Climate change impacts on mycorrhizae amplify nitrogen limitation on global plant growth

Renato K. Braghiere^{1,1}, Joshua Fisher^{2,2}, Rosie A. Fisher^{3,3}, Mingjie Shi^{4,4}, Brian N Steidinger^{5,5}, Benjamin N Sulman^{6,6}, Nadia Soudzilovskaia^{7,7}, Xiaojuan Yang^{8,8}, Jingjing Liang^{9,9}, Kabir G Peay^{5,5}, Thomas W Crowther^{10,10}, and Richard P. Phillips^{11,11}

¹NASA Jet Propulsion Laboratory
²Jet Propulsion Lab
³National Center for Atmospheric Research (UCAR)
⁴Jet Propulsion Lab (NASA)
⁵Stanford University
⁶Oak Ridge National Laboratory
⁷Leiden University
⁸Oak Ridge National Lab
⁹Purdue University
¹⁰ETH Zürich
¹¹Indiana University Bloomington

November 30, 2022

Abstract

Most tree species predominantly associate with a single type of mycorrhizal fungi, which can differentially affect plant nutrient acquisition and biogeochemical cycling. Here, we address for the first time the impact of mycorrhizal distributions on global carbon and nutrient cycling. Using the state-of-the-art carbon-nitrogen economics within the Community Land Model version 5 (CLM5) we found Net Primary Productivity (NPP) increased throughout the 21st century by 20%; however, as soil nitrogen has progressively become limiting, the costs to NPP for nitrogen acquisition — i.e., to mycorrhizae — have increased at a faster rate by 60%. This suggests that nutrient acquisition will increasingly demand a higher portion of assimilated carbon to support the same productivity. Uncertainties in mycorrhizal distributions are non-trivial, however, with uncertainties in NPP by up to 345 Tg C yr-1, depending on which published distribution is used. Remote sensing capabilities for mycorrhizal detection show promise for refining these estimates further.

1	Mycorrhizal distributions impact global patterns of carbon and nutrient cycling
2	R. K. Braghiere ^{1,2†} , J. B. Fisher ^{1,2} , R. A. Fisher ^{3,4} , M. Shi ^{1,2} , B. S. Steidinger ⁵ , B. N.
3	Sulman ⁶ , N. A. Soudzilovskaia ⁷ , X. Yang ⁶ , J. Liang ^{8,9} , K. G. Peay ⁵ , T. W. Crowther ¹⁰ , R. P.
4	Phillips ¹¹
5	¹ Jet Propulsion Laboratory, California Institute of Technology, 4800 Oak Grove Drive,
6	Pasadena, CA, 91109 USA.
7	² Joint Institute for Regional Earth System Science and Engineering, University of California at
8	Los Angeles, Los Angeles, CA, 90095 USA.
9	³ Climate and Global Dynamics Division, National Center for Atmospheric Research, Boulder,
10	CO, USA.
11	⁴ Laboratoire Évolution & Diversité Biologique, CNRS:UMR 5174, Université Paul Sabatier,
12	Toulouse, France.
13	⁵ Department of Biology, Stanford University, Stanford, CA, USA.
14	⁶ Environmental Sciences Division and Climate Change Science Institute, Oak Ridge National
15	Laboratory, Oak Ridge, TN, USA.
16	⁷ Environmental Biology Department, Institute of Environmental Sciences, Leiden University,
17	Leiden, The Netherlands.
18	⁸ Department of Forestry and Natural Resources, Purdue University, West Lafayette, IN, USA.
19	⁹ Research Center of Forest Management Engineering of State Forestry and Grassland
20	Administration, Beijing Forestry University, Beijing, China.
21	¹⁰ Department of Environmental Systems Science, ETH Zürich, Zürich, Switzerland.
22	¹¹ Department of Biology, Indiana University, 1001 E Third St, Bloomington, IN 47403, USA.
23	
24	Corresponding author: Dr. Renato K. Braghiere (renato.k.braghiere@jpl.nasa.gov)

- 25 † Current address: Jet Propulsion Laboratory, M/S 233-305F, 4800 Oak Grove Drive, Pasadena,
- 26 CA, 91109 USA.

27 Key Points:

37

nutrient cycling

28	• Global plant demand for N has increased 25% from 1850 to 2010, while the C cost
29	associated with it has increased 60% in the same period.
30	• NPP has increased by 20% from 1850 to 2010, but the NPP fraction used for nitrogen
31	acquisition increased from $\sim 1/4$ to $\sim 1/3$.
32	• Areas of savannas and forest-grasslands transition zones present a higher risk of nitrogen
33	limitation to plant growth.
34	
35	Keywords:
36	• Biogeochemistry, carbon cycling, climate change, Earth System modeling, mycorrhizae,

38 Abstract

39 Most tree species predominantly associate with a single type of mycorrhizal fungi, which can 40 differentially affect plant nutrient acquisition and biogeochemical cycling. Here, we address for 41 the first time the impact of mycorrhizal distributions on global carbon and nutrient cycling. 42 Using the state-of-the-art carbon-nitrogen economics within the Community Land Model version 43 5 (CLM5) we found Net Primary Productivity (NPP) increased throughout the 21st century by 44 20%; however, as soil nitrogen has progressively become limiting, the costs to NPP for nitrogen 45 acquisition — i.e., to mycorrhizae — have increased at a faster rate by 60%. This suggests that 46 nutrient acquisition will increasingly demand a higher portion of assimilated carbon to support 47 the same productivity. Uncertainties in mycorrhizal distributions are non-trivial, however, with 48 uncertainties in NPP by up to 345 Tg C yr⁻¹, depending on which published distribution is used. 49 Remote sensing capabilities for mycorrhizal detection show promise for refining these estimates 50 further.

51

52 Plain Language Summary

53 The majority of plants often join forces with specific types of fungi to improve their nutrient 54 acquisition capacity, which ultimately impact global photosynthesis. This is the first study to 55 explore the impacts of different types of fungi-root distributions on global carbon and nutrient 56 cycling. Using the land component of a state-of-the-art Earth System model we found that global 57 net carbon uptake increased throughout the 21st century by 20%, while the carbon spent on 58 nitrogen acquisition has increased at a faster rate by 60%. This study suggests that nutrient 59 acquisition by plants will increasingly demand a larger portion of net carbon to support the same 60 photosynthesis.

61

62

64 **1 Introduction**

65 Terrestrial ecosystems have been a persistent post-industrial carbon sink, absorbing almost a third of anthropogenic carbon emissions (Ciais et al., 2013; Schimel et al., 2015; 66 67 Friedlingstein et al., 2019). Studies suggest that terrestrial ecosystem productivity has increased 68 due to elevated CO₂ concentration (Keenan et al., 2016; Zhu et al., 2016; Chen et al., 2019), but 69 it remains unclear whether this will translate to increases in the terrestrial carbon sink in the 70 future (Friedlingstein et al., 2006, 2014; Zhang et al., 2019). It is widely expected that limiting 71 factors such as water (Trenberth et al., 2014; Kolus et al., 2019) and nutrients availability 72 (Zaehle et al., 2010; Fleischer et al., 2019; Terrer et al., 2019; Wieder et al., 2015, 2019) might 73 mediate the responses of terrestrial ecosystems to climate change. Disentangling these 74 mechanisms and exploring the consequences of atmospheric CO₂ increase requires assessment of 75 such mechanisms through Earth System models (ESMs), which allow comprehensive and 76 spatially explicit assessment of the impacts of future climate on biogeochemical cycles in 77 terrestrial ecosystems.

78 It has been estimated that a large part of plant nitrogen and phosphorus is provided by 79 fungal root symbionts (van der Heijden et al., 2015), thus it is likely that mycorrhizal 80 associations explain a large fraction of the variance in plant response to elevated CO₂ (Drake et 81 al., 2011; Orwin et al., 2011; Kivlin et al., 2013; Sulman et al., 2017; Terrer et al., 2016, 2018). 82 However, the global spatial distributions of these mechanisms as well as their potential impacts 83 are still uncertain (Norby et al., 2017; Sulman et al., 2019). Only a handful of ESMs consider 84 mycorrhizal nutrient acquisition when calculating carbon assimilation and allocation (Wang et 85 al., 2010; Zaehle et al., 2015; Goll et al., 2017). The Community Land Model version 5 (CLM5) 86 within the Community Earth System Model (CESM) currently enables an explicit representation 87 of the functional differences between different types of plant symbiotic associations (Fisher et 88 al., 2010; Brzostek et al., 2014; Shi et al., 2016; Fisher et al., 2019; Lawrence et al., 2019). 89 However, until recently, one of the major challenges in generating global estimates of nutrient 90 limitation on the global carbon cycle is related to a lack of understanding of the spatial 91 distribution of nutrient-acquiring plant-microbe symbioses. Despite the availability of regional 92 maps of present and past plant symbiotic status (Menzel et al., 2016; Swaty et al., 2016; 93 Brundrett, 2017; Jo et al., 2019), scientists have only recently begun to develop explicit global 94 data about mycorrhizal and nitrogen fixing associations (Davies-Barnard et al., 2020).

95 Recently, scientists developed methods for extrapolating spatially sparse measurements 96 into large-scale, spatially explicit maps suitable for applications within ESMs (Shi et al., 2016; 97 Soudzilovskaia et al., 2019; Steidinger et al., 2019; Sulman et al., 2019). These developments for 98 the first time enable examining how mycorrhizal distributions are related to the global carbon 99 and nitrogen cycles. In this study, we seek a better understanding of mycorrhizas on global 100 carbon and nitrogen cycles through incorporating multiple state-of-the-art spatial distributions of 101 mycorrhizal associations in a global ecosystem model. We first compare four existing global data 102 products of global spatial distributions of mycorrhizal associations. Second, we perform transient 103 global runs of CLM5 with increasing CO₂ concentration through the 20th and 21st centuries in 104 order to understand the impact of the CO₂ fertilization effect combined with different spatially 105 variable mycorrhizal representations. Finally, we evaluate the possible feedback effects that 106 changes in spatial mycorrhizal association due to climate change (Steidinger et al., 2019) may 107 have on the global carbon cycle.

108 2 Materials and Methods

109 2.1 Land Surface Model description: CLM5

110 CLM5 includes the Fixation and Uptake of Nitrogen (FUN) module calculating the 111 carbon costs for each pathway of plant nitrogen uptake - symbiotic nitrogen fixation, direct and 112 mycorrhizal uptake of soil nitrogen, and nitrogen retranslocation from leaves (Fisher et al., 2010; 113 Brzostek et al., 2014; Shi et al., 2016; Allen et al., 2020). Plants shift uptake pathways to 114 minimize the carbon costs of nitrogen uptake. FUN simulates uptake from the two major types of 115 fungi that plants associate with: arbuscular mycorrhizal (AM) and ectomycorrhizal (ECM) fungi. 116 Explicit representation of mycorrhizal associations improved the dynamic predictions of the 117 nitrogen retranslocated from leaves and taken up from the soil in previous ecosystem-scale 118 studies (Brzostek et al., 2014).

In order to generate the trade-offs between AM, ECM, and non-mycorrhizal root uptake, FUN within CLM5 uses an estimate of the percentage of aboveground biomass per grid cell that associates with each mycorrhizal type (Brzostek et al., 2014; Shi et al., 2016).

