biwa was severely limited by phosphorus, but not 362 nitrogen and organic carbon (Gurung & Urabe, 1999). 363

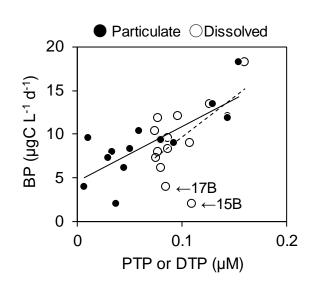
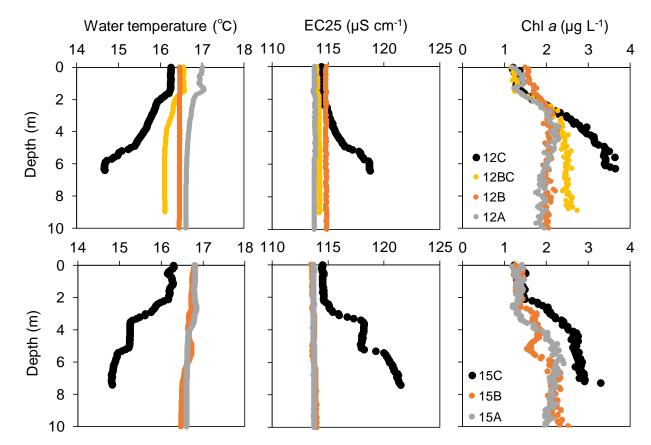
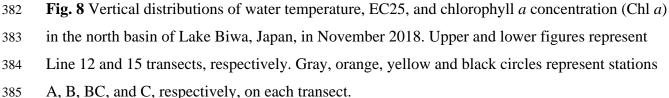




Fig. 7 Bacterial production (BP) versus particulate total phosphorus (PTP) and dissolved total phosphorus (DTP) concentrations in the surface waters of the north basin of Lake Biwa, Japan, in July 2018. The regression lines represent [BP] = $62 \times [PTP] + 4.6$ ($n = 14, R^2 = 0.64, p < 0.001$) and [BP] = $92 \times [DTP] + 0.44$ ($n = 14, R^2 = 0.31, p < 0.05$).

In November, BP was positively correlated with TP and chl *a* concentrations (Fig. 4). 371 Besides, BP was relatively high at eastern stations such as 12C and 15C (Fig. 3a). Compared to 372 central and western stations along Lines 12 and 15, water temperatures at the eastern Sts. 12C 373 374 and 15C were lower at depth, while specific conductivity (EC25) and chl a were relatively high (Fig. 8). EC25 can be used as a proxy for inflows of different water masses such as river water 375 and groundwater (Bischof et al., 2019). The concentrations of nutrients and dissolved organic 376 carbon are higher in river water and groundwater in the Lake Biwa watershed than in lake water 377 (Taniguchi & Tase, 1999; Mostofa et al., 2007). Thus, the results suggest that allochthonous 378 nutrients and organic matter entering predominantly from the eastern shore enhanced bacterial 379 activity directly, indirectly via phytoplankton activity, or both. 380





Our observation of a horizontal gradient of BP from the shore to the central waters, 386 especially in summer, is consistent with results from Lakes Michigan (Scavia & Laird, 1987) and 387 Constance (Güde, 1990). Ratios of maximum (max) over minimum (min) BP in horizontal 388 variability ranged from 1.6 to 19.5 (Table 1). Although the max/min ratio in Lake Biwa was 389 within this range $(5.2 \pm 3.0, n = 4, \text{Table 1})$, there were relatively high values of 4.1 in spring and 390 winter and 9.5 in summer. Reichart and Simon argued that the spatial variability is in the same 391 range as that of the diurnal range of bacterial growth dynamics (Reichart & Simon, 1996). The 392 ranges of BP diel variations were 1.8-3.8 (2.6 ± 0.8 , n = 4) in Lake Constance (Simon, 1994), 393 $1.3-5.0 (2.8 \pm 1.3, n = 8)$ in the Mediterranean Sea (Gasol et al., 1998), 1.7-2.5 (n = 2) in Lake 394 Michigan (Reichart & Simon, 1996) and 2.9 (n = 1) in Lake Hallwil (Filippini et al., 2008). 395 Although the diel BP variation was not assessed in the present study, the relatively high max/min 396

ratios for horizontal variation (~9.5) seem larger than the range of diel variation shown above.
Moreover, the variations of BP in each season were related to the variations of environmental
variables (Figs. 4, 7, and 8). Especially in Lake Biwa, BP is severely limited by phosphorus, but
not organic carbon (Gurung & Urabe, 1999). The results suggest that nutrient supply processes
strongly influence the horizontal variability of BP in this lake.