122 2.2 Coupling mycorrhizae spatial distribution into CLM5

123 Plant Functional Types (PFTs) are used to classify plants according to their physical, 124 phylogenetic, and phenological characteristics. The value of each parameter is determined or 125 inferred from observable characteristics. A spatial data product can be added as a 2D variable 126 varying as function of latitude and longitude, but because land surface models also work with the 127 concept of PFTs, adding a third dimension (i.e., latitude, longitude, and PFT) into the spatial 128 distribution can improve accuracy of processes and reduce model uncertainty (Braghiere et al., 129 2019). Here, given new datasets of spatial distributions of mycorrhizal associations based on 130 observations at different spatial resolutions, we modified CLM5 and added mycorrhizal 131 association types per PFT within a gridcell (latitude and longitude) to also consider landscape 132 heterogeneity within a model grid cell.

Four global maps of mycorrhizal association based on different assumptions and spatial resolutions were used to provide the percentage of ECM association (relative to AM) data for CLM5: Map A (Shi et al., 2016); Map B (Sulman et al., 2019), Map C (Steidinger et al., 2019), and Map D (Soudzilovskaia et al., 2019) (see **Fig. 1** and **Supplementary information** for details).

138 2.3 Simulation protocols

139 First, for each ECM map, initial ecosystem carbon and nitrogen stocks for 1850 were 140 generated using a spin-up approach using 1850 concentrations of CO₂ (284.7 ppm) and the 141 model's standard climate forcing dataset from the Global Soil Wetness Project Phase 3 version 1 142 (GSWP3v1) (Kim, 2017) at 1.9°x2.5° spatial resolution. The Model for Scale Adaptive River 143 Transport (MOSART) was turned on and ice evolution on land was turned off. Model runs were 144 performed with biogeochemistry mode on without crops for 200 years in 'accelerated 145 decomposition' mode (see Lawrence et al. (2019) for details) by cycling through the 1901–1920 146 climate forcing dataset and then for 400 years in regular spin-up mode until soil and plant carbon 147 and nitrogen stocks achieved steady state. Historical simulation was performed from 1850 to 148 2010 using transient GSWP3 climate, nitrogen deposition, and variable atmospheric CO₂ 149 concentration.

Second, in order to illustrate the model sensitivity to changes in global spatial patterns of plant symbiosis due to climate change, we used a projected map of plant symbiotic status for 2070 using a relative concentration pathway (RCP) of 8.5 W.m⁻² from Steidinger et al. (2019) versus the original map with present climate (Steidinger et al., 2019). We performed future runs (2015-2070) with the biogeochemistry mode on following the Shared Socio-Economic Pathway (SSP) number 5 (Kriegler et al., 2017). SSP5 scenarios are the only ones resulting in a radiative forcing pathway as high as the highest RCP8.5 used by Steidinger et al. (2019).

The SSP5 scenario includes extreme levels of fossil fuel use, up to a doubling of global food demand, and up to a tripling of energy demand and greenhouse gas emissions over the course of the century, marking the upper end of the scenario literature in several dimensions. We used future climatological forcing from the CESM2 simulation for the CMIP6 (Lawrence et al., 2016; O'Neill et al., 2016). We used the LMWG diagnostics package from NCAR (http://github.com/NCAR/CESM_postprocessing) and Python scripts to evaluate the differences

- 163 between each model run with CLM5.
- 164 2.4 Calculating nitrogen limitation

165 The risk of nitrogen limitation (NL) can be determined by evaluating if the growth rate 166 of NPP used for nitrogen uptake with time is larger than the growth rate of total NPP with time. 167 If the amount of NPP used for nitrogen uptake increases at a higher rate than the total NPP for a 168 particular grid cell, that grid cell is considered to be at risk of spending too much carbon on 169 nitrogen acquisition, and therefore, NL is closer to 1. On the contrary, if the amount of NPP used 170 for nitrogen uptake increases at a lower rate than the total NPP for a particular grid cell, that area 171 is not considered to be at risk of spending too much carbon on nitrogen acquisition. NL is 172 calculate as:

173

$$NL = 1. - \frac{\alpha_1(i,j)}{\alpha_2(i,j)}$$
(1.0)

174 where α_1 is the slope of the linear regression of NPP used for Nitrogen uptake per gridcell 175 (NPP_NUPTAKE(i,j)) with time and α_2 is the slope of the linear regression of NPP (NPP(i,j))

176 plus NPP_NUPTAKE(i,j) with time. Areas in red indicate higher risk of nitrogen limitation on

177 NPP based on the period from 1850 to 2010.

178 **3 Results and Discussion**

179 3.1 Different estimates of plant symbiotic status and impacts on nitrogen uptake180 pathways

181 To better visualize the differences from maps presented in Fig.1, the standard deviation 182 of the averaged difference between ECM fraction (%) of each one of the new maps and the 183 default CLM5 map is shown in Fig. 1e. All three data products agree that the default map in CLM5 overestimates ECM fraction in the boreal regions, as well as drier areas of the world, such 184 185 as the Atacama, Namibian, Somalian, Mongolian, Sonoran, and Australian deserts. Map C 186 resembles the default CLM5 map A, indicating an alignment of the assumptions that climate 187 variables are the main drivers of global biogeography of forest-tree symbioses and the 188 proposition that fixed values of mycorrhizal associations can be prescribed following PFTs 189 spatial distributions. The three maps disagree in the eastern USA, where map B indicates map A 190 overestimates ECM fraction, map C indicates the opposite, and map D shows small differences. 191 Over eastern Asia, the maps also disagree in the sign of changes of ECM fraction with respect to 192 map A. Map B shows no particular differences in Northeast China, map C indicates that map A 193 underestimates ECM fraction, while map D indicates the opposite. In central Europe, map C 194 strongly (+40%) revises the default CLM5 ECM fraction upwards, while maps B and D show a 195 much smaller positive difference in comparison to map A, except for parts of the Alps and parts 196 of the Iberic peninsula. Given that the map A is based on PFT values, the biases in particular 197 PFTs are presented in **Supplementary Fig. S1**.

Although all four maps agree in approximately 60% of the world area, some areas present large standard deviation values (> 30%), e.g., northern North America, throughout northern and eastern Asia, as well as parts of the tropical forests, i.e., northwest Amazon, the central part of the Congo Basin, and parts of the maritime continent. These areas would benefit from more field measurements of mycorrhizal association and further analysis.

Throughout all runs, the ECM-associated (NECM) and AM-associated (NAM) vegetation nitrogen uptake fluxes were the most impacted biogeochemical variables when including spatially explicit mycorrhizal status in CLM5, though the other nitrogen uptake pathways and their associated carbon costs were also impacted. There are four different representations of nitrogen acquisition pathways within CLM5: mycorrhizal uptake (NMYC), nitrogen fixation

208 (NFIX), nitrogen retranslocation from leaves (NRETRANS), and the non-mycorrhizal or direct

209 nitrogen uptake (NNONMYC). The sum of all different nitrogen acquisition pathways is the total

210 acquired nitrogen (TOTALN). **Table S2** shows the average carbon cost per unit of nitrogen

211 (gN.kgC⁻¹) in the period 2000-2010 for each different nitrogen uptake pathway as predicted by

212 CLM5.

On average for the period 2000-2010, the updated carbon cost per unit of nitrogen according to the three observation based maps (B,C, and D) increases 2.2%. The main areas where carbon costs of nitrogen uptake became higher are: (i) eastern North America, Europe, southeast Asia, and the tropics for mycorrhizal uptake; tropical and boreal forests for nitrogen fixation; and the tropics for nitrogen retranslocation (see **Supplementary material**). Changes in carbon costs of nitrogen acquisition via mycorrhizae uptake are 4.1% higher globally.

219 3.2 The effect of climate change and CO₂ fertilization on nitrogen limitation

220 To determine the climate change effect of nitrogen limitation on plant growth, Fig. 2 221 shows the global total NPP (PgC.yr⁻¹), global total carbon cost of nitrogen uptake 222 (NPP_NUPTAKE, PgC.yr⁻¹), global plant nitrogen demand (PLANT_NDEMAND, TgN.yr⁻¹), 223 and the global total nitrogen uptake (NUPTAKE, TgN.yr⁻¹). Nitrogen demand is calculated as the 224 total nitrogen that would be required if all assimilated carbon was allocated according to 225 idealized stoichiometric ratios. The CO₂ fertilization effect, with nitrogen deposition, and climate 226 change increased photosynthetic rates across the globe, represented by an increase in NPP from 227 40 PgCyr⁻¹ in 1850 to 47.5 PgCyr⁻¹ in 2010, an increase of about 20%. In turn, to support 228 elevated productivity, plants require more nitrogen, leading to an increase in plant nitrogen 229 demand from ~1600 TgN.yr⁻¹ in 1850 to 2000 TgN.yr⁻¹ in 2010, an increase of about 25%.

230 Although the rates of nitrogen uptake systematically increase in response to a higher 231 nitrogen demand, i.e., NUPTAKE of 800 TgN.yr⁻¹ in 1850 to 1000 TgN.yr⁻¹ in 2010, the 232 associated carbon cost of nitrogen acquisition increased at a faster rate, growing roughly 60% 233 more expensive in 2010 (17.5 PgCyr⁻¹) than it was in 1850 (11.2 PgCyr⁻¹). In terms of the 234 percentage of NPP spent in nitrogen acquisition, the values increased from about ~27.5% of NPP 235 in 1850 to ~32.5% of NPP in 2010. By 2075, it is projected that the NPP used for nitrogen 236 acquisition will reach 35% of total NPP (~22.5 PgCyr⁻¹), suggesting ecosystems will have much 237 less carbon available for allocation and plant growth, possibly becoming more susceptible to

extreme events that require extra carbon for re-growth, such as droughts, fires, and insectoutbreaks.

240 All transient runs from 1850 to 2010 with the new maps indicated a stronger effect of 241 climate and CO₂ fertilization on nitrogen limitation compared to map A. These findings highlight 242 that as estimated by CLM5, not only has plant demand for nitrogen increased at a faster rate than 243 actual nitrogen uptake, but that the carbon costs associated with nitrogen acquisition have 244 increased at a faster rate than the extra carbon gained through the CO₂ fertilization effect, i.e., 245 plants need to invest more carbon per unit of nitrogen uptaken. This pattern is projected to 246 continue in the future, which means that it is unlikely current plant growth rates will be sustained 247 globally.

Fig. 3a shows the risk of nitrogen limitation (NL) calculated as described in Eq. 1. According to the transient runs from 1850 to 2010 using the default CLM5 map A, tropical forests have a medium to low risk of being further limited by nitrogen, which is in agreement to some studies indicating that intact ancient tropical forests tend to accumulate and recycle large quantities of nitrogen relative to temperate forests (Hedin et al., 2009).

A part of South America, Africa, and Australia, associated with savannas and forestgrassland transition zones present a higher risk of nitrogen limitation to plant growth. Parts of the temperate forests in North America, Europe, and Asia, as well as northern areas of the planet in the presence of boreal forests present a medium to high risk of nitrogen limitation.