The average DBP of inshore stations was 2.1 times that of offshore stations in July, and 402 the inshore/offshore ratio was higher than those in the other seasons (0.58-1.0; Table 3). To 403 404 estimate the relative contribution of inshore BP to the whole north basin of Lake Biwa (about 450 km² for inshore and 168 km² for offshore), the average DBP for inshore and offshore 405 stations was multiplied by the respective areas. In July, the integrated inshore and offshore BP 406 values were 39.9 tC d^{-1} and 7.0 tC d^{-1} , respectively, suggesting that the inshore BP accounted for 407 85% of whole-lake BP (46.9 tC d⁻¹). The inshore BP in March, May, and November accounted 408 for 66.9%, 73.8%, and 61.1% of the whole-lake BP, respectively; for these months, the 409 percentage contributions of inshore waters were equivalent to or lower than the percentage of the 410 inshore area $(72.9\% = 450 \text{ km}^2/618 \text{ km}^2)$ and disproportionate contributions of inshore BP was 411 412 not confirmed. The result agrees with the results shown in Fig. 6: DBP in March, May, and 413 November were better explained not by the distance from shore but rather by the maximum depth. The results demonstrate that inshore BP disproportionately contributed to whole-lake BP 414 in the most productive season (July). 415

In conclusion, we quantified horizontal heterogeneity of BP in each season in Lake Biwa: 416 BP was regulated by primarily water temperature and secondarily phosphorus availability. 417 Dissolved organic carbon, including LMW- and HMW-DOC, did not link to horizontal 418 distributions of BP in the surface waters. The horizontal BP heterogeneity in the surface waters 419 was largest in summer (max/min ratio ~9.5), and minimum values were measured at offshore 420 421 stations. Moreover, mean DBP at inshore stations was 2.1 times that of offshore stations, 422 clarifying the substantial contribution of inshore to whole-lake BP, especially in the most productive season. 423

424

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- 432

433 **References**

- 434 Azam, F. (1998), Microbial Control of Oceanic Carbon Flux: The Plot Thickens, Science,
- 435 280(5364), 694-696, doi:10.1126/science.280.5364.694.
- 436 Benner, R., and B. Biddanda (1998), Photochemical transformations of surface and deep marine

dissolved organic matter: Effects on bacterial growth, *Limnol. Oceanogr.*, 43(6), 1373-1378,

- 438 doi:10.4319/lo.1998.43.6.1373.
- Billen, G., P. Servais, and S. Becquevort (1990), Dynamics of bacterioplankton in oligotrophic
- 440 and eutrophic aquatic environments: bottom-up or top-down control?, *Hydrobiologia*, 207(1),
- 441 37-42, doi:10.1007/BF00041438.
- 442 Bischof, S. M., B. R. Herwig, S. D. Sebestyen, M. A. Hanson, K. D. Zimmer, J. B. Cotner, and
- 443 T. J. Kroeger (2019), Further Development of a Specific Conductivity Approach to Measure
- 444 Groundwater Discharge Area within Lakes, *Journal of the American Water Resources*
- 445 Association, 55(2), 485-496, doi:10.1111/1752-1688.12730.
- 446 Filippini, M., N. Buesing, and M. O. Gessner (2008), Temporal dynamics of freshwater bacterio-
- 447 and virioplankton along a littoral-pelagic gradient, *Freshw. Biol.*, 53(6), 1114-1125,
- 448 doi:10.1111/j.1365-2427.2007.01886.x.
- 449 Güde, H. (1990), Bacterial Production and the Flow of Organic Matter in Lake Constance, in
- 450 Large Lakes: Ecological Structure and Function, edited by M. M. Tilzer and C. Serruya, pp.
- 451 489-502, Springer Berlin Heidelberg, Berlin, Heidelberg, doi:10.1007/978-3-642-84077-7_25.
- 452 Gasol, J. M., M. D. Doval, J. Pinhassi, J. I. Calderón-Paz, N. Guixa-Boixareu, D. Vaqué, and C.
- 453 Pedrós-Alió (1998), Diel variations in bacterial heterotrophic activity and growth in the
- northwestern Mediterranean Sea, *Mar. Ecol. Prog. Ser.*, *164*, 107-124, doi:10.3354/meps164107.