257

3.3 The feedback impacts of mycorrhizal changes due to climate change

Recent evidence suggests that anthropogenic influences, primarily nitrogen deposition and fire suppression, as well as climate change have increased AM tree dominance during the past three decades in the eastern United States (Jo et al., 2019). Globally, Steidinger et al. (2019) presented a study using the same environment-mycorrhizae relationships for current climate to project potential changes in the symbiotic status of forests in the future, suggesting that projected climate for 2070 reduces the abundance of ECM trees by as much as 10%, with major changes in ECM abundance along the boreal–temperate transition zone (**Fig. 3b**).

Although the magnitude of the time lag between climate change and ecosystem responses is unknown, the predicted decline in ECM trees aligns with previous simulated warming

experiments, which have demonstrated that some important ECM hosts decline at the boreal–
temperate zones under future climate conditions (Reich et al., 2015), and that ECM fungi
demonstrated increased responses of mycorrhizal fungal biomass under eCO₂ compared to AM
fungi (Dong et al., 2018), as the simulated response in the tropics (Fig. 3b).

Although it has been previously reported that climate change should impact forest
symbiosis, no study has ever evaluated the potential feedback of climate change effects on
mycorrhizal distribution onto nitrogen and carbon cycles. The difference in NPP for the period of
2016-2075 between the simulations using the future maps of ECM fraction and the simulations
using the present-day map C (Steidinger et al., 2019) are shown in Fig. 3c.

276 Large parts of South America, especially areas associated with savannas, present the 277 largest negative feedback effects on NPP due to future climate change impacts on mycorrhizal 278 association, followed by areas with boreal forests. The impact over tropical forests and areas in 279 China seem to benefit from a change in plant symbiotic status in the future. Although, these 280 results should be interpreted carefully due to the limitation of the original forest plot training data 281 in those areas of the globe used in Steidinger et al. (2019), machine learning algorithms indicate 282 more ECM fungi in the tropics in the future, possibly due to the effect eCO_2 on the tropical 283 climate.

284 In the SSP5-RCP8.5 runs from 2016 to 2075 with present-day plant symbiotic status, the 285 growth rate of nitrogen uptake was 4.8 TgN.yr⁻². In terms of carbon costs, NPP is projected to 286 increase at a rate of 265.5 TgC.yr⁻², while the carbon cost of nitrogen acquisition is projected to 287 increase at a rate of 130.4 TgC.yr⁻², an extra 135.2 TgC.yr⁻¹. The feedback effect of climate 288 change on the spatial distribution of plant symbiotic status decreases NPP globally (from 58.3 289 PgC.yr⁻¹ to 58.2 PgC.yr⁻¹), a negative impact of -23.1 TgC.yr⁻¹. The projected NPP increase rate with the future plant symbiotic status map is 266.2 TgC.yr⁻², 0.7 TgC.yr⁻² faster than the 290 291 projected NPP without changes in mycorrhizae associations. However, the carbon cost of nitrogen acquisition is projected to increase at a rate of 129.1 TgC.yr⁻², versus 130.0 TgC.yr⁻² in 292 293 the simulations without changes in the spatial distribution of plant symbiotic status. In terms of 294 total NPP globally, these changes are predicted to increase carbon costs of nitrogen acquisition 295 by 582.5 TgC.yr⁻¹, which significantly amplifies the effect of nutrient limitation on plant growth 296 worldwide.

297 4 Conclusions

To overcome the lack of global spatial representations of mycorrhizal associations, a few studies (Soudzilovskaia et al., 2019; Steidinger et al., 2019; Sulman et al., 2019) have combined a comprehensive quantitative evaluation of mycorrhizae distribution across biomes and continents, and assembled high-resolution digital maps of the global distribution of biomass fractions of different types of mycorrhizae associations.

In our analysis, we show that differences between data products have impacts upon the nitrogen and carbon cycles in CLM5. Nonetheless, this comparison did not aim to determine which map is the most realistic. Rather, we assessed the impact of different mycorrhizal representations in CLM5 to determine signs of changes in the global nitrogen and carbon cycles. In this study, we found a negative impact on future NPP due to feedback effects of climate change and CO₂ fertilization on mycorrhizae spatial distribution.

309 Although the transient runs with different spatial representations of plant symbiotic status 310 do not agree in terms of total values of nitrogen acquisition through different uptake pathways, or 311 their relative carbon costs, all experiments using the observation based maps do agree that the 312 increasing rate of plant nitrogen demand is higher than the rate of nitrogen uptake as previously 313 reported. Moreover, our simulations found that the carbon costs of nitrogen acquisition also 314 increase at a higher rate than NPP itself, indicating that plants need to invest more carbon per 315 unit of nitrogen uptake to sustain growth at current rates globally. To our knowledge, this is the 316 first study using observation-derived global maps of mycorrhizal association within an ESM to 317 estimate the impacts of climate change on mycorrhizas and its feedback on the global carbon and 318 nitrogen cycles.

319 Author Contributions

320 Conceptualization: R. K. Braghiere, J. B. Fisher, R. P. Phillips. Formal analysis: R. K.

321 Braghiere, J. B. Fisher, R. A. Fisher. Funding acquisition: J. B. Fisher, X. Yang, J. Liang, K. G.

- 322 Peay, T. W. Crowther, R. P. Phillips. Investigation: R. K. Braghiere, J. B. Fisher.
- 323 Methodology: R. K. Braghiere, J. B. Fisher, R. A. Fisher, M. Shi, B. N. Sulman, N. A.
- 324 Soudzilovskaia, X. Yang, R. P. Phillips. Supervision: J. B. Fisher, X. Yang. Validation: M. Shi,
- 325 B. S. Steidinger, B. N. Sulman, N. A. Soudzilovskaia, J. Liang, K. G. Peay, T. W. Crowther.

- 326 Writing original draft: R. K. Braghiere, J. B. Fisher. Writing review & editing: R. K.
- 327 Braghiere, J. B. Fisher, R. A. Fisher, M. Shi, B. S. Steidinger, B. N. Sulman, N. A.
- 328 Soudzilovskaia, X. Yang, J. Liang, K. G. Peay, T. W. Crowther, R. P. Phillips.

329 Acknowledgments

- 330 This research was carried out at the Jet Propulsion Laboratory, California Institute of
- 331 Technology, under a contract with the National Aeronautics and Space Administration.
- 332 California Institute of Technology. Government sponsorship acknowledged. This material is
- based upon work supported by the U.S. Department of Energy, Office of Science, Office of
- Biological and Environmental Research, Terrestrial Ecosystem Science program under Award
- Numbers DE-SC0008317 and DE-SC0016188. Funding was also provided by the NASA IDS
- 336 program. Copyright 2021. All rights reserved. We would like to acknowledge high-performance
- 337 computing support from Cheyenne (NCAR, 2020) provided by NCAR's Computational and
- 338 Information Systems Laboratory, sponsored by the National Science Foundation.

339 Data and code availability

- 340 A patch file with the modified version of CLM5 and all python scripts used for analyses and
- 341 plots are available in <u>https://doi.org/10.6084/m9.figshare.12919385.v1</u>.

342 **References**

- 343 Allen, K., Fisher, J. B., Phillips, R. P., Powers, J. S., & Brzostek, E. R. (2020). Modeling the
- 344 Carbon Cost of Plant Nitrogen and Phosphorus Uptake Across Temperate and Tropical
- 345 Forests. Frontiers in Forests and Global Change, 3.
- 346 https://doi.org/10.3389/ffgc.2020.00043
- 347 Braghiere, R. K., Quaife, T., Black, E., He, L., & Chen, J. M. (2019). Underestimation of Global
- 348 Photosynthesis in Earth System Models Due to Representation of Vegetation Structure.
- 349 *Global Biogeochemical Cycles*, *33*(11), 1358–1369. https://doi.org/10.1029/2018GB006135
- Brundrett, M. C. (2017). Distribution and Evolution of Mycorrhizal Types and Other Specialised
 Roots in Australia (pp. 361–394). https://doi.org/10.1007/978-3-319-56363-3_17
- 352 Brzostek, E. R., Fisher, J. B., & Phillips, R. P. (2014). Modeling the carbon cost of plant nitrogen

- 353 acquisition: Mycorrhizal trade-offs and multipath resistance uptake improve predictions of
- retranslocation. *Journal of Geophysical Research: Biogeosciences*, *119*(8), 1684–1697.
- 355 https://doi.org/10.1002/2014JG002660
- Chen, C., Park, T., Wang, X., Piao, S., Xu, B., Chaturvedi, R. K., et al. (2019). China and India
 lead in greening of the world through land-use management. *Nature Sustainability*, 2(2),
 122–129. https://doi.org/10.1038/s41893-019-0220-7
- Ciais, P., Sabine, C., Bala, G., Bopp, L., Brovkin, V., Canadell, J., et al. (2013). Carbon and
- 360 Other Biogeochemical Cycles. In T. F. Stocker, D. Qin, G.-K. Plattner, M. Tignor, S. K.
- 361 Allen, J. Boschung, et al. (Eds.), *Climate Change 2013 The Physical Science Basis* (pp.
- 362 465–570). Cambridge, United Kingdom and New York, NY, USA: Cambridge University
 363 Press.
- Davies-Barnard, T., Meyerholt, J., Zaehle, S., Friedlingstein, P., Brovkin, V., Fan, Y., et al.
 (2020). Nitrogen cycling in CMIP6 land surface models: progress and limitations. *Biogeosciences*, *17*, 5129–5148. https://doi.org/10.5194/bg-17-5129-2020
- Dong, Y., Wang, Z., Sun, H., Yang, W., & Xu, H. (2018). The Response Patterns of Arbuscular
 Mycorrhizal and Ectomycorrhizal Symbionts Under Elevated CO2: A Meta-Analysis.
- 369 *Frontiers in Microbiology*, 9. https://doi.org/10.3389/fmicb.2018.01248
- Drake, J. E., Gallet-Budynek, A., Hofmockel, K. S., Bernhardt, E. S., Billings, S. A., Jackson, R.
 B., et al. (2011). Increases in the flux of carbon belowground stimulate nitrogen uptake and
 sustain the long-term enhancement of forest productivity under elevated CO2. *Ecology Letters*, *14*(4), 349–357. https://doi.org/10.1111/j.1461-0248.2011.01593.x
- 374 Fisher, J. B., Sitch, S., Malhi, Y., Fisher, R. A., Huntingford, C., & Tan, S.-Y. (2010). Carbon
- 375 cost of plant nitrogen acquisition: A mechanistic, globally applicable model of plant
- 376 nitrogen uptake, retranslocation, and fixation. *Global Biogeochemical Cycles*, 24(1), n/a-
- 377 n/a. https://doi.org/10.1029/2009GB003621
- Fisher, R. A., Wieder, W. R., Sanderson, B. M., Koven, C. D., Oleson, K. W., Xu, C., et al.
 (2019). Parametric Controls on Vegetation Responses to Biogeochemical Forcing in the

- 380 CLM5. Journal of Advances in Modeling Earth Systems, 11(9), 2879–2895.
 381 https://doi.org/10.1029/2019MS001609
- Fleischer, K., Rammig, A., De Kauwe, M. G., Walker, A. P., Domingues, T. F., Fuchslueger, L.,
 et al. (2019). Amazon forest response to CO2 fertilization dependent on plant phosphorus
 acquisition. *Nature Geoscience*, *12*(9), 736–741. https://doi.org/10.1038/s41561-019-0404-
- 385

9

- Friedlingstein, P., Cox, P., Betts, R., Bopp, L., von Bloh, W., Brovkin, V., et al. (2006). Climate carbon cycle feedback analysis: Results from the C4MIP model intercomparison. *Journal of Climate*. https://doi.org/10.1175/JCLI3800.1
- Friedlingstein, P., Meinshausen, M., Arora, V. K., Jones, C. D., Anav, A., Liddicoat, S. K., &
 Knutti, R. (2014). Uncertainties in CMIP5 Climate Projections due to Carbon Cycle
- 391 Feedbacks. Journal of Climate, 27(2), 511–526. https://doi.org/10.1175/JCLI-D-12-00579.1
- Friedlingstein, P., Jones, M. W., O'Sullivan, M., Andrew, R. M., Hauck, J., Peters, G.
 P., et al. (2019). Global Carbon Budget 2019. *Earth System Science Data*, *11*(4), 1783–
 1838. https://doi.org/10.5194/essd-11-1783-2019

Goll, D. S., Winkler, A. J., Raddatz, T., Dong, N., Prentice, I. C., Ciais, P., & Brovkin, V.

396 (2017). Carbon–nitrogen interactions in idealized simulations with JSBACH (version 3.10).

- 397 *Geoscientific Model Development*, 10(5), 2009–2030. https://doi.org/10.5194/gmd-10-2009 398 2017
- Hedin, L. O., Brookshire, E. N. J., Menge, D. N. L., & Barron, A. R. (2009). The Nitrogen
 Paradox in Tropical Forest Ecosystems. *Annual Review of Ecology, Evolution, and*
- 401 *Systematics*, 40(1), 613–635. https://doi.org/10.1146/annurev.ecolsys.37.091305.110246
- 402 van der Heijden, M. G. A., Martin, F. M., Selosse, M.-A., & Sanders, I. R. (2015). Mycorrhizal
 403 ecology and evolution: the past, the present, and the future. *New Phytologist*, 205(4), 1406–
 404 1423. https://doi.org/10.1111/nph.13288
- Jo, I., Fei, S., Oswalt, C. M., Domke, G. M., & Phillips, R. P. (2019). Shifts in dominant tree
 mycorrhizal associations in response to anthropogenic impacts. *Science Advances*, 5(4),

407 eaav6358. https://doi.org/10.1126/sciadv.aav6358

- 408 Keenan, T. F., Prentice, I. C., Canadell, J. G., Williams, C., Wang, H., Raupach, M. R., &
- 409 Collatz, G. J. (2016). Recent pause in the growth rate of atmospheric CO2 due to enhanced
- 410 terrestrial carbon uptake. *Nature Communications*. https://doi.org/10.1038/ncomms13428
- 411 Kennedy, D., Swenson, S., Oleson, K. W., Lawrence, D. M., Fisher, R., Lola da Costa, A. C., &
- 412 Gentine, P. (2019). Implementing Plant Hydraulics in the Community Land Model, Version
- 413 5. Journal of Advances in Modeling Earth Systems, 11(2), 485–513.
- 414 https://doi.org/10.1029/2018MS001500
- 415 Kim, H. (2017). Global Soil Wetness Project Phase 3 Atmospheric Boundary Conditions
- 416 (Experiment 1). Data Integration and Analysis System (DIAS).
- 417 https://doi.org/https://doi.org/10.20783/DIAS.501
- Kivlin, S. N., Emery, S. M., & Rudgers, J. A. (2013). Fungal symbionts alter plant responses to
 global change. *American Journal of Botany*, *100*(7), 1445–1457.
 https://doi.org/10.3732/ajb.1200558
- 421 Kolus, H. R., Huntzinger, D. N., Schwalm, C. R., Fisher, J. B., McKay, N., Fang, Y., et al.
- 422 (2019). Land carbon models underestimate the severity and duration of drought's impact on
 423 plant productivity. *Scientific Reports*, 9(1), 2758. https://doi.org/10.1038/s41598-019424 39373-1
- 425 Kriegler, E., Bauer, N., Popp, A., Humpenöder, F., Leimbach, M., Strefler, J., et al. (2017).
- 426 Fossil-fueled development (SSP5): An energy and resource intensive scenario for the 21st
- 427 century. *Global Environmental Change*, 42, 297–315.
- 428 https://doi.org/10.1016/j.gloenvcha.2016.05.015
- 429 Lawrence, D. M., Hurtt, G. C., Arneth, A., Brovkin, V., Calvin, K. V., Jones, A. D., et al. (2016).
- 430 The Land Use Model Intercomparison Project (LUMIP) contribution to CMIP6: rationale
- 431 and experimental design. *Geoscientific Model Development*, 9(9), 2973–2998.
- 432 https://doi.org/10.5194/gmd-9-2973-2016
- 433 Lawrence, D. M., Fisher, R. A., Koven, C. D., Oleson, K. W., Swenson, S. C., Bonan, G., et al.

434	(2019). The Community Land Model Version 5: Description of New Features,
435	Benchmarking, and Impact of Forcing Uncertainty. Journal of Advances in Modeling Earth
436	Systems, 11(12), 4245-4287. https://doi.org/10.1029/2018MS001583
437	McGroddy, M. E., Daufresne, T., & Hedin, L. O. (2004). Scaling of C:N:P stoichiometry in
438	forests worldwide: implications of terrestrial redfield-type ratios. Ecology, 85(9), 2390-
439	2401. https://doi.org/10.1890/03-0351
440	Menzel, A., Hempel, S., Manceur, A. M., Götzenberger, L., Moora, M., Rillig, M. C., et al.
441	(2016). Distribution patterns of arbuscular mycorrhizal and non-mycorrhizal plant species
442	in Germany. Perspectives in Plant Ecology, Evolution and Systematics, 21, 78–88.
443	https://doi.org/10.1016/j.ppees.2016.06.002
444	NCAR. (2019). CLM5 Documentation Release, 337.
445	NCAR. (2020). CHEYENNE. https://doi.org/10.5065/D6RX99HX
446	Norby, R. J., De Kauwe, M. G., Walker, A. P., Werner, C., Zaehle, S., & Zak, D. R. (2017).
447	Comment on "Mycorrhizal association as a primary control of the CO 2 fertilization effect."
448	Science, 355(6323), 358.2-358. https://doi.org/10.1126/science.aai7976
449	O'Neill, B. C., Tebaldi, C., Van Vuuren, D. P., Eyring, V., Friedlingstein, P., Hurtt, G., et al.
450	(2016). The Scenario Model Intercomparison Project (ScenarioMIP) for CMIP6.
451	Geoscientific Model Development, 9(9), 3461-3482. https://doi.org/10.5194/gmd-9-3461-
452	2016
453	Orwin, K. H., Kirschbaum, M. U. F., St John, M. G., & Dickie, I. A. (2011). Organic nutrient
454	uptake by mycorrhizal fungi enhances ecosystem carbon storage: a model-based
455	assessment. Ecology Letters, 14(5), 493-502. https://doi.org/10.1111/j.1461-
456	0248.2011.01611.x

457 Reich, P. B., & Oleksyn, J. (2004). Global patterns of plant leaf N and P in relation to
458 temperature and latitude. *Proceedings of the National Academy of Sciences*, 101(30),
459 11001–11006. https://doi.org/10.1073/pnas.0403588101

460	Reich, Peter B., Sendall, K. M., Rice, K., Rich, R. L., Stefanski, A., Hobbie, S. E., &
461	Montgomery, R. A. (2015). Geographic range predicts photosynthetic and growth response
462	to warming in co-occurring tree species. Nature Climate Change, 5(2), 148–152.
463	https://doi.org/10.1038/nclimate2497
464	Riahi, K., van Vuuren, D. P., Kriegler, E., Edmonds, J., O'Neill, B. C., Fujimori, S., et al.
465	(2017). The Shared Socioeconomic Pathways and their energy, land use, and greenhouse
466	gas emissions implications: An overview. Global Environmental Change, 42, 153-168.
467	https://doi.org/10.1016/j.gloenvcha.2016.05.009
468	Schimel, D., Stephens, B. B., & Fisher, J. B. (2015). Effect of increasing CO 2 on the terrestrial
469	carbon cycle. Proceedings of the National Academy of Sciences, 112(2), 436-441.
470	https://doi.org/10.1073/pnas.1407302112
471	Shi, M., Fisher, J. B., Brzostek, E. R., & Phillips, R. P. (2016). Carbon cost of plant nitrogen
472	acquisition: global carbon cycle impact from an improved plant nitrogen cycle in the
473	Community Land Model. Global Change Biology, 22(3), 1299–1314.
474	https://doi.org/10.1111/gcb.13131
475	Smith, M. R., & Myers, S. S. (2018). Impact of anthropogenic CO2 emissions on global human
476	nutrition. Nature Climate Change, 8(9), 834-839. https://doi.org/10.1038/s41558-018-0253-
477	3
478	Soudzilovskaia, N. A., van Bodegom, P. M., Terrer, C., Zelfde, M. van't, McCallum, I., Luke
479	McCormack, M., et al. (2019). Global mycorrhizal plant distribution linked to terrestrial
480	carbon stocks. Nature Communications, 10(1), 5077. https://doi.org/10.1038/s41467-019-
481	13019-2
482	Steidinger, B. S., Crowther, T. W., Liang, J., Van Nuland, M. E., Werner, G. D. A., Reich, P. B.,
483	et al. (2019). Climatic controls of decomposition drive the global biogeography of forest-
484	tree symbioses. Nature, 569(7756), 404-408. https://doi.org/10.1038/s41586-019-1128-0
485	Sulman, B. N., Brzostek, E. R., Medici, C., Shevliakova, E., Menge, D. N. L., & Phillips, R. P.