- 455 Goosen, N. K., P. van Rijswijk, J. Kromkamp, and J. Peene (1997), Regulation of annual
- 456 variation in heterotrophic bacterial production in the Schelde estuary (SW Netherlands), Aquat.
- 457 *Microb. Ecol.*, *12*(3), 223-232, doi:10.3354/ame012223.
- 458 Gurung, T. B., and J. Urabe (1999), Temporal and Vertical Difference in Factors Limiting
- 459 Growth Rate of Heterotrophic Bacteria in Lake Biwa, *Microb. Ecol.*, 38(2), 136-145,
- 460 doi:10.1007/s002489900167.
- 461 Gurung, T. B., J. Urabe, K. Nozaki, C. Yoshimizu, and M. Nakanishi (2002), Bacterioplankton
- 462 production in a water column of Lake Biwa, Lakes and Reservoirs: Research and Management,
- 463 7(4), 317-323, doi:10.1046/j.1440-1770.2002.00197.x.
- Hayakawa, K. (2004), Seasonal variations and dynamics of dissolved carbohydrates in Lake
- 465 Biwa, *Organic Geochemistry*, *35*(2), 169-179, doi:10.1016/j.orggeochem.2003.09.002.
- 466 Kawasaki, N., K. Matsushige, K. Komatsu, A. Kohzu, F. W. Nara, F. Ogishi, M. Yahata, H.
- 467 Mikami, T. Goto, and A. Imai (2011), Fast and precise method for HPLC-size exclusion
- 468 chromatography with UV and TOC (NDIR) detection: importance of multiple detectors to
- 469 evaluate the characteristics of dissolved organic matter, *Water Res*, 45(18), 6240-6248,
- 470 doi:10.1016/j.watres.2011.09.021.
- 471 Mostofa, K. M., T. Yoshioka, E. Konohira, and E. Tanoue (2007), Dynamics and characteristics
- 472 of fluorescent dissolved organic matter in the groundwater, river and lake water, *Water Air Soil*
- 473 *Pollut.*, 184(1-4), 157-176, doi:10.1007/s11270-007-9405-1.
- 474 Nagata, T. (1986), Carbon and nitrogen content of natural planktonic bacteria, *Appl. Environ.*475 *Microbiol.*, 52(1), 28-32.
- 476 Nagata, T. (1987), Production rate of planktonic bacteria in the north basin of lake biwa, Japan,
- 477 *Appl. Environ. Microbiol.*, *53*(12), 2872-2882, doi:10.1128/AEM.53.12.2872-2882.1987.
- 478 Nakano, S.-i., T. Koitabashi, and T. Ueda (1998), Seasonal changes in abundance of
- heterotrophic nanoflagellates and their consumption of bacteria in Lake Biwa with special
- reference to trophic interactions with Daphnia galeata, *Archiv fur Hydrobiologie*, *142*(1), 21-34.
- 481 Nojiri, Y. (1987), Progress in water quality analysis (3), Journal of Groundwater Hydrology,
- 482 *29*(2), 107-111, doi:10.5917/jagh1987.29.107.