486 (2017). Feedbacks between plant N demand and rhizosphere priming depend on type of

- 487 mycorrhizal association. *Ecology Letters*, 20(8), 1043–1053.
- 488 https://doi.org/10.1111/ele.12802
- 489 Sulman, B. N., Shevliakova, E., Brzostek, E. R., Kivlin, S. N., Malyshev, S., Menge, D. N. L., &
- 490 Zhang, X. (2019). Diverse Mycorrhizal Associations Enhance Terrestrial C Storage in a
- 491 Global Model. *Global Biogeochemical Cycles*, *33*(4), 501–523.
- 492 https://doi.org/10.1029/2018GB005973
- Swaty, R., Michael, H. M., Deckert, R., & Gehring, C. A. (2016). Mapping the potential
 mycorrhizal associations of the conterminous United States of America. *Fungal Ecology*,
 24, 139–147. https://doi.org/10.1016/j.funeco.2016.05.005
- 496 Terrer, C., Vicca, S., Hungate, B. A., Phillips, R. P., & Prentice, I. C. (2016). Mycorrhizal
- 497 association as a primary control of the CO 2 fertilization effect. *Science*, *353*(6294), 72–74.
 498 https://doi.org/10.1126/science.aaf4610
- 499 Terrer, C., Vicca, S., Stocker, B. D., Hungate, B. A., Phillips, R. P., Reich, P. B., et al. (2018).
- Ecosystem responses to elevated CO 2 governed by plant-soil interactions and the cost of
 nitrogen acquisition. *New Phytologist*, 217(2), 507–522. https://doi.org/10.1111/nph.14872
- 502 Terrer, C., Jackson, R. B., Prentice, I. C., Keenan, T. F., Kaiser, C., Vicca, S., et al. (2019).
- Nitrogen and phosphorus constrain the CO2 fertilization of global plant biomass. *Nature Climate Change*, 9(9), 684–689. https://doi.org/10.1038/s41558-019-0545-2
- 505 Trenberth, K. E., Dai, A., van der Schrier, G., Jones, P. D., Barichivich, J., Briffa, K. R., &
- Sheffield, J. (2014). Global warming and changes in drought. *Nature Climate Change*, 4(1),
 17–22. https://doi.org/10.1038/nclimate2067
- 508 Wang, Y. P., Law, R. M., & Pak, B. (2010). A global model of carbon, nitrogen and phosphorus
- 509 cycles for the terrestrial biosphere. *Biogeosciences*, 7(7), 2261–2282.
- 510 https://doi.org/10.5194/bg-7-2261-2010
- 511 Wieder, W. R., Cleveland, C. C., Smith, W. K., & Todd-Brown, K. (2015). Future productivity
- and carbon storage limited by terrestrial nutrient availability. *Nature Geoscience*, 8(6), 441–
- 513 444. https://doi.org/10.1038/ngeo2413

514	Wieder, W. R., Lawrence, D. M., Fisher, R. A., Bonan, G. B., Cheng, S. J., Goodale, C. L., et al.
515	(2019). Beyond static benchmarking: Using experimental manipulations to evaluate land
516	model assumptions. Global Biogeochemical Cycles. https://doi.org/10.1029/2018GB006141
517	Zaehle, S., Friedlingstein, P., & Friend, A. D. (2010). Terrestrial nitrogen feedbacks may
518	accelerate future climate change. Geophysical Research Letters, 37(1), n/a-n/a.
519	https://doi.org/10.1029/2009GL041345
520	Zaehle, S., Jones, C. D., Houlton, B., Lamarque, JF., & Robertson, E. (2015). Nitrogen
521	Availability Reduces CMIP5 Projections of Twenty-First-Century Land Carbon Uptake.
522	Journal of Climate, 28(6), 2494-2511. https://doi.org/10.1175/JCLI-D-13-00776.1
523	Zhang, Y., Song, C., Band, L. E., & Sun, G. (2019). No Proportional Increase of Terrestrial
524	Gross Carbon Sequestration From the Greening Earth. Journal of Geophysical Research:
525	Biogeosciences, 2018JG004917. https://doi.org/10.1029/2018JG004917
526	Zhu, Z., Piao, S., Myneni, R. B., Huang, M., Zeng, Z., Canadell, J. G., et al. (2016). Greening of
527	the Earth and its drivers. <i>Nature Climate Change</i> . https://doi.org/10.1038/NCLIMATE3004
528	
529	
530	
531	
532	
002	
533	
534	
535	
536	

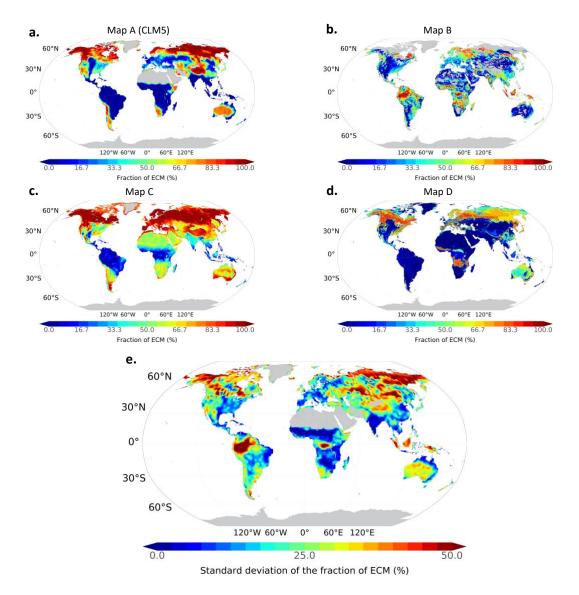




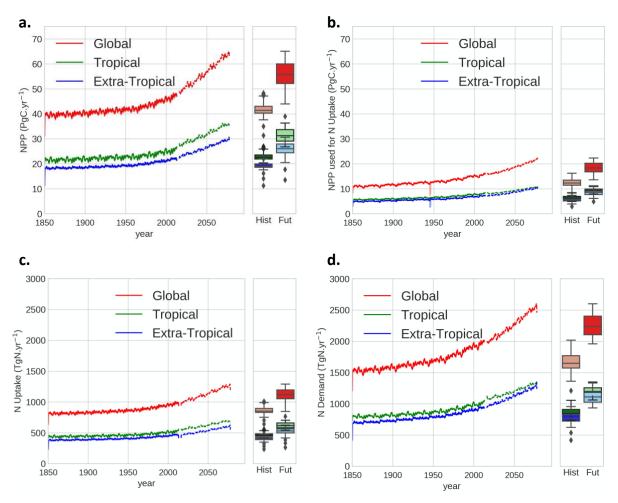
Figure 1. Global spatial distributions of ECM fraction (%). The remaining fraction is assumed to

be AM. **a.** Map A (Shi et al., 2016) (look-up table x PFTs in 1.9°x2.5°); **b.** Map B (Sulman et al.,

540 2019) (0.17°x0.17°); **c.** Map C (Steidinger et al., 2019) (1.0°x1.0° unmasked); and **d.** Map D

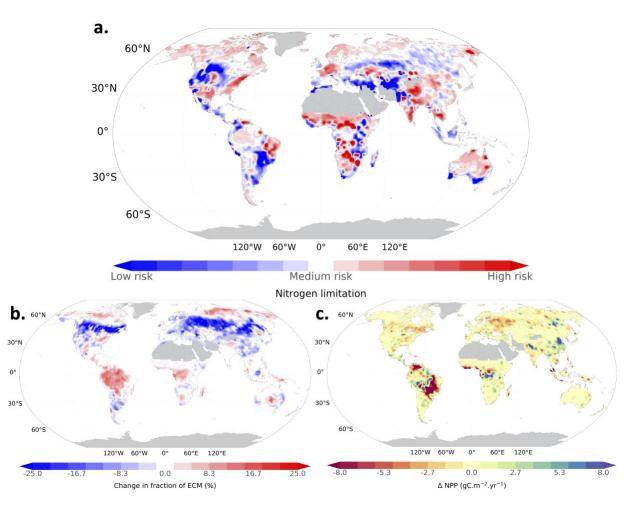
541 (Soudzilovskaia et al., 2019) $(0.17^{\circ}x0.17^{\circ})$; and **e.** standard deviation of all the four maps of ECM

542 fraction.



544

Figure 2. Trend in Net Primary Productivity and usage for nitrogen acquisition **a.** Global total NPP (PgC.yr⁻¹); **b.** global total carbon cost of nitrogen uptake (NPP_NUPTAKE, PgC.yr⁻¹); **c.** trend in nitrogen uptake and demand **a.** Global average nitrogen uptake (NUPTAKE, TgN.yr⁻¹); and **d.** global average plant nitrogen demand (PLANT_NDEMAND, TgN.yr⁻¹) for the transient historical run from 1850 to 2010 (continuous) and for the future projection SSP5 with RCP8.5 run from 2015 to 2070 (dashed) with CLM5. Tropical stands for the area of the globe between 23.5°S and 23.5°N. Extra-Tropical is the remaining area of the globe (90°S-23.5°S and 23.5°N-90°N).



555 Figure 3. a. Risk of nitrogen limitation. Areas in red indicate higher risk of nitrogen limitation 556 on NPP, and areas in blue indicate lower risk of nitrogen limitation on NPP; and projected 557 differences in NPP and mycorrhizae driven by climate change; **b.** The impact of climate change 558 on ECM fraction (%) derived from Steidinger et al. (2019) for 2070 following the RCP8.5 with 559 CMIP5 simulations; c. Difference in NPP (gC.m⁻².yr⁻¹) for future simulations (2016-2075) 560 between projected future map generated for the year of 2070 and the present-day map C 561 (Steidinger et al., 2019). The projected runs with CLM5 followed the SSP5 scenario in combination with RCP8.5 climate forcing from CESM, member of CMIP6 simulations. 562

1	Mycorrhizal distributions impact global patterns of carbon and nutrient cycling
2	R. K. Braghiere ^{1,2†} , J. B. Fisher ^{1,2} , R. A. Fisher ^{3,4} , M. Shi ^{1,2} , B. S. Steidinger ⁵ , B. N.
3	Sulman ⁶ , N. A. Soudzilovskaia ⁷ , X. Yang ⁶ , J. Liang ^{8,9} , K. G. Peay ⁵ , T. W. Crowther ¹⁰ , R. P.
4	Phillips ¹¹
5	¹ Jet Propulsion Laboratory, California Institute of Technology, 4800 Oak Grove Drive,
6	Pasadena, CA, 91109 USA.
7	² Joint Institute for Regional Earth System Science and Engineering, University of California at
8	Los Angeles, Los Angeles, CA, 90095 USA.
9	³ Climate and Global Dynamics Division, National Center for Atmospheric Research, Boulder,
10	CO, USA.
11	⁴ Laboratoire Évolution & Diversité Biologique, CNRS:UMR 5174, Université Paul Sabatier,

- 12 Toulouse, France.
- ⁵ Department of Biology, Stanford University, Stanford, CA, USA.
- ⁶Environmental Sciences Division and Climate Change Science Institute, Oak Ridge National
- 15 Laboratory, Oak Ridge, TN, USA.
- ¹⁶ ⁷ Environmental Biology Department, Institute of Environmental Sciences, Leiden University,
- 17 Leiden, The Netherlands.
- ¹⁸ ⁸ Department of Forestry and Natural Resources, Purdue University, West Lafayette, IN, USA.
- ¹⁹ ⁹Research Center of Forest Management Engineering of State Forestry and Grassland
- 20 Administration, Beijing Forestry University, Beijing, China.
- ²¹ ¹⁰ Department of Environmental Systems Science, ETH Zürich, Zürich, Switzerland.
- ¹¹ Department of Biology, Indiana University, 1001 E Third St, Bloomington, IN 47403, USA.

- 24 Corresponding author: Dr. Renato K. Braghiere (<u>renato.k.braghiere@jpl.nasa.gov</u>)
- 25 † Current address: Jet Propulsion Laboratory, M/S 233-305F, 4800 Oak Grove Drive, Pasadena,
- 26 CA, 91109 USA.