- 483 Otsuki, A., R. Goma, M. Aizaki, and Y. Nojiri (1993), Seasonal and spatial variations of
- 484 dissolved nitrogenous nutrient concentrations in hypertrophic shallow lake, with special
- 485 reference to dissolved organic nitrogen, Internationale Vereinigung für theoretische und
- 486 angewandte Limnologie: Verhandlungen, 25(1), 187-192,
- 487 doi:10.1080/03680770.1992.11900089.
- 488 Pace, M. L., and J. J. Cole (1994), Comparative and experimental approaches to top-down and
- 489 bottom-up regulation of bacteria, *Microb. Ecol.*, *28*(2), 181-193, doi:10.1007/BF00166807.
- 490 Pomeroy, L. R., and W. J. Wiebe (2001), Temperature and substrates as interactive limiting
- 491 factors for marine heterotrophic bacteria, *Aquat. Microb. Ecol.*, 23(2), 187-204,
- 492 doi:10.3354/ame023187.
- 493 Ram, A. S. P., Y. Nishimura, Y. Tomaru, K. Nagasaki, and T. Nagata (2010), Seasonal variation
- in viral-induced mortality of bacterioplankton in the water column of a large mesotrophic lake
- 495 (Lake Biwa, Japan), Aquat. Microb. Ecol., 58(3), 249-259, doi:10.3354/ame01381.
- Reichart, I., and M. Simon (1996), Horizontal variability of bacterioplankton growth dynamics in
 a large lake, *Aquat. Microb. Ecol.*, *11*, 31-41, doi:10.3354/ame011031.
- 498 Reitner, B., A. Herzig, and G. J. Herndl (1999), Dynamics in bacterioplankton production in a
- 499 shallow, temperate lake (Lake Neusiedl, Austria): evidence for dependence on macrophyte
- production rather than on phytoplankton, Aquat. Microb. Ecol., 19(3), 245-254,
- 501 doi:10.3354/ame019245.
- 502 Scavia, D., and G. A. Laird (1987), Bacterioplankton in Lake Michigan: Dynamics, controls, and
- significance to carbon flux, *Limnol. Oceanogr.*, 32(5), 1017-1033,
- 504 doi:10.4319/lo.1987.32.5.1017.
- 505 Shiah, F. K., and H. W. Ducklow (1994a), Temperature and Substrate Regulation of Bacterial
- 506 Abundance, Production and Specific Growth-Rate in Chesapeake Bay, USA, Mar. Ecol. Prog.
- 507 Ser., 103(3), 297-308, doi:10.3354/meps104297.
- 508 Shiah, F. K., and H. W. Ducklow (1994b), Temperature regulation of heterotrophic
- 509 bacterioplankton abundance, production, and specific growth rate in Chesapeake Bay, *Limnol*.
- 510 Oceanogr., 39(6), 1243-1258, doi:10.4319/lo.1994.39.6.1243.

- 511 Shiga-Prefecture (2017), Environmental white paper in 2017.
- 512 <u>https://www.pref.shiga.lg.jp/file/attachment/4016451.pdf</u>. Accessed 1 Feb 2019.
- 513 Shimotori, K., T. Satou, A. Imai, N. Kawasaki, K. Komatsu, A. Kohzu, N. Tomioka, R.
- 514 Shinohara, and S. Miura (2016), Quantification and characterization of coastal dissolved organic
- 515 matter by high-performance size exclusion chromatography with ultraviolet absorption,
- fluorescence, and total organic carbon analyses, *Limnology and Oceanography: Methods*,
- 517 *14*(10), 637-648, doi:10.1002/lom3.10118.
- 518 Simon, M. (1994), Diel Variability of Bacterioplankton Biomass Production and Cell
- 519 Multiplication in Lake Constance, *Archiv Fur Hydrobiologie*, *130*(3), 283-302.
- 520 Tanentzap, A. J., et al. (2017), Terrestrial support of lake food webs: Synthesis reveals controls
- 521 over cross-ecosystem resource use, *Sci Adv*, *3*(3), e1601765, doi:10.1126/sciadv.1601765.
- 522 Taniguchi, M., and N. Tase (1999), Nutrient discharge by groundwater and rivers into Lake
- 523 Biwa, Japan, IAHS PUBLICATION, 67-74.
- 524 They, N. H., and D. d. M. Marques (2019), The structuring role of macrophytes on plankton
- 525 community composition and bacterial metabolism in a large subtropical shallow lake, *Acta*
- 526 Limnologica Brasiliensia, 31, doi:10.1590/s2179-975x10017
- 527 Tsuchiya, K., T. Sano, N. Kawasaki, H. Fukuda, N. Tomioka, K. Hamasaki, Y. Tada, S.
- 528 Shimode, T. Toda, and A. Imai (2015), New radioisotope-free method for measuring bacterial
- 529 production using [15N5]-2'-deoxyadenosine and liquid chromatography mass spectrometry (LC-
- 530 MS) in aquatic environments, J. Oceanogr., 71(6), 675-683, doi:10.1007/s10872-015-0310-8.
- 531 Tsuchiya, K., T. Sano, N. Tomioka, A. Kohzu, K. Komatsu, R. Shinohara, S. Shimode, T. Toda,
- and A. Imai (2020a), Incorporation characteristics of exogenous 15N-labeled thymidine,
- deoxyadenosine, deoxyguanosine and deoxycytidine into bacterial DNA, *PLoS One*, 15(2),
- ⁵³⁴ e0229740, doi:10.1371/journal.pone.0229740.
- 535 Tsuchiya, K., et al. (2019), Seasonal variability and regulation of bacterial production in a
- shallow eutrophic lake, *Limnol. Oceanogr.*, 64(6), 2441-2454, doi:10.1002/lno.11196.
- 537 Tsuchiya, K., N. Tomioka, T. Sano, A. Kohzu, K. Komatsu, A. Imai, K. Hayakawa, T. Nagata,
- 538 T. Okamoto, and Y. Hirose (2020b), Decrease in bacterial production over the past three decades