27 Supplementary Materials Document

28 Includes:

29	- Supplementary Information
30	- Supplementary Tables
31	- Supplementary Figures
32	- Supplementary References
22	

33

34 Supplementary information

35 Land Surface model description: the Community Land Model version 5 (CLM5)

CLM5 is the land surface component of the Community Earth System Model 2 (CESM2; 36 37 https://www.cesm.ucar.edu/models/cesm2/). CLM5 includes three important changes to the representation of plant carbon and nitrogen dynamics: i) the Leaf Utilization of Nitrogen for 38 39 Assimilation (LUNA) module allows plants to adjust their partitioning of nitrogen among the maximum rate of carboxylation (V_{cmax}), the maximum rate of electron transport (J_{max}), and other 40 leaf nitrogen components, to achieve co-limitation of photosynthesis under the prevailing time-41 averaged environmental drivers (CO₂, temperature, humidity, soil moisture, radiation, and day 42 length) (Xu et al., 2012; Ali et al., 2016; Fisher et al., 2019); ii) the 'FlexCN' module allows 43 plants to alter and optimize their stoichiometry, removing the down-regulation of gross primary 44 productivity (GPP) that was used in CLM4 and CLM4.5 (Cheng et al., 2019; Ghimire et al., 45 2016). In the new allocation algorithm, the total nitrogen supply in each timestep is partitioned 46 among tissues in proportion to their relative 'demand' terms. Additional details on how 47 stoichiometry is optimized can be found in Lawrence et al. (2019) and Fisher et al. (2019); and 48 49 finally, iii) the Fixation and Uptake of Nitrogen (FUN) module implements a 'carbon cost' for each source of plant nitrogen uptake (Fisher et al., 2010; Brzostek et al., 2014; Shi et al., 2016; 50 51 Allen et al., 2020).

52 The carbon cost of nitrogen uptake from soil by mycorrhizal or non-mycorrhizal

pathways, for each soil layer *j*, is controlled by two uptake parameters that pertain respectively to the relationship between soil nitrogen and nitrogen uptake, and between fine root carbon density and nitrogen uptake. For mycorrhizal or non-mycorrhizal nitrogen uptake, the cost functions are given as:

$$N_{cost,pathway,j} = \frac{k_{n,pathway}}{N_{smin,j}} + \frac{k_{c,pathway}}{c_{root,j}}$$
(1.0)

where $k_{n,\text{pathway}}$ (kgC.m⁻²) and $k_{c,\text{pathway}}$ (kgC.m⁻²) varies according to whether the pathway considered is referring to a non-mycorrhizal (direct), ECM, or AM uptake. N_{smin,j} and c_{root,j} are the soil nitrogen content (gN.m⁻³) and fine root carbon density (gC.m⁻³), respectively. Please refer to CLM5 technical note and related publications (Fisher et al., 2019; Lawrence et al., 2019; NCAR, 2019) for the complete set of equations.

Shi et al. (2016) classified the Plant Functional Types (PFTs) in CLM, based upon known 63 64 associations between plant species and either arbuscular mycorrhizae (AM) or ectomycorrhizae (ECM) fungi described in the literature (Read, 1991; Allen et al., 1995; Phillips et al., 2013). 65 While some PFTs are usually AM-dominated (e.g., grasslands), others are usually ECM-66 dominated (e.g., boreal forest). PFT symbiont fraction estimates are available as ratios of the 67 AM-associated and ECM-associated plants of the CLM PFTs as a table in Shi et al. (2016). 68 These numbers are usually binary, associating one PFT with a single type of mycorrhizae, e.g., 69 70 0% or 100%, except for broadleaf deciduous temperate trees, which associates 50% with AM and 50% with ECM. 71

72 Coupling mycorrhizae spatial distribution into CLM5

In CLM5, within each grid cell, the soil area available for vegetation is divided into 73 patches that correspond to the area fraction of that PFT. For each PFT, a number of key 74 parameters are defined, such as the target tissue C:N values, stomatal water use efficiency, 75 76 maximum hydraulic conductivity and sensitivity to embolism (Kennedy et al., 2019), tissue allocation fractions (for leaves, fine roots, stem, and coarse roots), tissue turnover times, and the 77 78 rate at which litter class (labile, lignin, cellulose) decays and returns nutrients to the soil after death. Four global maps of mycorrhizal association based on different assumptions and spatial 79 80 resolutions were added into CLM5 to provide the percentage of ECM association (relative to

AM) data for CLM5: Map A (Shi et al., 2016); Map B (Sulman et al., 2019), Map C (Steidinger
et al., 2019), and Map D (Soudzilovskaia et al., 2019) (Fig. 1).

Map B was derived from Sulman et al. (2019), who assembled empirical AM data points presenting species number of AM fungi obtained from the MAARJAM database (Öpik et al., 2010), and ECM data points presenting species number of ECM fungi obtained from Tedersoo et al. (2014). These data were used to define niche models which were used to develop spatial maps of the relative probability of AM and ECM fungal presence within areal units of 10 arcmin.

These niche models were used to estimate ECM fraction by comparing the relative probability ofAM and ECM presence:

90

$$%ECM = 100*p(ECM)/(p(ECM) + p(AM))$$
 (2.0)

where p(ECM) and p(AM) are the probabilities of ECM or AM presence, respectively, from the
niche model in each grid cell.

93 Map C was derived from Steidinger et al. (2019), who proposed a global map of the symbiotic status of forests, using a database of over 1 million forest inventory plots containing 94 more than 28,000 tree species, and 70 global predictor layers: 19 climatic indices (relating to 95 annual, monthly, and quarterly temperature and precipitation variables), 14 soil chemical indices 96 (relating to soil nitrogen density, microbial nitrogen, C:N ratios and soil P fractions, pH and 97 cation exchange capacity), 26 vegetative indices (relating to leaf area index, total stem density, 98 99 enhanced vegetation index means and variances), and 5 topographic variables (relating to elevation and hillshade). Their maps provide quantitative estimates of the distribution of 100 101 above ground biomass fractions among AM, ECM, and N fixers plants within areal units of 0.5° and 1.0°. 102

Map D was proposed by Soudzilovskaia et al. (2019), who assembled a global database 103 on plant mycorrhizal type associations that included 2,169 studies and 27,736 species-by-site 104 records for 12,702 plant species and combined it with information about dominant plant species 105 and their growth form across distinct combinations of Bailey's with 98 ecoregions (Bailey, 2014) 106 and European Space Agency (ESA) land cover categories (ESA, 2017) with spatial resolution of 107 300 m. Their maps provide quantitative estimates of the distribution of aboveground biomass 108 fractions among AM, ECM, and ericoid mycorrhiza (ERM) plants withins areal units of 10 109 110 arcmin.

The maps D and B are principally different from maps A and C. Consequently, 111 conversions to unify the data for comparisons have to be applied. Map D shows fractions of 112 biomass for all plants, not only trees, while the map B shows the likelihood of occurrence of 113 ECM biomass in a grid cell based on a species distribution model fit to a genomic database. 114 Sulman et al. (2019) produced a range from very low likelihood of ECM fungal DNA being 115 present in observations to higher likelihood of ECM presence. In order to compare map B with 116 117 other maps, the ECM map was first combined with the AM map and normalized, producing a 118 spectrum that incorporates both mycorrhizal types.

119 A regridding process of the maps to CLM5 grid scales was applied by calculating an average value for ECM in percentage per PFT per gridcell based on the GLC2000 land cover 120 121 data (Bartholomé & Belward, 2005) at a spatial resolution of 500 m following a look-up table (Supplementary Table S1). The average value of ECM percentage was assigned to one of the 122 16 particular natural vegetation PFTs in CLM5 per gridcell, assuming that AM and ECM trees 123 do not differ in biomass. In this case, using basal area maps and biomass percentages map 124 125 interchangeably is acceptable in tree-dominated areas. In other areas, it is assumed that although differences in the data products might exist, the nature of the measure is assumed to have little 126 127 impact, as long as given in the format of a ratio of ECM over ECM plus AM present in the grid cells, due to the fact that CLM5 ingests the data as a ECM ratio per PFT. 128

- 129
- 130
- 131
- 132

133 **Table S1.** Look-up table between GLC Global Class and CLM PFTs.

CLM PFT	Classification	GLC Global Class
PFT 0	Bare soil (not vegetated)	(19)Bare Areas
PFT 1	Needleleaf evergreen temperate tree	(04)Tree Cover, needle-leaved, evergreen; (06)Tree Cover, mixed leaf type; (07)Tree Cover, regularly flooded, fresh water (& brackish); (08)Tree Cover, regularly flooded, saline water; (09)Mosaic; (10)Tree Cover, burnt; (17)Mosaic;

	1	
	Needleleaf evergreen boreal tree	(04) Tree Cover, needle-leaved, evergreen; (06) Tree Cover,
PFT 2		mixed leaf type; (07)Tree Cover, regularly flooded, fresh
		water (& brackish); (08) Tree Cover, regularly flooded, saline
		water; (09)Mosaic; (10)Tree Cover, burnt; (17)Mosaic;
	Needleleaf	(05)Tree Cover, needle-leaved, deciduous; (06)Tree Cover,
PFT 3	deciduous boreal	mixed leaf type; (07) Tree Cover, regularly flooded, fresh
1115		water (& brackish); (08)Tree Cover, regularly flooded, saline
	tree	water; (09)Mosaic; (10)Tree Cover, burnt; (17)Mosaic;
	Broadleaf	(01) Tree Cover, broadleaved, evergreen; (06) Tree Cover,
PFT 4		mixed leaf type; (07)Tree Cover, regularly flooded, fresh
ΓΓΙ 4	evergreen	water (& brackish); (08) Tree Cover, regularly flooded, saline
	tropical tree	water; (09)Mosaic; (10)Tree Cover, burnt; (17)Mosaic;
	Broadleaf evergreen temperate tree	(01) Tree Cover, broadleaved, evergreen; (06) Tree Cover,
PFT 5		mixed leaf type; (07)Tree Cover, regularly flooded, fresh
FFIJ		water (& brackish); (08) Tree Cover, regularly flooded, saline
		water; (09)Mosaic; (10)Tree Cover, burnt; (17)Mosaic;
		(02)Tree Cover, broadleaved, deciduous, closed; (06)Tree
	Broadleaf	Cover, mixed leaf type; (07)Tree Cover, regularly flooded,
PFT 6	deciduous	fresh water (& brackish); (08) Tree Cover, regularly flooded,
	tropical tree	saline water; (09)Mosaic; (10)Tree Cover, burnt; (17
)Mosaic;
		(02) Tree Cover, broadleaved, deciduous, closed; (06) Tree
	Broadleaf	Cover, mixed leaf type; (07) Tree Cover, regularly flooded,
PFT 7	deciduous	fresh water (& brackish); (08) Tree Cover, regularly flooded,
	temperate tree	saline water; (09)Mosaic; (10)Tree Cover, burnt; (17
)Mosaic;
		(02) Tree Cover, broadleaved, deciduous, closed; (06) Tree
	Broadleaf	Cover, mixed leaf type; (07) Tree Cover, regularly flooded,
PFT 8	deciduous boreal	fresh water (& brackish); (08) Tree Cover, regularly flooded,
	tree	saline water; (09)Mosaic; (10)Tree Cover, burnt; (17
)Mosaic;
	1	I

		
	Broadleaf evergreen shrub	(01) Tree Cover, broadleaved, evergreen; (06) Tree Cover,
		mixed leaf type; (09)Mosaic; (11)Shrub Cover, closed-open,
PFT 9		evergreen; (13)Herbaceous Cover, closed-open; (14)Sparse
		Herbaceous or sparse Shrub Cover; (15)Regularly flooded
		Shrub and/or Herbaceous Cover; (17)Mosaic; (18)Mosaic
		(03)Tree Cover, broadleaved, deciduous, open; (06)Tree
	Broadleaf	Cover, mixed leaf type; (09)Mosaic; (12)Shrub Cover,
PFT 10	deciduous	closed-open, deciduous; (13)Herbaceous Cover, closed-open;
PF1 10		(14)Sparse Herbaceous or sparse Shrub Cover; (15
	temperate shrub)Regularly flooded Shrub and/or Herbaceous Cover; (17
)Mosaic; (18)Mosaic
		(03)Tree Cover, broadleaved, deciduous, open; (06)Tree
	Dreadlasf	Cover, mixed leaf type; (09)Mosaic; (12)Shrub Cover,
DET 11	Broadleaf deciduous boreal shrub	closed-open, deciduous; (13)Herbaceous Cover, closed-open;
PFT 11		(14)Sparse Herbaceous or sparse Shrub Cover; (15
)Regularly flooded Shrub and/or Herbaceous Cover; (17
)Mosaic; (18)Mosaic
	C3 arctic grass	(09)Mosaic; (13)Herbaceous Cover, closed-open; (14
DET 10)Sparse Herbaceous or sparse Shrub Cover; (15)Regularly
PFT 12		flooded Shrub and/or Herbaceous Cover; (17)Mosaic; (18
)Mosaic
		(09)Mosaic; (13)Herbaceous Cover, closed-open; (14
	C3 nonarctic)Sparse Herbaceous or sparse Shrub Cover; (15)Regularly
PFT 13	grass	flooded Shrub and/or Herbaceous Cover; (17)Mosaic; (18
)Mosaic
		(09)Mosaic; (13)Herbaceous Cover, closed-open; (14
)Sparse Herbaceous or sparse Shrub Cover; (15)Regularly
PFT 14	C4 grass	flooded Shrub and/or Herbaceous Cover; (17)Mosaic; (18
)Mosaic
		(09)Mosaic; (16)Cultivated and managed areas; (17
PFT 15	Corn)Mosaic; (18)Mosaic

PFT 16	Wheat	(09)Mosaic; (16)Cultivated and managed areas; (17			
FFI IU)Mosaic; (18)Mosaic			
		(20)Water Bodies (natural & artificial); (21)Snow and Ice			
PFT 17	NaN	(natural & artificial); (22) Artificial surfaces and associated			
		areas; (23)No data			

- 135 *(09) Mosaic: Tree cover / Other natural vegetation; (17) Mosaic: Cropland / Tree Cover / Other
- 136 natural vegetation; (18) Mosaic: Cropland / Shrub or Grass Cover.
- 137
- 138 **Table S2.** Average carbon cost values per unit nitrogen (gN.kgC⁻¹) from 2000 to 2010 for each
- 139 different pathway and sum for all new maps and the default one in CLM5.

Pathway cost	Reference	TRANSIENT – 2000 – 2010				
$(gN.kgC^{-1})$	Map A (CLM5)	Map B	Map C	Map D	Average (B,C,D)	Change (%)
NMYC	1.15	1.15	1.21	1.04	1.13	1.4%
NFIX	104.00	103.80	105.20	107.60	105.53	-1.5%
NRETRANS	925.00	924.00	905.00	914.00	914.33	1.2%
NNONMYC	115.53	115.13	130.00	124.97	123.01	-6.5%
TOTALN	38.33	38.07	36.62	37.82	37.50	2.2%

140

- 142 **Table S3.** Average values from 2000 to 2010 of nitrogen uptake for each one of the different
- 143 pathways and sum for the spatially distributed PFT based.

Pathway	Reference	TRANSIENT – 2000 – 2010				
(TgNyr-1)	Map A (CLM5)	Map B	Map C	Map D		
NECM	10.7	10.8	14.8	7.5		
NAM	9.9	9.8	8.7	11.8		
NFIX	52.0	51.9	52.6	53.8		

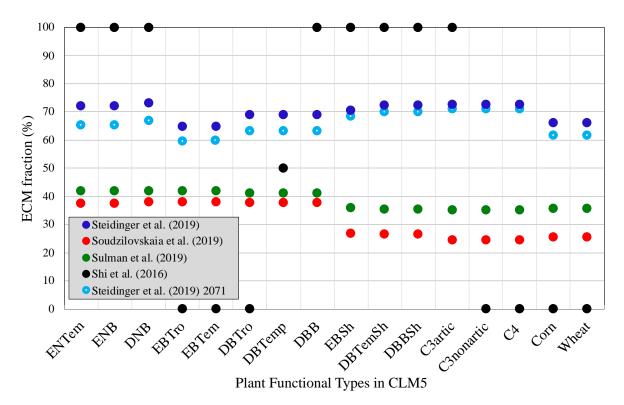
2000-2010

	NRETRANS	92.5	92.4	90.5	91.4	
	NNONMYC	808.7	805.9	793.0	799.8	
	TOTALN	973.7	970.8	959.5	964.4	
144						
145						
146						
147						
148						
149	Table S4. Average values from 2000 to 2010 of carbon costs of nitrogen uptake for each one of					
150	the different pathways and sum for the spatially distributed PFT based. The values of CLM4-					

151 FUN from Shi et al. (2016) are shown as reference.

	1995-2004	2000-2010			
Pathway	Reference	Reference	TRANSIENT - 2000 - 2010		
(PgCyr-1)	CLM4-	Map A	Map B	Map C	Map D
	FUN	(CLM5)			
NPP_MYC	1.2	17.9	17.9	19.4	18.6
NPP_NFIX	0.4	0.5	0.5	0.5	0.5
NPP_NRETRANS	0.6	0.1	0.1	0.1	0.1
NPP_TOTAL N	2.4	25.4	25.5	26.2	25.5
NPP_NPASSIVE	0.0	0.0	0.0	0.0	0.0
NPP_NDIRECT	0.2	7.0	7.0	6.1	6.4
	1				

1995-2004 2000-2010



154 **Figure S1.** PFT global average of ECM fraction in percentage for ref. (Sulman et al., 2019); ref.

155 (Steidinger et al., 2019) present and future (2071); ref. (Soudzilovskaia et al., 2019) and the base

156 map in CLM5 as in ref. (Shi et al., 2016).

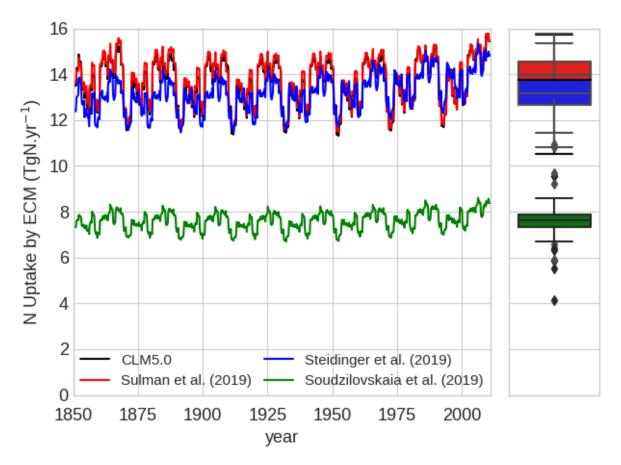
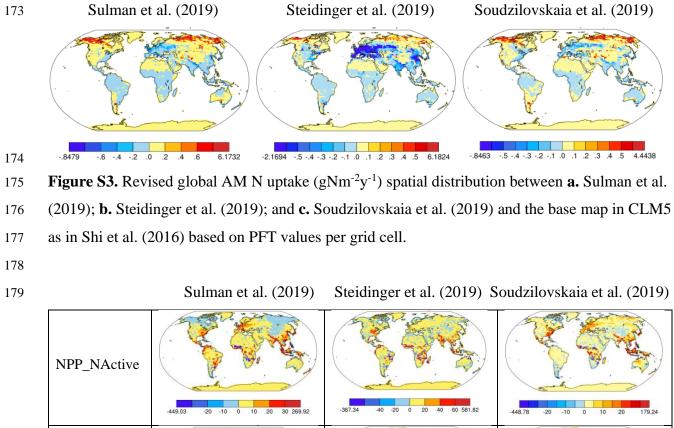


Figure S2. Nitrogen uptake through ectomycorrhizal association (NECM) in TgNyr⁻¹ for the
transient run (1850-2010) for ref. (Sulman et al., 2019); ref. (Steidinger et al., 2019); and ref.
(Soudzilovskaia et al., 2019) and the base map in CLM5 as in ref. (Shi et al., 2016) based on
fixed PFT values.



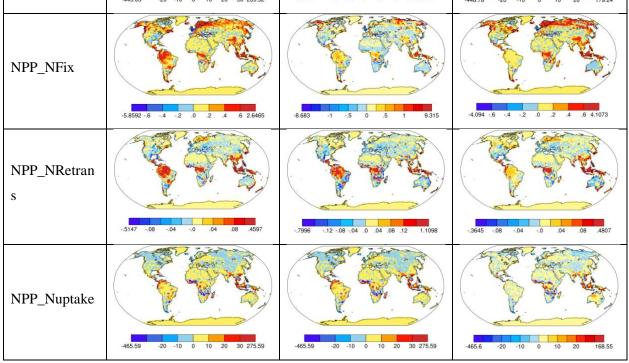


Figure S4. Revised carbon used for nitrogen uptake $(gCm^{-2}y^{-1})$ spatial distribution between a. Sulman et al. (2019); b. Steidinger et al. (2019); and c. Soudzilovskaia et al. (2019) and the base

- map in CLM as in Shi et al. (2016) based on PFT values per gridbox for different pathways:
 Mycorrhizal (NPP_NActive), Symbiotic BNF (NPP_NFix), retranslocated N (NPP_NRetrans),
- and total (NPP_Nuptake).
- 186
- 187 Sulman et al. (2019) Steidinger et al. (2019) Soudzilovskaia et al. (2019) 188 -.04 -.02 .00 .02 .04 .11771 -1.04959 .02 .04 -1.09864 -.04 .02 189 -.04 -.02 .00 -1.08009 .13618 -.02 -.00 .04 .32126

190 **Figure S5.** Revised Autotrophic Respiration (gCm⁻²y⁻¹) spatial distribution between **a.** Sulman et

al. (2019); **b.** Steidinger et al. (2019); and **c.** Soudzilovskaia et al. (2019) and the base map in

192 CLM as in Shi et al. (2016) based on fixed PFT values (**above**) and based on PFT values per

- 193 gridbox (**below**).
- 194
- 195
- 196
- 197
- 198
- 199
- 200
- 201
- 202
- 203
- 204
- 205
- 206
- 207
- 208
- 209

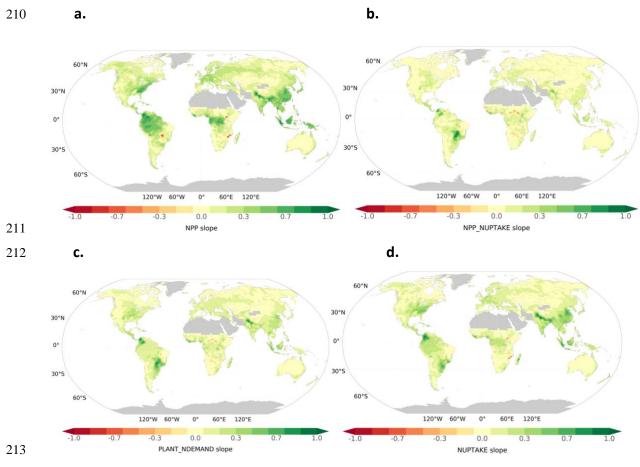


Figure S6. Normalized linear regression slope of a. NPP, b. NPP_NUPTAKE, c.