- 539 in the north basin of Lake Biwa, Japan, *Limnology*, 21(1), 87-96, doi:10.1007/s10201-019-
- 540 00582-2.
- 541 Wylie, J. L., and D. J. Currie (1991), The Relative Importance of Bacteria and Algae as Food
- 542 Sources for Crustacean Zooplankton, *Limnol. Oceanogr.*, *36*(4), 708-728,
- 543 doi:10.4319/lo.1991.36.4.0708.

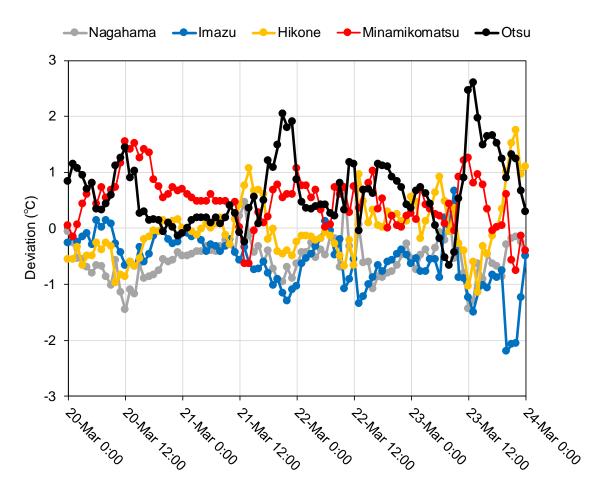
544 **Table A1** Summary of spearman's rank correlation coefficients (rs) between bacterial production

- 545 (BP) and environmental variables at the surface samples (< 10 m). PN; particulate nitrogen,
- 546 DTN; dissolved total nitrogen, DON; dissolved organic nitrogen, PP; particulate phosphorus,
- 547 DTP; dissolved total phosphorus, DOP; dissolved organic phosphorus, n.s.; not significant, ND:
- 548 not determined

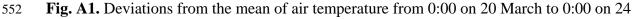
Season		PN	DTN	NO ₂₊₃	NH_4	DON	PP	DTP	PO_4	DOP
Spring (May)	rs n p	0.864 14 <0.001	0.4945 14 n.s.	0.4593 14 n.s.	-0.1077 14 n.s.	0.4989 14 n.s.	0.947 14 <0.001	0.692 14 0.0061	0.679 14 0.0075	0.684 14 0.0070
Summer (July)	rs n p	0.6689 14 0.0089	-0.1209 14 n.s.	-0.4158 14 n.s.	0.2967 14 n.s.	-0.0945 14 n.s.	0.7846 14 0.0009	0.3055 14 n.s.	ND	0.3275 14 n.s.

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553 March in 2018 observed at five weather observation stations around Lake Biwa, Japan: Imazu,

- 554 Nagahama (near Line 17), Hikone (near Line 15), Minamikomatsu (near Line 12) and Otsu (near
- south basin). The altitudes above sea level of these stations were similar (86–95 m). There were
- significant differences between the deviations among the stations: Nagahama = Imazu < Hikone
- 557 < Minamikomatsu = Otsu (Tukey-Kramer's multi comparison test; total n = 485, p < 0.01). The
- data were obtained from the Japan Meteorological Agency (https://www.jma.go.jp/jma/, last
- accessed on 6 March 2020).