- 215 PLANT_NDEMAND, and **d.** NUPTAKE with time.
- 216
- 217

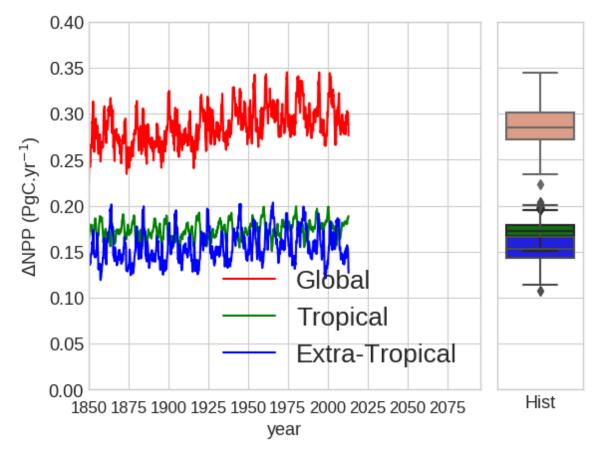
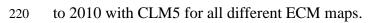


Figure S7. Global average maximum $\triangle NPP$ (PgC.yr⁻¹) for the transient historical runs from 1850



234 **References**

- Ali, A. A., Xu, C., Rogers, A., Fisher, R. A., Wullschleger, S. D., Massoud, E. C., et al. (2016).
 A global scale mechanistic model of photosynthetic capacity (LUNA V1.0). *Geoscientific Model Development*, 9(2), 587–606. https://doi.org/10.5194/gmd-9-587-2016
- Allen, E. B., Allen, M. F., Helm, D. J., Trappe, J. M., Molina, R., & Rincon, E. (1995). Patterns
- and regulation of mycorrhizal plant and fungal diversity. *Plant and Soil*, *170*(1), 47–62.
- 240 https://doi.org/10.1007/BF02183054
- Allen, K., Fisher, J. B., Phillips, R. P., Powers, J. S., & Brzostek, E. R. (2020). Modeling the
 Carbon Cost of Plant Nitrogen and Phosphorus Uptake Across Temperate and Tropical
 Forests. *Frontiers in Forests and Global Change*, *3*.
- 244 https://doi.org/10.3389/ffgc.2020.00043
- Bailey, R. G. (2014). *Ecoregions: The ecosystem geography of the oceans and continents*.
- *Ecoregions: The Ecosystem Geography of the Oceans and Continents*. Springer New York.
 https://doi.org/10.1007/978-1-4939-0524-9
- Bartholomé, E., & Belward, A. S. (2005). GLC2000: a new approach to global land cover
 mapping from Earth observation data. *International Journal of Remote Sensing*.
 https://doi.org/10.1080/01431160412331291297
- Braghiere, R. K., Quaife, T., Black, E., He, L., & Chen, J. M. (2019). Underestimation of Global
 Photosynthesis in Earth System Models Due to Representation of Vegetation Structure.
- 253 *Global Biogeochemical Cycles*, *33*(11), 1358–1369. https://doi.org/10.1029/2018GB006135
- Brzostek, E. R., Fisher, J. B., & Phillips, R. P. (2014). Modeling the carbon cost of plant nitrogen
 acquisition: Mycorrhizal trade-offs and multipath resistance uptake improve predictions of
- retranslocation. *Journal of Geophysical Research: Biogeosciences*, *119*(8), 1684–1697.
- 257 https://doi.org/10.1002/2014JG002660
- Cheng, S. J., Hess, P. G., Wieder, W. R., Thomas, R. Q., Nadelhoffer, K. J., Vira, J., et al.
- 259 (2019). Decadal fates and impacts of nitrogen additions on temperate forest carbon storage:
- a data-model comparison. *Biogeosciences*, *16*(13), 2771–2793. https://doi.org/10.5194/bg 16-2771-2019
- 262 ESA. (2017). CCI Land cover map 2015.
- Fisher, J. B., Sitch, S., Malhi, Y., Fisher, R. A., Huntingford, C., & Tan, S.-Y. (2010). Carbon
 cost of plant nitrogen acquisition: A mechanistic, globally applicable model of plant

- nitrogen uptake, retranslocation, and fixation. *Global Biogeochemical Cycles*, 24(1), n/a n/a. https://doi.org/10.1029/2009GB003621
- Fisher, R. A., Wieder, W. R., Sanderson, B. M., Koven, C. D., Oleson, K. W., Xu, C., et al.
- 268 (2019). Parametric Controls on Vegetation Responses to Biogeochemical Forcing in the
- 269 CLM5. Journal of Advances in Modeling Earth Systems, 11(9), 2879–2895.
- 270 https://doi.org/10.1029/2019MS001609
- Ghimire, B., Riley, W. J., Koven, C. D., Mu, M., & Randerson, J. T. (2016). Representing leaf
 and root physiological traits in CLM improves global carbon and nitrogen cycling
- 273 predictions. *Journal of Advances in Modeling Earth Systems*.
- 274 https://doi.org/10.1002/2015MS000538
- 275 Kennedy, D., Swenson, S., Oleson, K. W., Lawrence, D. M., Fisher, R., Lola da Costa, A. C., &
- 276 Gentine, P. (2019). Implementing Plant Hydraulics in the Community Land Model, Version
- 5. Journal of Advances in Modeling Earth Systems, 11(2), 485–513.
- 278 https://doi.org/10.1029/2018MS001500
- Kim, H. (2017). Global Soil Wetness Project Phase 3 Atmospheric Boundary Conditions
 (Experiment 1). Data Integration and Analysis System (DIAS).
- 281 https://doi.org/https://doi.org/10.20783/DIAS.501
- 282 Kriegler, E., Bauer, N., Popp, A., Humpenöder, F., Leimbach, M., Strefler, J., et al. (2017).
- Fossil-fueled development (SSP5): An energy and resource intensive scenario for the 21st
- century. *Global Environmental Change*, *42*, 297–315.
- 285 https://doi.org/10.1016/j.gloenvcha.2016.05.015
- Lawrence, D. M., Hurtt, G. C., Arneth, A., Brovkin, V., Calvin, K. V., Jones, A. D., et al. (2016).
- 287 The Land Use Model Intercomparison Project (LUMIP) contribution to CMIP6: rationale
- and experimental design. *Geoscientific Model Development*, 9(9), 2973–2998.
- 289 https://doi.org/10.5194/gmd-9-2973-2016
- 290 Lawrence, D. M., Fisher, R. A., Koven, C. D., Oleson, K. W., Swenson, S. C., Bonan, G., et al.
- 291 (2019). The Community Land Model Version 5: Description of New Features,
- 292 Benchmarking, and Impact of Forcing Uncertainty. *Journal of Advances in Modeling Earth*
- 293 Systems, 11(12), 4245–4287. https://doi.org/10.1029/2018MS001583
- NCAR. (2019). CLM5 Documentation Release, 337.
- 295 O'Neill, B. C., Tebaldi, C., Van Vuuren, D. P., Eyring, V., Friedlingstein, P., Hurtt, G., et al.

- 296 (2016). The Scenario Model Intercomparison Project (ScenarioMIP) for CMIP6.
- 297 *Geoscientific Model Development*, 9(9), 3461–3482. https://doi.org/10.5194/gmd-9-3461-
- 298 2016
- 299 Öpik, M., Vanatoa, A., Vanatoa, E., Moora, M., Davison, J., Kalwij, J. M., et al. (2010). The
- 300 online database MaarjAM reveals global and ecosystemic distribution patterns in arbuscular
- 301 mycorrhizal fungi (Glomeromycota). *New Phytologist*, *188*(1), 223–241.
- 302 https://doi.org/10.1111/j.1469-8137.2010.03334.x
- Phillips, R. P., Brzostek, E., & Midgley, M. G. (2013). The mycorrhizal-associated nutrient
 economy: a new framework for predicting carbon-nutrient couplings in temperate forests.
- 305 New Phytologist, 199(1), 41–51. https://doi.org/10.1111/nph.12221
- Read, D. J. (1991). Mycorrhizas in ecosystems. *Experientia*, 47(4), 376–391.
- 307 https://doi.org/10.1007/BF01972080
- Reich, P. B., Sendall, K. M., Rice, K., Rich, R. L., Stefanski, A., Hobbie, S. E., & Montgomery,
- R. A. (2015). Geographic range predicts photosynthetic and growth response to warming in
 co-occurring tree species. *Nature Climate Change*, 5(2), 148–152.
- 311 https://doi.org/10.1038/nclimate2497
- Riahi, K., van Vuuren, D. P., Kriegler, E., Edmonds, J., O'Neill, B. C., Fujimori, S., et al.
- 313 (2017). The Shared Socioeconomic Pathways and their energy, land use, and greenhouse
- 314 gas emissions implications: An overview. *Global Environmental Change*, *42*, 153–168.

315 https://doi.org/10.1016/j.gloenvcha.2016.05.009

- Shi, M., Fisher, J. B., Brzostek, E. R., & Phillips, R. P. (2016). Carbon cost of plant nitrogen
 acquisition: global carbon cycle impact from an improved plant nitrogen cycle in the
- Community Land Model. *Global Change Biology*, 22(3), 1299–1314.
- 319 https://doi.org/10.1111/gcb.13131
- 320 Soudzilovskaia, N. A., van Bodegom, P. M., Terrer, C., Zelfde, M. van't, McCallum, I., Luke
- 321 McCormack, M., et al. (2019). Global mycorrhizal plant distribution linked to terrestrial
- 322 carbon stocks. *Nature Communications*, *10*(1), 5077. https://doi.org/10.1038/s41467-019 323 13019-2
- 324 Steidinger, B. S., Crowther, T. W., Liang, J., Van Nuland, M. E., Werner, G. D. A., Reich, P. B.,
- et al. (2019). Climatic controls of decomposition drive the global biogeography of forest-
- tree symbioses. *Nature*, *569*(7756), 404–408. https://doi.org/10.1038/s41586-019-1128-0

- 327 Sulman, B. N., Shevliakova, E., Brzostek, E. R., Kivlin, S. N., Malyshev, S., Menge, D. N. L., &
- Zhang, X. (2019). Diverse Mycorrhizal Associations Enhance Terrestrial C Storage in a
- 329 Global Model. *Global Biogeochemical Cycles*, *33*(4), 501–523.
- 330 https://doi.org/10.1029/2018GB005973
- Tedersoo, L., Bahram, M., Põlme, S., Kõljalg, U., Yorou, N. S., Wijesundera, R., et al. (2014).
- Global diversity and geography of soil fungi. *Science*, *346*(6213), 1256688.
- 333 https://doi.org/10.1126/science.1256688
- Xu, C., Fisher, R., Wullschleger, S. D., Wilson, C. J., Cai, M., & McDowell, N. G. (2012).
- Toward a mechanistic modeling of nitrogen limitation on vegetation dynamics. *PLoS ONE*,
- 336 7(5), 1–11. https://doi.org/10.1371/journal.pone.0037914
- 337